

Package ‘betapart’

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Components

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Suggests vegan

Description Functions to compute pair-wise dissimilarities (distance matrices) and multiple-site dissimilarities, separating the turnover and nestedness-resultant components of taxonomic (incidence and abundance based), functional and phylogenetic beta diversity.

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NeedsCompilation no

Author Andres Baselga [aut, cre],
David Orme [aut],
Sebastien Villegier [aut],
Julien De Bortoli [aut],
Fabien Leprieur [aut],
Maxime Logez [aut],
Renato Henriques-Silva [ctb]

Maintainer Andres Baselga <andres.baselga@usc.es>

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bbsData

BBS data by state for two timeslices

Description

The data consists of binary presence/absence matrices for 569 bird species across 49 US states for two time slices (1980 - 1985 and 2000 - 2005). Only species (identified by AOU number) recorded during both time periods are included. The data are taken from the North American Breeding Bird Survey dataset and from a version of the database downloaded in May 2009.

Usage

```
data(bbsData)
```

Format

Two matrices (bbs1980 and bbs2000) of identical structure showing the presence/absence of the species as binary data.

state US states by USPS two letter codes.

aou Species identity by AOU species ID numbers.

Source

<http://www.pwrc.usgs.gov/BBS/>

Examples

```
data(bbsData)
str(bbs1980)
str(bbs2000)
```

beta.multi	<i>Multiple-site dissimilarities</i>
------------	--------------------------------------

Description

Computes 3 multiple-site dissimilarities accounting for the spatial turnover and the nestedness components of beta diversity, and the sum of both values

Usage

```
beta.multi(x, index.family="sorensen")
```

Arguments

`x` data frame, where rows are sites and columns are species. Alternatively `x` can be a `betapart` object derived from the `betapart.core` function

`index.family` family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Value

The function returns a list with the three multiple site dissimilarity values.

For `index.family="sorensen"` the three indices are:

`beta.SIM` value of the turnover component, measured as Simpson dissimilarity

`beta.SNE` value of the nestedness component, measured as nestedness-resultant fraction of Sorensen dissimilarity

`beta.SOR` value of the overall beta diversity, measured as Sorensen dissimilarity

For `index.family="jaccard"` the three indices are:

`beta.JTU` value of the turnover component, measured as turnover fraction of Jaccard dissimilarity

`beta.JNE` value of the nestedness component, measured as nestedness-resultant fraction of Jaccard dissimilarity

`beta.JAC` value of the overall beta diversity, measured as Jaccard dissimilarity

Author(s)

Andrés Baselga and David Orme

References

- Baselga, A. 2010. Partitioning the turnover and nestedness components of beta diversity. *Global Ecology and Biogeography* 19:134-143
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

See Also

[beta.pair](#), [beta.sample](#), [betapart.core](#), [beta.temp](#)

Examples

```
data(ceram.s)
ceram.beta<-beta.multi(ceram.s, index.family="sor")
```

beta.multi.abund	<i>Abundance-based multiple-site dissimilarities</i>
------------------	------------------------------------------------------

Description

Computes 3 multiple-site dissimilarities accounting for the (i) balanced variation and (ii) abundance gradient components of dissimilarity, and the sum of both values (i.e. total abundance-based dissimilarity)

Usage

```
beta.multi.abund(x, index.family="bray")
```

Arguments

x	data frame, where rows are sites and columns are species. Alternatively x can be a betapart.abund object derived from the betapart.core.abund function
index.family	family of dissimilarity indices, partial match of "bray" or "ruzicka".

Value

The function returns a list with the three multiple site dissimilarity values.

For index.family="bray" the three indices are:

beta.BRAY.BAL	value of the balanced variation component of Bray-Curtis multiple-site dissimilarity
beta.BRAY.GRA	value of the abundance-gradient component of Bray-Curtis multiple-site dissimilarity
beta.BRAY	value of the overall dissimilarity, measured as Bray-Curtis multiple-site dissimilarity

For index.family="ruzicka" the three indices are:

beta.RUZ.BAL	value of the balanced variation component of Ruzicka multiple-site dissimilarity
beta.RUZ.GRA	value of the abundance-gradient component of Ruzicka multiple-site dissimilarity
beta.RUZ	value of the overall dissimilarity, measured as Ruzicka multiple-site dissimilarity

Author(s)

Andrés Baselga

References

Baselga, A. 2017. Partitioning abundance-based multiple-site dissimilarity into components: balanced variation in abundance and abundance gradients. *Methods in Ecology and Evolution* 8: 799-808

See Also

[beta.pair.abund](#), [beta.sample.abund](#), [betapart.core.abund](#), [beta.multi](#)

Examples

```
require(vegan)
data(BCI)
beta.multi.abund(BCI, index.family="bray")
```

beta.pair

Incidence-based pair-wise dissimilarities

Description

Computes 3 distance matrices accounting for the (i) turnover (replacement), (ii) nestedness-resultant component, and (iii) total dissimilarity (i.e. the sum of both components).

Usage

```
beta.pair(x, index.family = "sorensen")
```

Arguments

x data frame, where rows are sites and columns are species. Alternatively x can be a betapart object derived from the betapart.core function

index.family family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Value

The function returns a list with three dissimilarity matrices. For `index.family="sorensen"` the three matrices are:

<code>beta.sim</code>	dist object, dissimilarity matrix accounting for spatial turnover (replacement), measured as Simpson pair-wise dissimilarity
<code>beta.sne</code>	dist object, dissimilarity matrix accounting for nestedness-resultant dissimilarity, measured as the nestedness-fraction of Sorensen pair-wise dissimilarity
<code>beta.sor</code>	dist object, dissimilarity matrix accounting for total dissimilarity, measured as Sorensen pair-wise dissimilarity (a monotonic transformation of beta diversity)

For `index.family="jaccard"` the three matrices are:

<code>beta.jtu</code>	dist dissimilarity matrix accounting for spatial turnover, measured as the turnover-fraction of Jaccard pair-wise dissimilarity
<code>beta.jne</code>	dist object, dissimilarity matrix accounting for nestedness-resultant dissimilarity, measured as the nestedness-fraction of Jaccard pair-wise dissimilarity
<code>beta.jac</code>	dist object, dissimilarity matrix accounting for beta diversity, measured as Jaccard pair-wise dissimilarity (a monotonic transformation of beta diversity)

Author(s)

Andrés Baselga and David Orme

References

- Baselga, A. 2010. Partitioning the turnover and nestedness components of beta diversity. *Global Ecology and Biogeography* 19:134-143
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

See Also

[beta.pair.abund](#), [codebeta.multi](#), [beta.sample](#), [betapart.core](#), [beta.temp](#)

Examples

```
data(ceram.s)
ceram.dist<-beta.pair(ceram.s, index.family="jac")
```

beta.pair.abund	<i>Abundance-based pair-wise dissimilarities</i>
-----------------	--------------------------------------------------

Description

Computes 3 distance matrices accounting for the (i) balanced variation in abundances, (ii) abundance gradients, and (iii) total dissimilarity (i.e. the sum of both components).

Usage

```
beta.pair.abund(x, index.family = "bray")
```

Arguments

x	data frame, where rows are sites and columns are species. Alternatively x can be a betapart.abund object derived from the betapart.core.abund function
index.family	family of dissimilarity indices, partial match of "bray" or "ruzicka".

Value

The function returns a list with three dissimilarity matrices. For index.family="bray" the three matrices are:

beta.bray.bal	dist object, dissimilarity matrix accounting for the dissimilarity derived from balanced variation in abundance between sites
beta.bray.gra	dist object, dissimilarity matrix accounting for the dissimilarity derived from unidirectional abundance gradients
beta.bray	dist object, dissimilarity matrix accounting for total abundance-based dissimilarity between sites, measured as the Bray-Curtis index

For index.family="ruzicka" the three matrices are:

beta.ruz.bal	dist object, dissimilarity matrix accounting for the dissimilarity derived from balanced variation in abundance between sites
beta.ruz.gra	dist object, dissimilarity matrix accounting for the dissimilarity derived from unidirectional abundance gradients
beta.ruz	dist object, dissimilarity matrix accounting for total abundance-based dissimilarity between sites, measured as the Ruzicka index

Author(s)

Andrés Baselga

References

Baselga, A. 2013. Separating the two components of abundance-based dissimilarity: balanced changes in abundance vs. abundance gradients. *Methods in Ecology and Evolution* 4: 552–557

Legendre, P. 2014. Interpreting the replacement and richness difference components of beta diversity. *Global Ecology and Biogeography*, 23: 1324–1334

See Also

[beta.multi.abund](#), [beta.sample.abund](#), [betapart.core.abund](#), [beta.pair](#)

Examples

```
require(vegan)
data(BCI)
BCI.pair<-beta.pair.abund(BCI, index.family="bray")
```

beta.para.control *Specifying Control Values for Internal Parallel Cluster*

Description

The values supplied in the `beta.para.control()` call replace the defaults, and a list with all settings (i.e., values for all possible arguments) is returned. The returned list is used to define the internal parallel cluster of the `functional.betapart.core` function.

Usage

```
beta.para.control(nc = floor(parallel::detectCores()/2), type = "PSOCK",
                 LB = TRUE, size = 1)
```

Arguments

<code>nc</code>	number of cores to use. Default is half of the available cores.
<code>type</code>	character - the type of cluster to be used, either "SOCK", "PSOCK" or "FORK" (not on Windows).
<code>LB</code>	logical indicating if load balancing has to be used. Default is TRUE
<code>size</code>	number of operation run on each core at each iteration. Default is 1.

Value

a list with components for each of the possible arguments.

Author(s)

Maxime Logez

Examples

```
str(beta.para.control(nc = 2, LB = FALSE))
```

beta.sample	<i>Resampling multiple-site dissimilarity for n sites</i>
-------------	-----------------------------------------------------------

Description

Resamples the 3 multiple-site dissimilarities (turnover, nestedness-resultant fraction, and overall beta diversity) for a subset of sites of the original data frame.

Usage

```
beta.sample(x, index.family="sorensen", sites=nrow(x$data), samples = 1)
```

Arguments

x	data frame, where rows are sites and columns are species. Alternatively x can be a <code>betapart</code> object derived from the <code>betapart.core</code> function.
index.family	family of dissimilarity indices, partial match of "sorensen" or "jaccard".
sites	number of sites for which multiple-site dissimilarities will be computed. If not specified, default is all sites.
samples	number of repetitions. If not specified, default is 1.

Value

The function returns a list with a dataframe with the resampled 3 multiple-site dissimilarities (turnover fraction, nestedness-resultant fraction and overall dissimilarity; see [beta.multi](#)), a vector with the respective means and a vector with the respective standard deviation.

For `index.family="sorensen"`:

sampled.values	dataframe containing beta.SIM, beta.SNE and beta.SOR for all samples
mean.values	vector containing the mean values of beta.SIM, beta.SNE and beta.SOR among samples
sd.values	vector containing the sd values of beta.SIM, beta.SNE and beta.SOR among samples

For `index.family="jaccard"`:

sampled.values	dataframe containing beta.JTU, beta.JNE and beta.JAC for all samples
mean.values	vector containing the mean values of beta.JTU, beta.JNE and beta.JAC among samples
sd.values	vector containing the sd values of beta.JTU, beta.JNE and beta.JAC among samples

Author(s)

Andrés Baselga and David Orme

References

Baselga, A. 2010. Partitioning the turnover and nestedness components of beta diversity. *Global Ecology and Biogeography* 19:134-143

Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

See Also

[beta.multi](#), [beta.sample](#), [beta.temp](#)

Examples

```
# Read the data for Northern and Southern European cerambycids
data(ceram.s)
data(ceram.n)

# Resample 100 times the multiple-site dissimilarities
# for 10 countries.
beta.ceram.s<-beta.sample(ceram.s, index.family="sor", sites=10, samples=100)
beta.ceram.n<-beta.sample(ceram.n, index.family="sor", sites=10, samples=100)

# Plot the distributions of beta.SIM in Southern Europe (red)
# and Northern Europe (blue)
plot(density(beta.ceram.s$sampled.values$beta.SIM), col="red", xlim=c(0,1))
lines(density(beta.ceram.n$sampled.values$beta.SIM), col="blue")

# Compute the p-value of difference in beta.SIM between South and North
# (i.e. the probability of finding in the North a higher value than
# in the South)
p.value.beta.SIM<-length(which(beta.ceram.s$sampled.values$beta.SIM<
beta.ceram.n$sampled.values$beta.SIM))/100

p.value.beta.SIM
# The result is 0 and we used 100 samples, so p<0.01
```

beta.sample.abund *Resampling abundance-based multiple-site dissimilarity for n sites*

Description

Resamples the 3 abundance-based multiple-site dissimilarities (balanced variation fraction, abundance-gradient fraction, and overall dissimilarity) for a subset of sites of the original data frame.

Usage

```
beta.sample.abund(x, index.family="bray", sites = nrow(x), samples = 1)
```

Arguments

x	data frame, where rows are sites and columns are species
index.family	family of dissimilarity indices, partial match of "bray" or "ruzicka".
sites	number of sites for which multiple-site dissimilarities will be computed. If not specified, default is all sites.
samples	number of repetitions. If not specified, default is 1.

Value

The function returns a list with a dataframe with the resampled 3 multiple-site dissimilarities (balanced variation fraction, abundance-gradient fraction and overall dissimilarity; see [beta.multi.abund](#)), a vector with the respective means and a vector with the respective standard deviation.

For index.family="bray":

sampled.values	dataframe containing beta.BRAY.BAL, beta.BRAY.GRA and beta.BRAY for all samples
mean.values	vector containing the mean values of beta.BRAY.BAL, beta.BRAY.GRA and beta.BRAY among samples
sd.values	vector containing the sd values of beta.BRAY.BAL, beta.BRAY.GRA and beta.BRAY among samples

For index.family="ruzicka":

sampled.values	dataframe containing beta.RUZ.BAL, beta.RUZ.GRA and beta.RUZ for all samples
mean.values	vector containing the mean values of beta.RUZ.BAL, beta.RUZ.GRA and beta.RUZ among samples
sd.values	vector containing the sd values of beta.RUZ.BAL, beta.RUZ.GRA and beta.RUZ among samples

Author(s)

Andrés Baselga

References

Baselga, A. 2017. Partitioning abundance-based multiple-site dissimilarity into components: balanced variation in abundance and abundance gradients. *Methods in Ecology and Evolution* 8: 799-808

See Also

[beta.multi.abund](#), [beta.sample](#)

Examples

```
require(vegan)
data(BCI)
beta.sample.abund(BCI, index.family="bray", sites=10, samples=100)
```

`beta.temp`*Temporal change in community composition*

Description

Computes the dissimilarity for each locality between time 1 and time 2, considering the turnover and nestedness components of temporal change, and the sum of both values (overall change)

Usage

```
beta.temp(x, y, index.family="sorensen")
```

Arguments

`x` data frame for time 1, where rows are sites and columns are species
`y` data frame for time 2, where rows are sites and columns are species. `x` and `y` must contain exactly the same sites and species
`index.family` family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Value

The function returns a data frame where rows are sites and columns are pairwise dissimilarity values between cell composition in time 1 and time 2. For `index.family="sorensen"` the indices are `beta.sim`, `beta.sne`, and `beta.sor`. For `index.family="jaccard"` the indices are `beta.jtu`, `beta.sne`, and `beta.jac`.

Author(s)

Andrés Baselga and David Orme

References

Baselga, A. 2010. Partitioning the turnover and nestedness components of beta diversity. *Global Ecology and Biogeography* 19:134-143

Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

See Also

[beta.multi](#), [beta.pair](#), [beta.sample](#), [betapart.core](#),

Examples

```
data(bbsData)
bbs.t <- beta.temp(bbs1980, bbs2000, index.family="sor")
```

`betapart`*Partitioning beta diversity into turnover and nestedness components*

Description

betapart allows computing pair-wise dissimilarities (distance matrices) and multiple-site dissimilarities, separating the turnover and nestedness-resultant components of taxonomic (incidence and abundance based), functional and phylogenetic beta diversity.

Details

The partitioning of incidence-based dissimilarity can be performed for two different families of indices: Sorensen and Jaccard. The pairwise function `beta.pair` yields 3 distance matrices accounting for the spatial turnover and the nestedness components of beta-diversity. The third distance matrix accounts for the sum of both components, i.e. total dissimilarity (a monotonic transformation of beta diversity). The multiple site function `beta.multi` yields the spatial turnover and the nestedness components of overall dissimilarity, and the sum of both components, total dissimilarity. The basic calculations for all these multiple-site measures and pairwise dissimilarity matrices can be computed using the function `betapart.core`, which returns an object of class `betapart`. This is useful for large datasets as the consuming calculations are done only once, and its result can then be used for computing many indices. The multiple-site values can be randomly sampled a specified number of times for a specified number of sites using the function `beta.sample`. The aforementioned indices used for assessing spatial patterns can also be used for measuring temporal changes in community composition with the function `beta.temp`. Likewise, an analogous framework has been implemented for separating the two components of abundance-based dissimilarity (balanced changes in abundance vs. abundance gradients) using commands `beta.pair.abund`, `beta.multi.abund`, `betapart.core.abund`, and `beta.sample.abund`. The framework has been extended for functional beta diversity with commands `functional.betapart.core`, `functional.beta.pair` and `functional.beta.multi`, and for phylogenetic beta diversity with commands `phylo.betapart.core`, `phylo.beta.pair` and `phylo.beta.multi`. The package also allows fitting negative exponential or power law distance-decay models for assessing the relationship between assemblage (dis)similarity and spatial (or other) distance. `decay.model` will fit the distance-decay function via GLM, `plot.decay` will plot the distance-decay pattern, and `boot.coefs.decay` will bootstrap the parameters of the distance-decay model.

Author(s)

Andrés Baselga, David Orme, Sébastien Villéger, Julien De Bortoli and Fabien Leprieur

References

- Baselga, A. 2010. Partitioning the turnover and nestedness components of beta diversity. *Global Ecology and Biogeography* 19:134-143
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232
- Baselga, A. 2013. Separating the two components of abundance-based dissimilarity: balanced changes in abundance vs. abundance gradients. *Methods in Ecology and Evolution*, 4: 552-557

- Baselga, A. 2017. Partitioning abundance-based multiple-site dissimilarity into components: balanced variation in abundance and abundance gradients. *Methods in Ecology and Evolution* 8: 799-808
- Baselga A, Leprieur, F. 2015. Comparing methods to separate components of beta diversity. *Methods in Ecology and Evolution* 6: 1069-1079
- Baselga A, Orme CDL. 2012. betapart: an R package for the study of beta diversity. *Methods Ecol. Evol.* 3: 808-812
- Gómez-Rodríguez, C. & Baselga, A. 2018. Variation among European beetle taxa in patterns of distance decay of similarity suggests a major role of dispersal processes. *Ecography*, in press
- Legendre P. 2014. Interpreting the replacement and richness difference components of beta diversity. *Global Ecology and Biogeography*, 23: 1324–1334
- Leprieur F, Albouy C, De Bortoli J, Cowman PF, Belwood DR, Mouillot D. 2012. Quantifying phylogenetic beta diversity: distinguishing between "true" turnover of lineages and phylogenetic diversity gradients. *PLoS One* 7(8): e42760
- Villéger, S. Grenouillet, G., Brosse, S. 2013. Decomposing functional beta-diversity reveals that low functional beta-diversity is driven by low functional turnover in European fish assemblages. *Global Ecology and Biogeography*, 22: 671-681

betapart.core

Core calculations of betapart

Description

Computes the basic quantities needed for computing the multiple-site beta diversity measures and pairwise dissimilarity matrices.

Usage

```
betapart.core(x)
```

Arguments

x data frame, where rows are sites and columns are species

Value

The function returns an object of class betapart with the following elements:

sumSi	the sum of the species richness values of all sites
St	the total richness in the dataset
a	the multiple-site analog of the shared species term
shared	a matrix containing the number of species shared between pairs of sites
not.shared	a matrix containing the number of species not shared between pairs of sites: b, c

- sum.not.shared a matrix containing the total number of species not shared between pairs of sites:
b+c
- max.not.shared a matrix containing the total maximum number of species not shared between
pairs of sites: max(b,c)
- min.not.shared a matrix containing the total minimum number of species not shared between
pairs of sites: min(b,c)

Author(s)

Andrés Baselga and David Orme

References

- Baselga, A. 2010. Partitioning the turnover and nestedness components of beta diversity. *Global Ecology and Biogeography* 19:134-143
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

See Also

[beta.multi](#), [beta.pair](#), [beta.sample](#), [beta.temp](#),

Examples

```
data(ceram.s)
ceram.core.s<-betapart.core(ceram.s)
ceram.dist.jac<-beta.pair(ceram.core.s, index.family="jac")
ceram.multi.sor<-beta.multi(ceram.core.s, index.family="sor")
```

betapart.core.abund *Core calculations of betapart for abundance-based dissimilarity measures*

Description

Computes the basic quantities needed for computing the abundance-based multiple-site dissimilarity measures and pairwise dissimilarity matrices.

Usage

```
betapart.core.abund(x)
```

Arguments

x data frame, where rows are sites and columns are species

Value

The function returns an object of class `betapart.abund` with the following elements:

`multiple.shared.abund`
the multiple-site intersection component in terms of abundances (AM)

`pair.shared.abund`
a matrix containing the agreement in abundance between pairs of sites (A)

`min.not.shared.abund`
a matrix containing the minimum disagreement in abundance between pairs of sites: $\min(B,C)$

`max.not.shared.abund`
a matrix containing the maximum disagreement in abundance between pairs of sites between pairs of sites: $\max(B,C)$

`pair.not.shared.abund`
a matrix containing the total disagreement in abundance between pairs of sites: $B+C$

Author(s)

Andrés Baselga

References

- Baselga, A. 2013. Separating the two components of abundance-based dissimilarity: balanced changes in abundance vs. abundance gradients. *Methods in Ecology and Evolution*, 4: 552–557
- Legendre, P. 2014. Interpreting the replacement and richness difference components of beta diversity. *Global Ecology and Biogeography*, 23: 1324–1334
- Baselga, A. 2017. Partitioning abundance-based multiple-site dissimilarity into components: balanced variation in abundance and abundance gradients. *Methods in Ecology and Evolution*, 8: 799–808

See Also

[beta.multi.abund](#), [beta.pair.abund](#), [beta.sample.abund](#), [betapart.core](#)

Examples

```
require(vegan)
data(BCI)
core.BCI<-betapart.core.abund(BCI)
```

boot.coefs.decay	<i>Bootstrapping the parameters of distance-decay models computed with decay.model()</i>
------------------	------------------------------------------------------------------------------------------

Description

Takes the output of `decay.model()` and bootstraps the parameters (intercept and slope).

Usage

```
boot.coefs.decay(x, R)
```

Arguments

x	the output of <code>decay.model()</code> .
R	the number of bootstrap samples.

Value

The function returns a list with:

model.type	functional form of the model, either negative exponential or power law.
y.type	similarities or dissimilarities.
boot.coefs	a matrix with the coefficients bootstrapped distributions, including intercept values in the first column, and slope values in the second column.
original.coefs	model coefficients as estimated with a GLM using <code>decay.model()</code> .
mean.boot	the mean of the bootstrapped distributions.
sd.boot	the standard deviation of the bootstrapped distributions.

Author(s)

Andrés Baselga

References

Gómez-Rodríguez, C. & Baselga, A. 2018. Variation among European beetle taxa in patterns of distance decay of similarity suggests a major role of dispersal processes. *Ecography* 41: 1825–1834

See Also

[decay.model](#)

Examples

```

require(vegan)

data(BCI)
## UTM Coordinates (in metres)
UTM.EW <- rep(seq(625754, 626654, by=100), each=5)
UTM.NS <- rep(seq(1011569, 1011969, by=100), len=50)

spat.dist<-dist(data.frame(UTM.EW, UTM.NS))

dissim.BCI<-beta.pair.abund(BCI)$beta.bray.bal

BCI.decay.pow<-decay.model(dissim.BCI, spat.dist, model.type="pow", perm=10)

boot.coefs.decay(BCI.decay.pow, 10)

```

bray.part

*Partitioning pair-wise Bray-Curtis dissimilarities***Description**

Computes 3 distance matrices accounting for the balanced variation and abundance gradient components of Bray-Curtis dissimilarity, and the sum of both values (i.e. Bray-Curtis dissimilarity)

Usage

```
bray.part(x)
```

Arguments

x data frame of species abundances, where rows are sites and columns are species.

Value

The function returns a list with three dissimilarity matrices.

bray.bal	dist object, dissimilarity matrix accounting for the dissimilarity derived from balanced variation in abundance between sites
bray.gra	dist object, dissimilarity matrix accounting for the dissimilarity derived from unidirectional abundance gradients
bray	dist object, dissimilarity matrix accounting for total abundance-based dissimilarity between sites, measured as the Bray-Curtis index

Author(s)

Andrés Baselga

References

Baselga, A. 2013. Separating the two components of abundance-based dissimilarity: balanced changes in abundance vs. abundance gradients. *Methods in Ecology and Evolution* 4: 552–557

See Also

[beta.pair](#)

Examples

```
require(vegan)
data(BCI)
BCI.matrices<-bray.part(BCI)
```

ceram.n

Cerambycidae from Northern European Countries

Description

The `ceram.n` data frame has 19 rows and 634 columns. Columns are presence/absence values of 634 species. The variable names are formed from the scientific names. The case names are standard country abbreviations, excepting RSS (Southern European Russia), RSC (Central European Russia) and RSN (Northern European Russia).

Usage

```
data(ceram.n)
```

Source

<http://www.cerambycidae.net/>

References

1. Danilevsky, M. L. 2007. A check-list of Longicorn Beetles (Coleoptera, Cerambycoidea) of Europe. Available at <http://www.cerambycidae.net/>
2. Baselga, A. 2008. Determinants of species richness, endemism and turnover in European longhorn beetles. *Ecography* 31:263-271

ceram.s

*Cerambycidae from Southern European Countries***Description**

The `ceram.s` data frame has 15 rows and 634 columns. Columns are presence/absence values of 634 species. The variable names are formed from the scientific names. The case names are standard country abbreviations, excepting SS (Serbia) and CBH (Croatia and Bosnia-Herzegovina).

Usage

```
data(ceram.s)
```

Source

<http://www.cerambycidae.net/>

References

1. Danilevsky, M. L. 2007. A check-list of Longicorn Beetles (Coleoptera, Cerambycoidea) of Europe. Available at <http://www.cerambycidae.net/>
2. Baselga, A. 2008. Determinants of species richness, endemism and turnover in European longhorn beetles. *Ecography* 31:263-271

decay.model

*Fitting distance decay models to pair-wise assemblage similarity***Description**

Fits a negative exponential or power law function (via GLM) describing (i) the decay of assemblage similarity with spatial (or any other) distance, or, equivalently, (ii) the increase of assemblage dissimilarity with distance.

Usage

```
decay.model(y, x, model.type="exponential", y.type="similarities", perm=100)
```

Arguments

<code>y</code>	codedist object, either containing similarities or dissimilarities between pairs of assemblages.
<code>x</code>	codedist object, containing distances (spatial or other) between pairs of assemblages.
<code>model.type</code>	functional form of the model, either negative exponential or power law, partial match of "exponential" or "power".

y.type	polarity of the codedist object (i.e. 1 means total similarity or total dissimilarity), partial match of "similarities" or "dissimilarities".
perm	number of randomizations to assess significance.

Value

The function returns a list with:

data	dataframe containing distances (spatial or other) and similarities (or dissimilarities).
model	the fitted GLM.
model.type	functional form of the model, either negative exponential or power law.
y.type	similarities or dissimilarities.
pseudo.r.squared	similarities or dissimilarities.
a.intercept	intercept of the model, i.e. similarity or dissimilarity at distance=0.
b.slope	slope of the model, i.e. rate at which similarity decreases with distance, or dissimilarity increases with distance in a negative exponential or power law model.
p.value	significance of the model, as estimated from a randomization test.

Author(s)

Andrés Baselga

References

Nekola, J.C. & McGill, B.J. 2014. Scale dependency in the functional form of the distance decay relationship. *Ecography*, 37, 309-320

Gómez-Rodríguez, C. & Baselga, A. 2018. Variation among European beetle taxa in patterns of distance decay of similarity suggests a major role of dispersal processes. *Ecography* 41: 1825–1834

See Also

[beta.pair](#), [beta.pair.abund](#)

Examples

```
require(vegan)

data(BCI)
## UTM Coordinates (in metres)
UTM.EW <- rep(seq(625754, 626654, by=100), each=5)
UTM.NS <- rep(seq(1011569, 1011969, by=100), len=50)

spat.dist<-dist(data.frame(UTM.EW, UTM.NS))

dissim.BCI<-beta.pair.abund(BCI)$beta.bray.bal
```

```

plot(spat.dist, dissim.BCI, ylim=c(0,1), xlim=c(0, max(spat.dist)))

BCI.decay.exp<-decay.model(dissim.BCI, spat.dist, y.type="dissim", model.type="exp", perm=10)

BCI.decay.pow<-decay.model(dissim.BCI, spat.dist, y.type="dissim", model.type="pow", perm=10)

plot.decay(BCI.decay.exp, col=rgb(0,0,0,0.5))
plot.decay(BCI.decay.exp, col="red", remove.dots=TRUE, add=TRUE)
plot.decay(BCI.decay.pow, col="blue", remove.dots=TRUE, add=TRUE)

```

functional.beta.multi *Multiple-site functional dissimilarities*

Description

Computes 3 multiple-site functional dissimilarities accounting for the spatial turnover and the nestedness components of functional beta diversity, and the sum of both values. Functional dissimilarities are based on volume of convex hulls intersections in a multidimensional functional space.

Usage

```
functional.beta.multi(x, traits, index.family="sorensen", warning.time=TRUE)
```

Arguments

x	data frame, where rows are sites and columns are species. Alternatively x can be a functional.betapart object derived from the functional.betapart.core function
traits	if x is not a functional.betapart object, a data frame, where rows are species and columns are functional space dimensions (i.e. quantitative traits or synthetic axes after PCoA). Number of species in each site must be strictly higher than number of dimensions. Number of dimensions should not exceed 4 and number of sites should not exceed 10. See Details.
index.family	family of dissimilarity indices, partial match of "sorensen" or "jaccard".
warning.time	a logical value indicating whether computation of multiple-site dissimilarities would stop if number of dimensions exceeds 4 or if number of sites exceeds 10. If turn to FALSE, computation process can be tracked in the step.fbc.txt file, see Details.

Details

For multiple-site dissimilarities metrics ($N > 2$ sites), the volume of the union of the N convex hulls is computed using the inclusion-exclusion principle (Villéger et al., 2011). It requires to compute the volume of all the intersections between 2 to N convex hulls. Intersection between $k > 2$ convex hulls is computed as the intersection between the two convex hulls shaping intersections between the corresponding $k-1$ convex hulls, e.g. $V(A_n B_n C) = V((A_n B)_n (B_n C))$. For N sites, computing

multiple-site dissimilarity metrics thus requires computing $2^N - (N+1)$ pair-wise intersections between convex hulls in a multidimensional functional space. Computation time of the intersection between two convex hulls increases with the number of dimensions (D) of the functional space. Therefore, to prevent from running very long computation process `warning.time` is set by default to stop the function if $D > 4$ or $N > 10$. Computation progress can be tracked in the "step.fbc.txt" file written in the working directory. This table shows proportion of steps completed for computing convex hull volume shaping each site ("FRi") and intersections between them ("intersection_k").

Note that the `betapart` package now supports external parallel computing for null models. However, this functionality is only available in `functional.betapart.core`. In this case, use the `functional.betapart` object as `x` in this function. See `functional.betapart.core` for more details.

Value

The function returns a list with the three multiple site functional dissimilarity values.

For `index.family="sorensen"` the three indices are:

<code>beta.SIM</code>	value of the functional turnover component, measured as Simpson derived functional dissimilarity
<code>beta.SNE</code>	value of the functional nestedness component, measured as nestedness-resultant fraction of Sorensen derived functional dissimilarity
<code>beta.SOR</code>	value of the overall functional beta diversity, measured as Sorensen derived functional dissimilarity

For `index.family="jaccard"` the three indices are:

<code>beta.JTU</code>	value of the functional turnover component, measured as turnover fraction of Jaccard derived functional dissimilarity
<code>beta.JNE</code>	value of the functional nestedness component, measured as nestedness-resultant fraction of Jaccard derived functional dissimilarity
<code>beta.JAC</code>	value of the overall functional beta diversity, measured as Jaccard derived functional dissimilarity

Author(s)

Sébastien Villéger, Andrés Baselga and David Orme

References

- Villéger S., Novack-Gottshal P. & Mouillot D. 2011. The multidimensionality of the niche reveals functional diversity changes in benthic marine biotas across geological time. *Ecology Letters* 14: 561-568
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21: 1223-1232
- Villéger, S. Grenouillet, G., Brosse, S. 2013. Decomposing functional beta-diversity reveals that low functional beta-diversity is driven by low functional turnover in European fish assemblages. *Global Ecology and Biogeography* 22: 671-681

See Also

[functional.beta.pair](#), [functional.betapart.core](#), [beta.multi](#)

Examples

```
##### 4 communities in a 2D functional space (convex hulls are rectangles)
traits.test<-cbind( c(1,1,1,2,2,3,3,4,4,5,5) , c(1,2,4,1,2,3,5,1,4,3,5) )
dimnames(traits.test)<-list(paste("sp",1:11,sep="") , c("Trait 1","Trait 2") )

comm.test<-matrix(0,4,11,dimnames=list( c("A","B","C","D") , paste("sp",1:11,sep="") ) )
comm.test["A",c(1,2,4,5)]<-1
comm.test["B",c(1,3,8,9)]<-1
comm.test["C",c(6,7,10,11)]<-1
comm.test["D",c(2,4,7,9)]<-1

plot(5,5,xlim=c(0,6), ylim=c(0,6), type="n", xlab="Trait 1",ylab="Trait 2")
points(traits.test[,1],traits.test[,2], pch=21,cex=1.5,bg="black")
rect(1,1,4,4, col="#458B0050", border="#458B00") ; text(2.5,2.5,"B",col="#458B00",cex=1.5)

polygon(c(2,1,3,4), c(1,2,5,4), col="#DA70D650", border="#DA70D6") ;
text(2.5,3,"D",col="#DA70D6",cex=1.5)
rect(1,1,2,2, col="#FF000050", border="#FF0000") ; text(1.5,1.5,"A",col="#FF0000",cex=1.5)

rect(3,3,5,5, col="#1E90FF50", border="#1E90FF") ; text(4,4.2,"C",col="#1E90FF",cex=1.5)

test.multi<-functional.beta.multi(x=comm.test, traits=traits.test, index.family = "jaccard" )
test.multi

test.multi.ABC<-functional.beta.multi(x=comm.test[c("A","B","C"),], traits=traits.test,
index.family = "jaccard" )
test.multi.ABC

test.multi.ABD<-functional.beta.multi(x=comm.test[c("A","B","D"),], traits=traits.test,
index.family = "jaccard" )
test.multi.ABD
```

`functional.beta.pair` *Pair-wise functional dissimilarities*

Description

Computes 3 distance matrices accounting for the spatial turnover and nestedness components of functional beta diversity, and the sum of both values. Functional dissimilarities are based on volume of convex hulls intersections in a multidimensional functional space.

Usage

```
functional.beta.pair(x, traits, index.family="sorensen")
```


Arguments

x	data frame, where rows are sites and columns are species. Alternatively x can be a functional.beta.part object derived from the functional.beta.part.core function
traits	if x is not a functional.beta.part object, a data frame, where rows are species and columns are functional space dimensions (i.e. quantitative traits or synthetic axes after PCoA). Number of species in each site must be strictly higher than number of dimensions.
index.family	family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Details

Note that the the betapart package now supports external parallel computing for null models. However, this functionality is only available in functional.beta.part.core. In this case, use the functional.beta.part object as x in this function. See functional.beta.part.core for more details.

Value

The function returns a list with three functional dissimilarity matrices.

For index.family="sorensen" the three matrices are:

funct.beta.sim	dist object, dissimilarity matrix accounting for functional turnover, measured as Simpson derived pair-wise functional dissimilarity
funct.beta.sne	dist object, dissimilarity matrix accounting for nestedness-resultant functional dissimilarity, measured as the nestedness-fraction of Sorensen derived pair-wise functional dissimilarity
funct.beta.sor	dist object, dissimilarity matrix accounting for functional beta diversity, measured as Sorensen derived pair-wise functional dissimilarity

For index.family="jaccard" the three matrices are:

funct.beta.jtu	dist object, dissimilarity matrix accounting for functional turnover, measured as the turnover-fraction of Jaccard derived pair-wise functional dissimilarity
funct.beta.jne	dist object, dissimilarity matrix accounting for nestedness-resultant functional dissimilarity, measured as the nestedness-fraction of Jaccard derived pair-wise functional dissimilarity
funct.beta.jac	dist object, dissimilarity matrix accounting for functional beta diversity, measured as Jaccard derived pair-wise functional dissimilarity

Author(s)

Sébastien Villéger, Andrés Baselga and David Orme

References

- Villéger S., Novack-Gottshal P. & Mouillot D. 2011. The multidimensionality of the niche reveals functional diversity changes in benthic marine biotas across geological time. *Ecology Letters* 14: 561-568
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21: 1223-1232
- Villéger, S. Grenouillet, G., Brosse, S. 2013. Decomposing functional beta-diversity reveals that low functional beta-diversity is driven by low functional turnover in European fish assemblages. *Global Ecology and Biogeography* 22: 671-681

See Also

[functional.beta.multi](#), [functional.betapart.core](#), [beta.pair](#)

Examples

```
##### 4 communities in a 2D functional space (convex hulls are rectangles)
traits.test<-cbind( c(1,1,1,2,2,3,3,4,4,5,5) , c(1,2,4,1,2,3,5,1,4,3,5) )
dimnames(traits.test)<-list(paste("sp",1:11,sep="") , c("Trait 1","Trait 2") )

comm.test<-matrix(0,4,11,dimnames=list( c("A","B","C","D") , paste("sp",1:11,sep="") ) )
comm.test["A",c(1,2,4,5)]<-1
comm.test["B",c(1,3,8,9)]<-1
comm.test["C",c(6,7,10,11)]<-1
comm.test["D",c(2,4,7,9)]<-1

plot(5,5,xlim=c(0,6), ylim=c(0,6), type="n", xlab="Trait 1",ylab="Trait 2")
points(traits.test[,1],traits.test[,2], pch=21,cex=1.5,bg="black")
rect(1,1,4,4, col="#458B0050", border="#458B00") ; text(2.5,2.5,"B",col="#458B00",cex=1.5)

polygon(c(2,1,3,4), c(1,2,5,4), col="#DA70D650", border="#DA70D6") ;
text(2.5,3,"D",col="#DA70D6",cex=1.5)
rect(1,1,2,2, col="#FF000050", border="#FF0000") ; text(1.5,1.5,"A",col="#FF0000",cex=1.5)

rect(3,3,5,5, col="#1E90FF50", border="#1E90FF") ; text(4,4.2,"C",col="#1E90FF",cex=1.5)

test.pair<-functional.beta.pair(x=comm.test, traits=traits.test, index.family = "jaccard" )
lapply(test.pair,round,2)
```

functional.betapart.core

Core calculations of functional dissimilarities metrics

Description

Computes the basic quantities needed for computing the multiple-site functional beta diversity measures and pairwise functional dissimilarity matrices. This version of the function now supports external parallelization for null models.

Usage

```
functional.betapart.core(x, traits, multi = TRUE, warning.time = TRUE,
return.details = FALSE, fbc.step = FALSE,
parallel = FALSE,
opt.parallel = beta.para.control())
```

Arguments

x	data frame, where rows are sites and columns are species.
traits	data frame, where rows are species and columns are functional space dimensions (i.e. quantitative traits or synthetic axes after PCoA). Number of species in each site must be strictly higher than number of dimensions.
multi	a logical value indicating whether basic quantities for multiple-site functional beta-diversity should be computed. See Details.
warning.time	a logical value indicating whether computation of multiple-site dissimilarities would stop if number of dimensions exceeds 4 or if number of sites exceeds 10. If turn to FALSE, computation process can be tracked in the step.fbc.txt file, see Details.
return.details	a logical value indicating whether volume and coordinates of vertices of convex hulls shaping each site and their intersections in the functional space should be returned.
fbc.step	a logical value indicating whether the computation progress tracking file "step.fbc.txt" should be created; Setting it to FALSE will speed up the function. It is automatically turn to FALSE when parallel is TRUE.
parallel	a logical value indicating internal parallelization is used to compute pairwise dissimilarities, see Examples. If multi is set to TRUE parallelization will be turned-off.
opt.parallel	a list of values to replace the default values returned by the function beta.para.control , to customize the cluster used for parallel computing. Defaults to an empty list.

Details

For multiple-site dissimilarities metrics ($N > 2$ sites), the volume of the union of the N convex hulls is computed using the inclusion-exclusion principle (Villéger et al., 2011). It requires to compute the volume of all the intersections between 2 to N convex hulls. Intersection between $k > 2$ convex hulls is computed as the intersection between the two convex hulls shaping intersections between the corresponding $k-1$ convex hulls, e.g. $V(A_n B_n C) = V((A_n B)_n (B_n C))$. For N sites, computing multiple-site dissimilarity metrics thus requires computing $2^N - (N+1)$ pair-wise intersections between convex hulls in a multidimensional functional space. Computation time of the intersection between two convex hulls increases with the number of dimensions (D) of the functional space. Therefore, to prevent from running very long computation process `warning.time` is set by default to stop the function if $D > 4$ or $N > 10$.

If `fbc.step` is set to TRUE, computation progress can be tracked in the "step.fbc.txt" file written in the working directory. This table shows proportion of steps completed for computing convex hull volume shaping each site ("FRi") and intersections between them ("intersection_k"). This

is only possible when computations are not performed in parallel, and this whatever the type of parallelization used (external or internal).

If `parallel` is set to `TRUE`, computation will be run through the creation of a cluster. This is interesting when beta diversity computation is long. When the number of sites increase and/or when the taxonomic richness is highly variable between sites, parallelization becomes more and more interesting. On small matrices, the running time could inflate due to the creation of the cluster and its management.

Value

The function returns an object of class `betapart` with the following elements:

<code>sumFRi</code>	the sum of the functional richness values of all sites
<code>FRt</code>	the total functional richness in the dataset
<code>a</code>	the multiple-site analog of the shared functional richness term
<code>shared</code>	a matrix containing the functional richness shared between pairs of sites
<code>not.shared</code>	a matrix containing the functional richness not shared between pairs of sites: b, c
<code>sum.not.shared</code>	a matrix containing the total functional richness not shared between pairs of sites: b+c
<code>max.not.shared</code>	a matrix containing the total maximum functional richness not shared between pairs of sites: max(b,c)
<code>min.not.shared</code>	a matrix containing the total minimum functional richness not shared between pairs of sites: min(b,c)
<code>details</code>	if <code>return.details=TRUE</code> a list of two lists: <code>\$CH</code> a list with a vector (<code>FRi</code>) of functional richness in each site (i.e. convex hull volume) and <code>coord_vertices</code> a list of <code>N</code> matrices with the coordinates of species being vertices in the <code>D</code> -dimensions functional space. <code>\$intersections</code> a list of 3 lists: <code>\$combinations</code> , <code>N-1</code> matrices with all combinations of 2 to <code>N</code> sites (numbers are rank of sites in <code>x</code>); <code>\$volumes</code> , <code>N-1</code> vectors with the volume inside the intersection between each combination of sites; <code>\$coord_vertices</code> , list of <code>N-1</code> matrices with the coordinates of the vertices shaping these intersections (NA if no intersection).

Author(s)

Sébastien Villéger, Andrés Baselga, David Orme, Renato Henriques-Silva, Maxime Logez

References

- Villéger S., Novack-Gottshal P. & Mouillot D. 2011. The multidimensionality of the niche reveals functional diversity changes in benthic marine biotas across geological time. *Ecology Letters*. 14, 561-568
- Baselga, A. 2012. The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232
- Villéger, S. Grenouillet, G., Brosse, S. 2012. Decomposing functional beta-diversity reveals that low functional beta-diversity is driven by low functional turnover in European fish assemblages. *Global Ecology and Biogeography*, in press

See Also

[functional.beta.multi](#), [functional.beta.pair](#), [betapart.core](#)

Examples

```
##### 4 communities in a 2D functional space (convex hulls are rectangles)
traits.test<-cbind( c(1,1,1,2,2,3,3,4,4,5,5) , c(1,2,4,1,2,3,5,1,4,3,5) )
dimnames(traits.test)<-list(paste("sp",1:11,sep="") , c("Trait 1","Trait 2") )

comm.test<-matrix(0,4,11,dimnames=list( c("A","B","C","D") , paste("sp",1:11,sep="") ) )
comm.test["A",c(1,2,4,5)]<-1
comm.test["B",c(1,3,8,9)]<-1
comm.test["C",c(6,7,10,11)]<-1
comm.test["D",c(2,4,7,9)]<-1

plot(5,5,xlim=c(0,6), ylim=c(0,6), type="n", xlab="Trait 1",ylab="Trait 2")
points(traits.test[,1],traits.test[,2], pch=21,cex=1.5,bg="black")
rect(1,1,4,4, col="#458B00", border="#458B00") ; text(2.5,2.5,"B",col="#458B00",cex=1.5)

polygon(c(2,1,3,4), c(1,2,5,4), col="#DA70D6", border="#DA70D6")
text(2.5,3,"D",col="#DA70D6",cex=1.5)
rect(1,1,2,2, col="#FF0000", border="#FF0000") ; text(1.5,1.5,"A",col="#FF0000",cex=1.5)

rect(3,3,5,5, col="#1E90FF", border="#1E90FF") ; text(4,4.2,"C",col="#1E90FF",cex=1.5)

test.core<-functional.betapart.core(x=comm.test, traits=traits.test, multi=TRUE,
                                   return.details=FALSE)

test.core

# using functional.betapart.core to get details on intersections
# when only pairwise dissimilarity is computed
test.core.pair<-functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
                                         return.details=TRUE)

test.core.pair

# for multiple dissimilarity
test.core.multi<-functional.betapart.core(x=comm.test, traits=traits.test, multi=TRUE,
                                          return.details=TRUE)

test.core.multi

# using core outputs to compute pairwise and multiple functional dissimilarities
functional.beta.pair(x=test.core.pair, index.family = "jaccard" )
functional.beta.multi(x=test.core.multi, index.family = "jaccard" )

## Not run:
## Not run:
##### using internal parallelisation to fasten pairwise dissimilarity
test.core.pair<- functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
                                         return.details=FALSE, fbc.step = FALSE,
```

```

parallel = FALSE)

# by default it uses half of the cores and 1 task per run (this can be customised)
# test.core.pairp <- functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
#                                           return.details=FALSE, fbc.step = FALSE,
#                                           parallel = TRUE)

# you can set the number of core to use :
test.core.pairp <- functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
                                           return.details=FALSE, fbc.step = FALSE,
                                           parallel = TRUE, opt.parallel = beta.para.control(nc = 2))
all.equal(test.core.pair, test.core.pairp)

# library(microbenchmark)
# microbenchmark(serial =
#                 functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
#                                           return.details=FALSE, fbc.step = FALSE,
#                                           parallel = FALSE),
#                 nc2 =
#                 functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
#                                           return.details=FALSE, fbc.step = FALSE,
#                                           parallel = TRUE,
#                                           opt.parallel = beta.para.control(nc = 2)),
#                 nc4 =
#                 functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
#                                           return.details=FALSE, fbc.step = FALSE,
#                                           parallel = TRUE,
#                                           opt.parallel = beta.para.control(nc = 4))
# )

# If the number of species is very different among communities
# load-balancing parallelisation could be very efficient
# especially when the number of community is high
test.core.pairp <- functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
                                           return.details=FALSE, fbc.step = FALSE,
                                           parallel = TRUE,
                                           opt.parallel = beta.para.control(nc = 2, LB = TRUE))

# use you can use fork cluster (but not on Windows)
#test.core.pairp <- functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
#                                           return.details=FALSE, fbc.step=FALSE,
#                                           parallel=TRUE,
#                                           opt.parallel=beta.para.control(nc=2, type="FORK"))

# finally you can customise the number of task run at each time
test.core.pairp <- functional.betapart.core(x=comm.test, traits=traits.test, multi=FALSE,
                                           return.details=FALSE, fbc.step = FALSE,
                                           parallel = TRUE,
                                           opt.parallel = beta.para.control(nc = 2, size = 6))

# using internal parallelisation is not always useful, especially on small data set
# load balancing is very helpful when species richness are highly variable

```

```

# Null model using 'external' parallel computing

# Example 1: pairwise functional beta diversity (functional.beta.pair)
# Note that this is an example with a small number of samples and null model
# permutations for illustration.
# Real null model analyses should have a much greater number of samples and permutations.

##### 4 communities in a 3D functional space

comm.test<-matrix(0,4,11,dimnames=list( c("A","B","C","D") , paste("sp", 1:11, sep = "")))
comm.test["A",c(1,2,4,5)]<-1
comm.test["B",c(1,3,8,9)]<-1
comm.test["C",c(6,7,10,11)]<-1
comm.test["D",c(2,4,7,9)]<-1

set.seed(1)
traits.test<-matrix(rnorm(11*3, mean=0, sd=1), 11, 3)
dimnames(traits.test)<-list(paste("sp", 1:11, sep="") , c("Trait 1","Trait 2", "Trait 3"))

# Required packages
library(doParallel)
library(picante)
library(fastmatch)
library(foreach)

# define number of cores
# Use parallel::detectCores() to determine number of cores available in your machine
nc <- 2

# 4 cores would be better (nc <- 4)

# create cluster
cl <- parallel::makeCluster(nc)

# register parallel backend
registerDoParallel(cl)

# define number of permutations for the null model (the usual is 1000)
# make sure that nperm/nc is a whole number so that all cores have the same number
# of permutations to work on
nperm <- 100

test.score <- functional.betapart.core(x = comm.test, traits = traits.test,
                                     multi = FALSE, warning.time = FALSE,
                                     return.details = FALSE,
                                     fbc.step = FALSE,
                                     parallel = FALSE)

obs.pair.func.dis <- functional.beta.pair(x = test.score, index.family = "sorensen")

# transform functional.beta.pair results into a matrix
obs.pair.func.dis <- do.call(rbind, obs.pair.func.dis)

```

```

# set names for each pair of site
pair_names <- combn(rownames(comm.test), 2, FUN = paste, collapse = "_")
colnames(obs.pair.func.dis) <- pair_names

# export necessary variables and functions to the cluster of cores
parallel::clusterExport(cl = cl, c("comm.test", "traits.test"),
                       envir = environment())

# creation of an iterator to run 1 comparisons on each core at time
it <- itertools::isplitIndices(nperm, chunkSize = 1)

# parallel computation
null.pair.func.dis <- foreach(n = it, .combine = c,
                             .packages=c("picante", "betapart", "fastmatch")) %dopar% {

# it enables to adjust the number of permutations (nt) done on each run
nt <- length(n)
null.pair.temp <- vector(mode = "list", length = nt)

# for each core "n" perform "nt" permutations
for (j in 1:nt){

# randomize community with chosen null model
# for this particular example we used the "independent swap algorithm"
# but the user can choose other types of permutation, or create it's own null model
null.comm.test <- randomizeMatrix(comm.test, null.model="independentswap",
                                  iterations=1000)

# execute functional.betapart.core function identifying each "n" core with the
# core.ident argument
null.test.score <- try(functional.betapart.core(null.comm.test, traits = traits.test,
                                              multi = FALSE, warning.time = FALSE,
                                              return.details = FALSE, fbc.step = FALSE,
                                              parallel = FALSE), silent=TRUE)

# when using 'external' parallelisation it is necessary to set parralel to FALSE

# in this artificial example there are a few combinations of species that the convex hull
# cannot be calculated due to
# some odd geometric combination so we need to re-permute the community matrix
while(inherits(null.test.score, "try-error")){

null.comm.test <- randomizeMatrix(comm.test, null.model="independentswap",
                                  iterations=1000)
null.test.score <- try(functional.betapart.core(x = null.comm.test,
                                              traits = traits.test, multi = FALSE,
                                              warning.time = FALSE, return.details = FALSE,
                                              fbc.step = FALSE, parallel = FALSE), silent=TRUE)
}

# compute the pairwise beta-diversity null values and input them in the temporary
# result matrix

```



```

res <- functional.beta.pair(x = null.test.score, index.family = "sorensen" )
null.pair.temp[[j]] <- do.call(rbind, res)

}
#retrieve the results from each core
null.pair.temp
}

# stop cluster
parallel::stopCluster(cl)

# compute the mean, standard deviation and p-values of dissimilarity metrics
# for each pair of site

mean.null.pair.func <-matrix(numeric(),ncol = ncol(obs.pair.func.dis),
                             nrow = nrow(obs.pair.func.dis))
sd.null.pair.func <-matrix(numeric(),ncol = ncol(obs.pair.func.dis),
                             nrow = nrow(obs.pair.func.dis))
p.pair.func.dis<-matrix(numeric(),ncol = ncol(obs.pair.func.dis),
                             nrow = nrow(obs.pair.func.dis))

# for each one of the 3 null dissimilarity metrics (SIN, SNE and SOR)
for (j in 1:nrow(obs.pair.func.dis)){
  matnull <- sapply(null.pair.func.dis, function(x) x[j,])
  mean.null.pair.func[j,] <- rowMeans(matnull)
  sd.null.pair.func[j,] <- sqrt(rowSums((matnull-mean.null.pair.func[j,])^2)/(nperm-1))
  p.pair.func.dis[j,] <- rowSums(matnull>=obs.pair.func.dis[j,])
  p.pair.func.dis[j,] <- (pmin(p.pair.func.dis[j,], nperm-p.pair.func.dis[j,])+1)/(nperm+1)
  # the +1 is to take into account that the observed value is one of the possibilities
}

# compute standardized effect sizes
ses.pair.func.dis <- (obs.pair.func.dis - mean.null.pair.func)/sd.null.pair.func

# Example 2: multiple functional beta diversity (functional.beta.multi)
# Note that this is an example with a small number of samples and null model
# permutations for illustration.
# Real null model analyses should have a much greater number of samples
# and permutations.

##### 4 communities in a 3D functional space

comm.test<-matrix(0,4,11,dimnames=list( c("A","B","C","D"),
                                       paste("sp", 1:11, sep = "")))

comm.test["A",c(1,2,4,5)]<-1
comm.test["B",c(1,3,8,9)]<-1
comm.test["C",c(6,7,10,11)]<-1
comm.test["D",c(2,4,7,9)]<-1

set.seed(1)
traits.test<-matrix(rnorm(11*3, mean=0, sd=1), 11, 3)

```

```

dimnames(traits.test)<-list(paste("sp", 1:11, sep=""),
                           c("Trait 1","Trait 2", "Trait 3"))

# Required packages
require(doParallel)
require(picante)
require(fastmatch)
require(foreach)

# define number of cores
# Use parallel::detectCores() to determine number of cores available in your machine
nc <- 2

# create cluster
cl <- parallel::makeCluster(nc)

# register parallel backend
registerDoParallel(cl)

# define number of permutations for the null model (the usual is 1000)
# make sure that nperm/nc is a whole number so that all cores have the same number
# of permutations to work on
nperm <- 10

# compute observed values for multiple functional dissimilarities
test.score <- functional.betapart.core(x = comm.test, traits = traits.test,
                                     multi = TRUE, warning.time = FALSE,
                                     return.details = FALSE,
                                     fbc.step = FALSE,
                                     parallel = FALSE)
obs.multi.func.dis <- do.call(cbind, functional.beta.multi(x=test.score,
                                                         index.family = "sorensen"))

# export necessary variables and functions to the cluster of cores
parallel::clusterExport(cl = cl, c("comm.test", "traits.test"),
                        envir=environment())

it <- itertools::isplitIndices(nperm, chunkSize = 1)

null.multi.func.dis <- foreach(n = it, .combine = rbind,
                              .packages = c("picante","betapart","fastmatch")) %dopar% {

# for each core, create temporary matrix to store 3 null multiple functional
# dissimilarity indices (SIN, SNE,SOR)
null.multi.temp <- matrix(numeric(), ncol = 3, nrow = length(n),
                          dimnames = list(NULL, c("funct.beta.SIM",
                                                    "funct.beta.SNE",
                                                    "funct.beta.SOR")))

# number of tasks per core (i.e., permutations per core)
nt <- length(n)

```

```

# for each core "n" perform "nt" permutations
for (j in 1:nt) {

# randomize community matrix with chosen null model (for this example
# we chose the "independent swap" algorithm)
null.comm.test <- picante::randomizeMatrix(comm.test,
                                           null.model="independentswap", iterations=1000)

# execute functional.betapart.core function identifying each "n" core
# with the core.ident argument for external parallelization,
null.test.score <- try(functional.betapart.core(x = null.comm.test,
                                              traits = traits.test, multi = TRUE, warning.time = FALSE,
                                              return.details = FALSE, fbc.step = FALSE,
                                              parallel = FALSE), silent = TRUE)

# in this artificial example there are a few combinations of species
# that the convex hull
# cannot be calculated due to some odd geometric combination so we
# need to re-permute the community matrix

while(inherits(null.test.score, "try-error")){
null.comm.test <- randomizeMatrix(comm.test, null.model="independentswap", iterations=1000)
null.test.score <- try(functional.betapart.core(x = null.comm.test, traits = traits.test,
                                              multi = TRUE, warning.time = FALSE,
                                              return.details=FALSE, fbc.step=FALSE,
                                              parallel = FALSE), silent = TRUE)
}
# input null values in the temporary result matrix
null.multi.temp[j,] <- unlist(functional.beta.multi(x = null.test.score,
                                                  index.family = "sorensen"))
}
# recover results from each core
null.multi.temp
}

# close cluster
parallel::stopCluster(cl)

# result matrix
result <- matrix(numeric(), ncol = 3, nrow = 3, dimnames = list(c("obs", "ses", "p"),
                                                              colnames(obs.multi.func.dis)))

# input observed values for the multiple functional dissimilarity indices (SIN, SNE,SOR)
result[1,] = obs.multi.func.dis

# compute standardized effect sizes (ses) for the multiple functional
# dissimilarity indices (SIN, SNE,SOR)
result[2,] <- (obs.multi.func.dis-colMeans(null.multi.func.dis, na.rm=TRUE))/
  apply(null.multi.func.dis,2, sd, na.rm=TRUE)

# compute p-values for the multiple functional dissimilarity indices (SIN, SNE,SOR)
for (i in 1:3) {
  result[3, i] <- sum(obs.multi.func.dis[i]<=null.multi.func.dis[,i])
}

```

```

    result[3, i] <- (pmin(result[3, i], nperm - result[3, i]) + 1)/(nperm+1)
  }
# the +1 is to take into account that the observed value is one of the possibilities

result
###
## End(Not run)

## End(Not run)

```

phylo.beta.multi *Multiple-site phylogenetic dissimilarities*

Description

Computes 3 distance values accounting for the multiple-site phylogenetic turnover and nestedness components of phylogenetic beta diversity, and the sum of both values. Phylogenetic dissimilarities are based on Faith's phylogenetic diversity.

Usage

```
phylo.beta.multi(x, tree, index.family="sorensen")
```

Arguments

x	a community matrix or data frame, where rows are sites and columns are species. Alternatively x can be a phylo.betapart object derived from the phylo.betapart.core function
tree	a phylogenetic tree of class phylo with tips names identical to species names from the community matrix.
index.family	family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Details

The Sorensen dissimilarity index allows computing the PhyloSor index (Bryant et al. 2008) whereas the Jaccard dissimilarity index allows computing the UniFrac index (Lozupone & Knight 2005).

Value

The function returns a list with three phylogenetic dissimilarity values.

For index.family="sorensen" the three values are:

phylo.beta.sim	dist object, dissimilarity value accounting for phylogenetic turnover, measured as Simpson derived multiple-site phylogenetic dissimilarity
phylo.beta.sne	dist object, dissimilarity value accounting for nestedness-resultant phylogenetic dissimilarity, measured as the nestedness-fraction of Sorensen derived multiple-site phylogenetic dissimilarity

phylo.beta.sor dist object, dissimilarity value accounting for phylogenetic beta diversity, measured as Sorensen derived multiple-site phylogenetic dissimilarity

For index.family="jaccard" the three values are:

phylo.beta.jtu dist object, dissimilarity value accounting for phylogenetic turnover, measured as the turnover-fraction of Jaccard derived multiple-site phylogenetic dissimilarity

phylo.beta.jne dist object, dissimilarity value accounting for nestedness-resultant phylogenetic dissimilarity, measured as the nestedness-fraction of Jaccard derived multiple-site phylogenetic dissimilarity

phylo.beta.jac dist object, dissimilarity value accounting for phylogenetic beta diversity, measured as Jaccard derived multiple-site phylogenetic dissimilarity

Author(s)

Julien De Bortoli (juldebortoli@yahoo.fr), Fabien Leprieur (fabien.leprieur@univ-montp2.fr), Andrés Baselga and David Orme

References

Baselga A. (2012) The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

Bryant JA, Lamanna C, Morlon H, Kerkhoff AJ, Enquist BJ, et al. (2008) Microbes on mountainsides: Contrasting elevational patterns of bacterial and plant diversity. *Proceedings of the National Academy of Sciences of the United States of America* 105: 11505-11511.

Faith DP, Lozupone CA, Nipperess D, Knight R (2009) The Cladistic Basis for the Phylogenetic Diversity (PD) Measure Links Evolutionary Features to Environmental Gradients and Supports Broad Applications of Microbial Ecology's "Phylogenetic Beta Diversity" Framework. *Int J Mol Sci* 10: 4723-4741. doi: 10.3390/ijms10114723.

Leprieur F, Albouy C, De Bortoli J, Cowman PF, Bellwood DR, et al. (2012) Quantifying Phylogenetic Beta Diversity: Distinguishing between "True" Turnover of Lineages and Phylogenetic Diversity Gradients. *PLoS ONE* 7(8): e42760. doi:10.1371/journal.pone.0042760

Lozupone C, Knight R (2005) UniFrac: a new phylogenetic method for comparing microbial communities. *Applied and Environmental Microbiology* 71: 8228-8235.

See Also

[phylo.betapart.core](#), [beta.multi](#)

Examples

```
# toy tree for 6 species (sp1 to sp6)
require(ape)
toy.tree<-read.tree(text="(((sp1:1,sp2:1):5,(sp3:3,sp4:3):3):2,(sp5:7,sp6:7):1);")
plot(toy.tree)

# toy community table with 6 assemblages (A to F) with 6 species (sp1 to sp6)
```

```

toy.comm<-matrix(nrow=6, ncol=6)
rownames(toy.comm)<-c("A", "B", "C", "D", "E", "F")
colnames(toy.comm)<-c("sp1", "sp2", "sp3", "sp4", "sp5", "sp6")
toy.comm[1,]<-c(1,1,1,0,0,0)
toy.comm[2,]<-c(0,1,1,1,0,0)
toy.comm[3,]<-c(0,0,1,1,1,0)
toy.comm[4,]<-c(0,0,1,1,1,1)
toy.comm[5,]<-c(0,0,0,1,1,1)
toy.comm[6,]<-c(1,0,0,1,1,1)

toy.phylobetamulti<-phylo.beta.multi(toy.comm, toy.tree, index.family="sor")
toy.betamulti<-beta.multi(toy.comm, index.family="sor")

```

phylo.beta.pair *Pair-wise phylogenetic dissimilarities*

Description

Computes 3 distance matrices accounting for the phylogenetic turnover and nestedness components of phylogenetic beta diversity, and the sum of both values. Phylogenetic dissimilarities are based on Faith's phylogenetic diversity.

Usage

```
phylo.beta.pair(x, tree, index.family="sorensen")
```

Arguments

x	a community matrix or data frame, where rows are sites and columns are species. Alternatively x can be a phylo.betapart object derived from the phylo.betapart.core function
tree	a phylogenetic tree of class phylo with tips names identical to species names from the community matrix.
index.family	family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Details

The Sorensen dissimilarity index allows computing the PhyloSor index (Bryant et al. 2008) whereas the Jaccard dissimilarity index allows computing the UniFrac index (Lozupone & Knight 2005).

Value

The function returns a list with three phylogenetic dissimilarity matrices.

For index.family="sorensen" the three matrices are:

phylo.beta.sim	dist object, dissimilarity matrix accounting for phylogenetic turnover, measured as Simpson derived pair-wise phylogenetic dissimilarity
----------------	------------------------------------------------------------------------------------------------------------------------------------------

phylo.beta.sne dist object, dissimilarity matrix accounting for nestedness-resultant phylogenetic dissimilarity, measured as the nestedness-fraction of Sorensen derived pair-wise phylogenetic dissimilarity

phylo.beta.sor dist object, dissimilarity matrix accounting for phylogenetic beta diversity, measured as Sorensen derived pair-wise phylogenetic dissimilarity

For `index.family="jaccard"` the three matrices are:

phylo.beta.jtu dist object, dissimilarity matrix accounting for phylogenetic turnover, measured as the turnover-fraction of Jaccard derived pair-wise phylogenetic dissimilarity

phylo.beta.jne dist object, dissimilarity matrix accounting for nestedness-resultant phylogenetic dissimilarity, measured as the nestedness-fraction of Jaccard derived pair-wise phylogenetic dissimilarity

phylo.beta.jac dist object, dissimilarity matrix accounting for phylogenetic beta diversity, measured as Jaccard derived pair-wise phylogenetic dissimilarity

Author(s)

Julien De Bortoli (juldebortoli@yahoo.fr), Fabien Leprieur (fabien.leprieur@univ-montp2.fr), Andrés Baselga and David Orme

References

Baselga A. (2012) The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232

Bryant JA, Lamanna C, Morlon H, Kerkhoff AJ, Enquist BJ, et al. (2008) Microbes on mountainsides: Contrasting elevational patterns of bacterial and plant diversity. *Proceedings of the National Academy of Sciences of the United States of America* 105: 11505-11511.

Faith DP, Lozupone CA, Nipperess D, Knight R (2009) The Cladistic Basis for the Phylogenetic Diversity (PD) Measure Links Evolutionary Features to Environmental Gradients and Supports Broad Applications of Microbial Ecology's "Phylogenetic Beta Diversity" Framework. *Int J Mol Sci* 10: 4723-4741. doi: 10.3390/ijms10114723.

Leprieur F, Albouy C, De Bortoli J, Cowman PF, Bellwood DR, et al. (2012) Quantifying Phylogenetic Beta Diversity: Distinguishing between "True" Turnover of Lineages and Phylogenetic Diversity Gradients. *PLoS ONE* 7(8): e42760. doi:10.1371/journal.pone.0042760

Lozupone C, Knight R (2005) UniFrac: a new phylogenetic method for comparing microbial communities. *Applied and Environmental Microbiology* 71: 8228-8235.

See Also

[phylo.betapart.core](#), [beta.pair](#)

Examples

```
# toy tree for 6 species (sp1 to sp6)
require(ape)
```

```

toy.tree<-read.tree(text="(((sp1:1,sp2:1):5,(sp3:3,sp4:3):3):2,(sp5:7,sp6:7):1);")
plot(toy.tree)

# toy community table with 6 assemblages (A to F) with 6 species (sp1 to sp6)
toy.comm<-matrix(nrow=6, ncol=6)
rownames(toy.comm)<-c("A","B","C","D","E","F")
colnames(toy.comm)<-c("sp1","sp2","sp3","sp4","sp5","sp6")
toy.comm[1,]<-c(1,1,1,0,0,0)
toy.comm[2,]<-c(0,1,1,1,0,0)
toy.comm[3,]<-c(0,0,1,1,1,0)
toy.comm[4,]<-c(0,0,1,1,1,1)
toy.comm[5,]<-c(0,0,0,1,1,1)
toy.comm[6,]<-c(1,0,0,1,1,1)

toy.phylobetapair<-phylo.beta.pair(toy.comm, toy.tree, index.family="sor")
toy.betapair<-beta.pair(toy.comm, index.family="sor")
plot(toy.betapair$beta.sim,toy.phylobetapair$phylo.beta.sim)
plot(toy.betapair$beta.sne,toy.phylobetapair$phylo.beta.sne)

```

phylo.betapart.core *Core calculations of phylogenetic dissimilarities metrics*

Description

Computes the basic quantities needed for computing the multiple-site phylogenetic beta diversity measures and pairwise phylogenetic dissimilarity matrices.

Usage

```
phylo.betapart.core(x, tree)
```

Arguments

x	a community matrix or data frame, where rows are sites and columns are species.
tree	a phylogenetic tree of class phylo with tips names identical to species names from the community matrix.

Value

The function returns a list with:

sumSi	the sum of the phylogenetic diversity values of all sites
St	the total phylogenetic diversity in the dataset
shared	a matrix containing the phylogenetic diversity shared between pairs of sites
sum.not.shared	a matrix containing the total phylogenetic diversity not shared between pairs of sites: b+c

- max.not.shared a matrix containing the total maximum phylogenetic diversity not shared between pairs of sites: max(b,c)
- min.not.shared a matrix containing the total minimum phylogenetic diversity not shared between pairs of sites: min(b,c)

Author(s)

Julien De Bortoli (juldebortoli@yahoo.fr), Fabien Leprieur(fabien.leprieur@univ-montp2.fr), Andrés Baselga and David Orme

References

- Baselga A. (2012) The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. *Global Ecology and Biogeography* 21, 1223-1232
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- Faith DP, Lozupone CA, Nipperess D, Knight R (2009) The Cladistic Basis for the Phylogenetic Diversity (PD) Measure Links Evolutionary Features to Environmental Gradients and Supports Broad Applications of Microbial Ecology's "Phylogenetic Beta Diversity" Framework. *Int J Mol Sci* 10: 4723-4741. doi: 10.3390/ijms10114723.
- Leprieur F, Albouy C, De Bortoli J, Cowman PF, Bellwood DR, et al. (2012) Quantifying Phylogenetic Beta Diversity: Distinguishing between "True" Turnover of Lineages and Phylogenetic Diversity Gradients. *PLoS ONE* 7(8): e42760. doi:10.1371/journal.pone.0042760
- Lozupone C, Knight R (2005) UniFrac: a new phylogenetic method for comparing microbial communities. *Applied and Environmental Microbiology* 71: 8228-8235.

See Also

[phylo.beta.pair](#), [phylo.beta.multi](#)

Examples

```
# toy tree for 6 species (sp1 to sp6)
require(ape)
toy.tree<-read.tree(text="(((sp1:1,sp2:1):5,(sp3:3,sp4:3):3):2,(sp5:7,sp6:7):1);")
plot(toy.tree)

# toy community table with 6 assemblages (A to F) with 6 species (sp1 to sp6)
toy.comm<-matrix(nrow=6, ncol=6)
rownames(toy.comm)<-c("A","B","C","D","E","F")
colnames(toy.comm)<-c("sp1","sp2","sp3","sp4","sp5","sp6")
toy.comm[1,]<-c(1,1,1,0,0,0)
toy.comm[2,]<-c(0,1,1,1,0,0)
toy.comm[3,]<-c(0,0,1,1,1,0)
toy.comm[4,]<-c(0,0,1,1,1,1)
toy.comm[5,]<-c(0,0,0,1,1,1)
toy.comm[6,]<-c(1,0,0,1,1,1)
```

```
toy.phylocore<-phylo.betapart.core(toy.comm, toy.tree)
```

plot.decay	<i>Plotting distance decay curves from models computed with decay.model()</i>
------------	-------------------------------------------------------------------------------

Description

Takes the output of decay.model() and plots a distance-decay curve, either a negative exponential or power law function as estimated with decay.model().

Usage

```
## S3 method for class 'decay'
plot(x, xlim=c(0,max(x$data[,1])), ylim=c(0,1),
add=FALSE, remove.dots=FALSE, col="black", pch=1, lty=1, lwd=5, cex=1, ...)
```

Arguments

x	the output of decay.model().
xlim	the range of spatial distances to be plotted, default is from 0 to the maximum distance in the data.
ylim	the range of assemblage similarities or dissimilarities to be plotted, default is from 0 to 1.
add	add to the previous plot.
remove.dots	remove the dots from the plot, thus retaining just the decay curve.
col	colour used.
pch	symbol used for points.
lty	line type.
lwd	line width.
cex	scale of text and symbols.
...	other parameters for plotting functions.

Author(s)

Andrés Baselga

References

Gómez-Rodríguez, C. & Baselga, A. 2018. Variation among European beetle taxa in patterns of distance decay of similarity suggests a major role of dispersal processes. *Ecography*, in press

See Also[decay.model](#)**Examples**

```
require(vegan)

data(BCI)
## UTM Coordinates (in metres)
UTM.EW <- rep(seq(625754, 626654, by=100), each=5)
UTM.NS <- rep(seq(1011569, 1011969, by=100), len=50)

spat.dist<-dist(data.frame(UTM.EW, UTM.NS))

dissim.BCI<-beta.pair.abund(BCI)$beta.bray.bal

plot(spat.dist, dissim.BCI, ylim=c(0,1), xlim=c(0, max(spat.dist)))

BCI.decay.exp<-decay.model(dissim.BCI, spat.dist, model.type="exp", perm=100)

BCI.decay.pow<-decay.model(dissim.BCI, spat.dist, model.type="pow", perm=100)

plot.decay(BCI.decay.exp, col=rgb(0,0,0,0.5))
plot.decay(BCI.decay.exp, col="red", remove.dots=TRUE, add=TRUE)
plot.decay(BCI.decay.pow, col="blue", remove.dots=TRUE, add=TRUE)
```

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