

# Package ‘polySegratioMM’

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**Type** Package

**Title** Bayesian mixture models for marker dosage in autopolyploids

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**Depends** R (>= 2.12.0), polySegratio

**Imports** gtools, coda, lattice

**Description** Fits Bayesian mixture models to estimate marker dosage for dominant markers on autopolyploids using JAGS (1.0 or greater) as outlined in Baker et al (2010). May be used in conjunction with polySegratio for simulation studies and comparison with standard methods.

**License** GPL-3

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polySegratioMM-package

*Marker dosage for autopolyploids by Bayesian mixture models*

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**Description**

These functions provide tools for estimating marker dosage for dominant markers in regular autopolyploids via Bayesian mixture model. Wrappers are provided for generating MCMC samples using the JAGS software. Convergence diagnostics and posterior distribution densities are provided by the coda package.

**Details**

Package: polySegratioMM  
 Type: Package  
 Version: 0.6-2  
 Date: 2012-04-10  
 License: GPL-3

The simplest way to fit a model is to use `runSegratioMM`. Given segregation ratios and a ploidy level, a mixture model is constructed with default priors and initial values and JAGS run to produce an MCMC sample for statistical inference.

A standard model may be set up with `setModel` where two parameters are set, namely `ploidy.level` or the number of homologous chromosomes set either as a numeric or as a character string and also `n.components` or the number of components for mixture model (less than or equal to maximum number of possible dosages).

Vague or strong priors may be constructed automatically using `setPriors`. Plots of standard conjugate distributions may be obtained using `DistributionPlotBinomial` `DistributionPlotGamma` and `DistributionPlotNorm`.

If necessary, other operations like setting up initial values or the control files for JAGS may be set using `setInits` `setControl` `dumpData` `dumpInits` `writeControlFile` `writeJagsFile`. Once the BUGS files and JAGS control files are set up then JAGS may be run using `runJags` and results read using `readJags`.

Convergence diagnostics may be carried out using coda or the convenience wrapper `diagnosticsJagsMix`.

Dose allocation can be carried out using `dosagesJagsMix`.

Plots may be produced and objects printed and summarised using standard `print` and `plot` methods. Plots of theoretical binomial distributions with different ploidy levels and sample sizes may be obtained with `plotFitted`. In addition, `plotFitted` produces a lattice plot of the observed segregation ratios and fitted mixture model on the logit scale.

## Author(s)

Peter Baker <p.baker1@uq.edu.au>

## References

- Baker P, Jackson P, and Aitken K. (2010) Bayesian estimation of marker dosage in sugarcane and other autopolyploids. *TAG Theoretical and Applied Genetics* **120** (8): 1653-1672.
- J B S Haldane (1930) Theoretical genetics of autopolyploids. *Journal of genetics* **22** 359–372
- Ripol, M I et al (1999) Statistical aspects of genetic mapping in autopolyploids. *Gene* **235** 31–41
- JAGS <http://www-fis.iarc.fr/~martyn/software/jags/> and [http://streaming.stat.iastate.edu/wiki/index.php/JAGS\\_Guide](http://streaming.stat.iastate.edu/wiki/index.php/JAGS_Guide)

## Examples

```
## simulate small autooctaploid data set of 100 markers for 50 individuals
## with %70 Single, %20 Double and %10 Triple Dose markers
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=400,n.individuals=275)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8) # autooctaploid mode with 3 components

## Not run:
## fit simple model in one hit with default priors, inits etc
## warning: this is too small an MCMC sample so should give inaccurate
```

```
## answers but it could still take quite a while
x.run <- runSegratioMM(sr, x, burn.in=2000, sample=5000)
print(x.run)

## plot observed segregation ratios, fitted model and expected distribution
plot(x.run, theoretical=TRUE)

## End(Not run)
```

---

calculateDIC	<i>Compute DIC for fitted mixture model</i>
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### Description

Computes and returns the Deviance Information Critereon (DIC) as suggested by Celeaux et al (2006) as their DIC\$\_4\$ for Bayesian mixture models

### Usage

```
calculateDIC(mcmc.mixture, model, priors, seg.ratios, chain=1, print.DIC=FALSE)
```

### Arguments

mcmc.mixture	Object of type <a href="#">segratioMCMC</a> produced by coda usually by using <a href="#">readJags</a>
model	object of class <code>modelSegratioMM</code> specifying model parameters, ploidy etc
priors	Object of class <code>priorsSegratioMM</code>
seg.ratios	Object of class <a href="#">segRatio</a> contains the segregation ratios for dominant markers and other information such as the number of dominant markers per individual
chain	Which chain to use when compute dosages (Default: 1)
print.DIC	Whether to print DIC

### Value

A scalar DIC is returned

### Author(s)

Peter Baker <p.baker1@uq.edu.au>

### References

- G Celeaux et. al. (2006) Deviance Information Criteria for Missing Data Models *Bayesian Analysis* **4** 23pp
- D Spiegelhalter et. el. (2002) Bayesian measures of model complexity and fit *JRSS B* **64** 583–640

**See Also**

[dosagesMCMC readJags](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## compute segregation ratios
sr <- segregationRatios(a1$markers)

## set up model, priors, inits etc and write files for JAGS
x <- setModel(3,8)
x2 <- setPriors(x)
dumpData(sr, x)
inits <- setInits(x,x2)
dumpInits(inits)
writeJagsFile(x, x2, stem="test")

## Not run:
## run JAGS
small <- setControl(x, burn.in=200, sample=500)
writeControlFile(small)
rj <- runJags(small) ## just run it
print(rj)

## read mcmc chains and print DIC
xj <- readJags(rj)
print(calculateDIC(xj, x, x2, sr))

## End(Not run)
```

---

diagnosticsJagsMix      *MCMC diagnostics for polyploid segregation ratio mixture models*

---

**Description**

Produce and/or plot various diagnostic measures from coda package for Bayesian mixture models for assessing marker dosage in autopolyploids

**Usage**

```
diagnosticsJagsMix(mcmc.mixture, diagnostics = TRUE, plots = FALSE,
  index = -c( grep("T\\[",varnames(mcmc.mixture$mcmc.list)),
             grep("b\\[",varnames(mcmc.mixture$mcmc.list)) ),
  trace.plots = FALSE, auto.corr = FALSE, density.plots = FALSE,
  xy.plots = FALSE, hpd.intervals = FALSE, hdp.prob = 0.95,
  return.results = FALSE)
```

**Arguments**

<code>mcmc.mixture</code>	Object of class <a href="#">segratioMCMC</a> or <a href="#">runJagsWrapper</a> after JAGS run produced by coda
<code>diagnostics</code>	if TRUE then print several coda dignostic tests
<code>plots</code>	if TRUE then produce several coda dignostic plots
<code>index</code>	index of parameters for disgnostic tests/plots (Default: mixture model (and random effects) parameters)
<code>trace.plots</code>	if TRUE plot mcmc traces (default: FALSE)
<code>auto.corr</code>	if TRUE produce autocorrelations of mcmc's (default: FALSE)
<code>density.plots</code>	if TRUE plot parameter densities (default: FALSE)
<code>xy.plots</code>	if TRUE plot traces using 'lattice' (default: FALSE)
<code>hpd.intervals</code>	if TRUE print and return highest posterior density intervals for parameters specified by <code>index</code>
<code>hpd.prob</code>	probability for <code>hpd.intervals</code>
<code>return.results</code>	if TRUE return results as list

**Value**

If `return.results` is TRUE then a list is returned with components depending on various settings of arguments

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[mcmc](#) [autocorr.diag](#) [raftery.diag](#) [geweke.diag](#) [gelman.diag](#) [trellisplots](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)

## Not run:
## fit simple model in one hit

x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)
print(x.run)
diagnosticsJagsMix(x.run)
diagnosticsJagsMix(x.run, plot=TRUE)

## End(Not run)
```

---

 DistributionPlotBinomial

*Distribution Plot*


---

### Description

Plots probability density function given the parameters. May be useful when investigating parameter choice for prior distributions.

### Usage

```
DistributionPlotBinomial(size = 200, prob = 0.5,
  xlab = "Number of Successes", ylab = "Probability Mass", signif.digits = 3,
  main = paste("Binomial Distribution: n =", size, "p =",
  signif(prob, digits = signif.digits)))
```

```
DistributionPlotGamma(shape = 1, rate = 1, length = 100, xlab = "x",
  ylab = "Density", main = bquote(paste("Gamma Distribution: ", alpha,
  "=", .(signif(shape, digits = signif.digits)), ", ", beta, "=",
  .(signif(rate, digits = signif.digits)))), signif.digits = 3)
```

```
DistributionPlotNorm(mean = 0, sd = 1, length = 100, xlab = "x", ylab =
  "Density", main = bquote(paste("Normal Distribution: ", mu, "=",
  .(signif(mean, digits = signif.digits)), ", ", sigma, "=", .(signif(sd,
  digits = signif.digits)))), signif.digits = 3)
```

### Arguments

size	number of trials (Binomial)
prob	probability of success (Binomial)
shape	shape parameter. Must be strictly positive. (Gamma)
rate	an alternative way to specify the scale (Gamma)
mean	mean (Normal)
sd	standard deviation (Normal)
xlab	x-axis label
ylab	y-axis label
signif.digits	number of significant digits for default main title
main	title for plot
length	Number of points to use for obtaining a smooth curve

### Details

Based on functions in package Rcmdr

**Value**

None.

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[Rcmdr](#) [Binomial](#) [Normal](#) [GammaDist](#)

**Examples**

```
## Binomial distribution
DistributionPlotBinomial()
DistributionPlotBinomial(size=20, prob=0.2)

## Gamma distribution
DistributionPlotGamma()

## Normal distribution
DistributionPlotNorm()
```

---

dosagesJagsMix

*Compute dosages under specified Bayesian mixture model*

---

**Description**

Computes and returns estimated dosages under specified model using posterior probabilities derived from mcmc chains by the proportion of samples in each dosage class.

**Usage**

```
dosagesJagsMix(mcmc.mixture, jags.control, seg.ratio, chain = 1,
max.post.prob = TRUE, thresholds = c(0.5, 0.6, 0.7, 0.8, 0.9, 0.95,
0.99), print = FALSE, print.warning = TRUE, index.sample = 20)
```

**Arguments**

mcmc.mixture	Object of type <a href="#">segratioMCMC</a> produced by coda usually by using <a href="#">readJags</a>
jags.control	Object of class <a href="#">jagsControl</a> for setting up JAGS command file
seg.ratio	Object of class <a href="#">segRatio</a> contains the segregation ratios for dominant markers and other information such as the number of dominant markers per individual
chain	Which chain to use when compute dosages (Default: 1)
max.post.prob	Logical for producing dose allocations based on the maximum posterior probability (Default: TRUE)



thresholds	Numeric vector of thresholds for allocating dosages when the posterior probability to a particular dosage class is above the threshold
print	Logical indicating whether or not to print intermediate results (Default: FALSE)
print.warning	Logical to print warnings if there is more than one marker with the maximum posterior probability
index.sample	Numeric vector indicating which markers to print if print is TRUE. If index.sample is of length 1 then a random sample of size index.sample is selected

**Value**

An object of class `dosagesMCMC` is returned with components:

p.dosage	Matrix of posterior probabilities of dosages for each marker dosage
dosage	Matrix of allocated dosages based on posterior probabilities. The columns correspond to different 'thresholds' and if requested, the last column is allocated on basis of max.post
thresholds	vector of cutoff probabilities for dosage class
chain	Chain used to compute dosages
max.post	maximum dosage posterior probabilities for each marker
index.sample	Numeric vector indicating which markers to print if print is TRUE. If index.sample is of length 1 then a random sample of size index.sample is selected

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[dosagesMCMC](#) [readJags](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## compute segregation ratios
sr <- segregationRatios(a1$markers)

## set up model, priors, inits etc and write files for JAGS
x <- setModel(3,8)
x2 <- setPriors(x)
dumpData(sr, x)
inits <- setInits(x,x2)
dumpInits(inits)
writeJagsFile(x, x2, stem="test")

## Not run:
## run JAGS
small <- setControl(x, burn.in=200, sample=500)
```

```

writeControlFile(small)
rj <- runJags(small) ## just run it
print(rj)

## read mcmc chains and produce dosage allocations
xj <- readJags(rj)
dd <- dosagesJagsMix(xj, small, sr)
print(dd)

## End(Not run)

```

---

dumpData

*Dumps segregation ratio data to file for subsequent JAGS run*


---

### Description

Given segregation ratio data provided as an object of class `segRatio`, data are dumped in R format for use by JAGS

### Usage

```

dumpData(seg.ratio, model, stem = "test", fix.one = TRUE,
         data.file = paste(stem, "-data.R", sep = ""))

```

### Arguments

<code>seg.ratio</code>	Object of class <code>segRatio</code> contains the segregation ratios for dominant markers and other information such as the number of dominant markers per individual
<code>model</code>	Object of class <code>modelSegratioMM</code> containing mixture model information
<code>stem</code>	File name stem for data file (default "test")
<code>fix.one</code>	Logical to fix the dosage of the observation closest to the centre of each component on the logit scale. This can greatly assist with convergence (Default: TRUE)
<code>data.file</code>	Data file name which is automatically generated from <code>stem</code> if not specified

### Value

None.

### Author(s)

Peter Baker <p.baker1@uq.edu.au>

### See Also

[segRatio dump](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## compute segregation ratios
sr <- segregationRatios(a1$markers)

## set up model for 3 components of autooctaploid
x <- setModel(3,8)

dumpData(sr, x)
```

---

hexmarkers	<i>Simulated autopolyploid dominant markers from 200 hexaploid individuals</i>
------------	--------------------------------------------------------------------------------

---

**Description**

These data were simulated as 500 markers for 200 “auto-hexaploid individuals” exhibiting no overdispersion. The underlying percentages of single double and triple dose markers are 70%, 20% and 10%, respectively.

**Usage**

```
hexmarkers
```

**Format**

An object of S3 class `sim.autoMarkers` containing 500 simulated dominant markers for 200 auto-hexaploid individuals.

**References**

Haldane, J B S. 1930. Theoretical genetics of autopolyploids. *Journal of Genetics* 22: 359-372.

Baker P, Jackson P, and Aitken K. 2010. Bayesian estimation of marker dosage in sugarcane and other autopolyploids. *TAG Theoretical and Applied Genetics* 120 (8): 1653-1672.

---

hexmarkers.overdisp	<i>Simulated overdispersed autopolyploid dominant markers from 200 hexaploid individuals</i>
---------------------	----------------------------------------------------------------------------------------------

---

### Description

These data are simulated as 500 markers for 200 “auto–hexaploid individuals” exhibiting overdispersion with the parameter  $\text{shape1} = 25$ . The underlying percentages of single double and triple dose markers are 70%, 20% and 10%, respectively.

### Usage

```
hexmarkers.overdisp
```

### Format

An object of S3 class `sim.autoMarkers` containing 500 simulated dominant markers for 200 auto–hexaploid individuals.

### References

- Haldane, J B S. 1930. Theoretical genetics of autopolyploids. *Journal of Genetics* 22: 359-372.
- Baker P, Jackson P, and Aitken K. 2010. Bayesian estimation of marker dosage in sugarcane and other autopolyploids. *TAG Theoretical and Applied Genetics* 120 (8): 1653-1672.

---

mcmcHexRun	<i>Results of MCMC estimation for simulated overdispersed markers</i>
------------	-----------------------------------------------------------------------

---

### Description

MCMC was performed using the wrapper function `runSegratioMM` to run JAGS for a Bayesian mixture model on the segregation ratios obtained using the simulated data `hexmarkers.overdisp`. These data were simulated as 500 markers for 200 “auto–hexaploid individuals” exhibiting overdispersion with  $\text{shape1}=25$ . The underlying percentages of single double and triple dose markers are 70%, 20% and 10%, respectively.

### Usage

```
mcmcHexRun
```

### Format

An object of S3 class `runJagsWrapper` with various components including summaries and diagnostics.

## References

Baker P, Jackson P, and Aitken K. 2010. Bayesian estimation of marker dosage in sugarcane and other autopolyploids. *TAG Theoretical and Applied Genetics* 120 (8): 1653-1672.

---

plot.segratioMCMC      *MCMC plots for segregation ratio mixture models*

---

## Description

Standard MCMC trace and density plots for specified mixture model parameters and posterior probability distributions for specified markers

## Usage

```
## S3 method for class 'segratioMCMC'  
plot(x, ..., row.index = c(1:10), var.index = c(1:6),  
marker.index = c(1:8))
```

## Arguments

x	object of class segratioMCMC
...	extra options for printing
row.index	which rows to print (Default: first 10)
var.index	which mixture model variable to summarise (Default: all)
marker.index	which markers to summarise (Default: 1:8)

## Value

None.

## Author(s)

Peter Baker <p.baker1@uq.edu.au>

## See Also

[dosagesMCMC](#) [readJags](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)

## Not run:
## fit simple model in one hit and summarise

x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)
plot(x.run$mcmc.mixture)

## End(Not run)
```

---

plotFitted

*Plot observed segregation ratios and fitted and theoretical models*


---

**Description**

Plots histogram of observed segregation ratios on logit scale along with scaled density of fitted components corresponding to dosage classes. Plots of expected theoretical distributions can be plotted with or without segregation ratio data.

**Usage**

```
## S3 method for class 'runJagsWrapper'
plot(x, theoretical=FALSE, ...)

plotFitted(seg.ratios, summary.mixture, add.random.effect=TRUE,
  theoretical=FALSE, model=NULL, theory.col="red",
  xaxis=c("logit","raw"), ylim=NULL, NCLASS=NULL, n.seq=100,
  xlab="logit(Segregation Ratio)", ylab="Density", density.plot=FALSE,
  fitted.lwd=2, fitted.col="blue", bar.col="lightgreen", cex=1,
  warnings = FALSE, main=NULL, ...)

plotTheoretical(ploidy.level=8, seg.ratios=NULL, n.components=NULL,
  expected.segratio=NULL, proportions=c(0.65,0.2,0.1,0.03,0.01,0.01, 0, 0),
  n.individuals=200, xaxis=c("raw","logit"),
  type.parents=c("heterogeneous","homozygous"), xlim=c(0,1),
  NCLASS=NULL, xlab="Segregation Ratio", ylab="Density",
  density.plot=FALSE, fitted.lwd=2, fitted.col="blue", cex=1,
  warnings = TRUE, main=NULL, ...)
```

**Arguments**

<code>x</code>	object of class <code>runJagsWrapper</code> produced by using <code>runSegratioMM</code> to set up and fit mixture model
<code>seg.ratios</code>	segregation ratios as class <code>segRatio</code>
<code>summary.mixture</code>	mcmc summary data produce by <code>summary.segratioMCMC</code>
<code>add.random.effect</code>	add random variance component to fitted distribution plot if model includes a random effect (default: TRUE)
<code>theoretical</code>	whether to plot the expected theoretical distribution under the fitted model (default: FALSE)
<code>model</code>	object of class <code>modelSegratioMM</code> specifying model if plotting expected theoretical distribution
<code>theory.col</code>	colour for expected theoretical distribution (default: "red")
<code>ploidy.level</code>	the number of homologous chromosomes
<code>n.components</code>	number of components for mixture model
<code>expected.segratio</code>	may be specified or automatically calculated from ploidy level etc
<code>xaxis</code>	whether to plot on "logit" or "raw" scale. Defaults to "logit" if plotting segregation ratios or "raw" for theoretical distributions
<code>proportions</code>	for no. of markers in each component of theoretical distribution plot
<code>n.individuals</code>	for theoretical distribution plot - taken from segregation ratios if supplied
<code>type.parents</code>	"heterogeneous" if parental markers are 0,1 or "homogeneous" if parental markers are both 1
<code>ylim</code>	<code>c(lower,upper)</code> yaxis limits for histogram of segregation ratios
<code>xlim</code>	<code>c(lower,upper)</code> xaxis limits for segregation ratios
<code>NCLASS</code>	number of classes for histogram (Default: 100)
<code>n.seq</code>	number of points to use for plotting fitted mixture
<code>xlab</code>	x-axis label
<code>ylab</code>	y-axis label
<code>density.plot</code>	whether to plot a smoothed density as well as segregation data and fitted and/or theoretical distributions (default: FALSE)
<code>main</code>	title for plot
<code>fitted.lwd</code>	width for fitted line
<code>fitted.col</code>	colour for fitted line
<code>bar.col</code>	colour for histogram
<code>cex</code>	character expansion for text (see <a href="#">par</a> )
<code>warnings</code>	print warnings like number of components etc (Default: FALSE)
<code>...</code>	extra options for plot

**Details**

`plotFitted` plot histogram of observed segregation ratios on logit scale along with scaled density of fitted components corresponding to dosage classes using trellis

`plotTheoretical` plot expected distribution of autopolyploid dominant markers on probability (0,1) scale. Segregation ratios may also be plotted

`plot.runJagsWrapper` plots the fitted values of object of class `runJagsWrapper` which has been produced by using `runSegratioMM` to set up and fit mixture model

Note that since trellis graphics are employed, plots may need to be printed in order to see them

**Value**

None.

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[summary.mcmc](#) [mcmc](#) [segratioMCMC](#) [readJags](#) [diagnosticsJagsMix](#) [runSegratioMM](#)

**Examples**

```
## simulate small autooctaploid data set
plotTheoretical(8, proportion=c(0.7,0.2,0.1),n.individuals=50)
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)

## fit simple model in one hit and summarise
## Not run:
x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)
print(x.run)

## plot fitted model using 'plotFitted'
plotFitted(sr, x.run$summary)
a.plot <- plotFitted(sr, x.run$summary, density.plot=TRUE)
print(a.plot)
## or the easier way
plot(x.run, theoretical=TRUE)

## End(Not run)
```



---

print.dosagesMCMC      *Doses from Bayesian mixture model*

---

## Description

Prints objects of S3 class dosagesMCMC or segratioMCMC

## Usage

```
## S3 method for class 'dosagesMCMC'
print(x, ..., index.sample = 20)

## S3 method for class 'segratioMCMC'
print(x, ..., row.index = c(1:10), var.index = c(1:6), marker.index
      = c(1:8), chain = 1)
```

## Arguments

x	object of class dosagesMCMC or segratioMCMC
...	extra options for printing
index.sample	which markers to print (Default: 20 markers at random)
row.index	which rows to print (Default: first 10)
var.index	which mixture model variable to summarise (Default: all)
marker.index	which markers to summarise (Default: 1:8)
chain	which chain to print (Default: 1)

## Value

None.

## Author(s)

Peter Baker <p.baker1@uq.edu.au>

## See Also

[dosagesMCMC readJags](#)

## Examples

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)
```

```
## fit simple model in one hit

## Not run:
x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)

print(x.run$doses)

## End(Not run)
```

---

print.runJags

*Running JAGS*

---

## Description

Print details and timing of JAGS run and summaries of results

## Usage

```
## S3 method for class 'runJags'
print(x, ...)
## S3 method for class 'runJagsWrapper'
print(x, ...)
```

## Arguments

x	Objects of class runJags or runJagsWrapper
...	extra printing options

## Details

print.runJags can be employed when runJags is called directly and reports timings and dates while print.runJagsWrapper provides summary statistics when runSegratioMM is used.

## Value

None.

## Author(s)

Peter Baker <p.baker1@uq.edu.au>

## See Also

[runJags](#) [runSegratioMM](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)

## Not run:
## fit simple model in one hit

x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)
print(x.run)

## End(Not run)
```

---

readJags

*Read MCMC sample(s) from a JAGS run*


---

**Description**

wrapper to `read.openbugs` which returns object of class `mcmc.list` and so can be used to specify the start and end iterations for the MCMC sample(s) and also specify thinning

**Usage**

```
readJags(run.jags, quiet = TRUE, ...)
```

**Arguments**

<code>run.jags</code>	object of class <code>runJAGS</code> produced by running JAGS
<code>quiet</code>	logical to return program output (Default: TRUE)
<code>...</code>	other options for <code>read.openbugs</code>

**Value**

Returns object of class `segratioMCMC` with components

<code>run.jags</code>	object of class <code>runJAGS</code> produced by running JAGS
<code>mcmc.list</code>	object of class <code>mcmc.list</code> containing the MCMC sample(s)

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[mcmc.list](#) [setPriors](#) [setInits](#) [expected.segRatio](#) [segRatio](#) [setControl](#) [dumpData](#) [dumpInits](#)  
or for an easier way to run a segregation ratio mixture model see [runSegratioMM](#)

## Examples

```

library(polySegratio)

## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)
x2 <- setPriors(x)
cat(x$bugs.code,x2$bugs.code,sep="\n")

x3 <- setModel(3,8, random.effect = TRUE)
x4 <- setPriors(x3, type="strong")

dumpData(sr, x3)
inits <- setInits(x,x2)
dumpInits(inits)
##x.priors <- setPriors(x, "vague")
writeJagsFile(x, x2, stem="test")

small <- setControl(x, burn.in=20, sample=50)
writeControlFile(small)
## Not run:
rj <- runJags(small) ## just run it

xj <- readJags(rj)
print(xj)

## End(Not run)

```

---

runJags

*Run JAGS to create MCMC sample for segregation ratio mixture model*


---

## Description

Runs external program JAGS and returns MCMC list for processing by coda.

## Usage

```
runJags(jags.control, jags = "jags", quiet = FALSE,
       cmd.file = paste(jags.control$stem, ".cmd", sep = ""), timing = TRUE)
```

## Arguments

`jags.control` Object of class `jagsControl` containing MCMC burn in, sample and thinning as well as relevant files for BUGS commands, inits and data

jags	Name of JAGS program assumed to be in PATH. However, jags may explicitly set here to include the full path name
quiet	Logical to return program output (Default: FALSE)
cmd.file	JAGS .cmd command file (Default: deduced from jags.control)
timing	Logical to return timing information such as date started and ended and elapsed user and system time

**Value**

Returns object of class runJAGS with components

jags.control	Object of class jagsControl
exit	integer indicating return error (0 if no errors)
cmd.file	JAGS command file
start.time	time JAGS run started
end.time	time JAGS run finished
elapsed.time	elapsed user and system time
call	function call

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[setPriors](#) [setInits](#) [expected.segRatio](#) [segRatio](#) [setControl](#) [dumpData](#) [dumpInits](#) or for an easier way to run a segregation ratio mixture model see [runSegratioMM](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
sr <- segregationRatios(a1$markers)

## set up model with 3 components
x <- setModel(3,8)
x2 <- setPriors(x)
dumpData(sr, x)
inits <- setInits(x,x2)
dumpInits(inits)
##x.priors <- setPriors(x, "vague")
writeJagsFile(x, x2, stem="test")

## Not run:
small <- setControl(x, burn.in=20, sample=50)
writeControlFile(small)
rj <- runJags(small) ## just run it
print(rj)

## End(Not run)
```

---

runSegratioMM                      *Run a Bayesian mixture model for marker dosage with minimal effort*

---

### Description

Given segregation ratios and a ploidy level, a mixture model is constructed with default priors and initial values and JAGS run to produce an MCMC sample for statistical inference. Returns an object of S3 class runJagsWrapper

### Usage

```
runSegratioMM(seg.ratios, model, priors = setPriors(model),
  inits = setInits(model, priors), jags.control =
  setControl(model, stem, burn.in = burn.in, sample = sample, thin = thin),
  burn.in = 2000, sample = 5000, thin = 1, stem = "test", fix.one = TRUE,
  print = TRUE, plots = TRUE, print.diagnostics = TRUE,
  plot.diagnostics = TRUE, run.diagnostics.later=FALSE )
```

### Arguments

seg.ratios	Object of class <code>segRatio</code> contains the segregation ratios for dominant markers and other information such as the number of dominant markers per individual
model	object of class <code>modelSegratioMM</code> specifying model parameters, ploidy etc
priors	object of class <code>priorsSegratioMM</code> indicating priors that are “vague”, “strong” or “specified”
inits	A list of initial values usually produced by <code>setInits</code>
jags.control	Object of class <code>jagsControl</code> containing MCMC burn in, sample and thinning as well as relevant files for BUGS commands, inits and data
burn.in	size of MCMC burn in (Default: 2000)
sample	size of MCMC sample (default: 5000)
thin	thinning interval between consecutive observations (default: 1 or no thinning)
stem	text to be used as part of JAGS .cmd file name
fix.one	Logical to fix the dosage of the observation closest to the centre of each component on the logit scale. This can greatly assist with convergence (Default: TRUE)
print	logical for printing monitoring and summary information (default: TRUE)
plots	logical to plotting MCMC posterior distributions (default: TRUE)
print.diagnostics	logical for printing diagnostic statistics (default: TRUE)
plot.diagnostics	logical for diagnostic plots (default: TRUE)
run.diagnostics.later	should diagnostics be run later which may help if there are convergence problems (Default: FALSE)

**Value**

Returns object of class runJagsWrapper with components

seg.ratios	Object of class <a href="#">segRatio</a> contains the segregation ratios for dominant markers
model	object of class <a href="#">modelSegratioMM</a> specifying model parameters, ploidy etc
priors	Object of class <a href="#">priorsSegratioMM</a> specifying prior distributions
inits	A list of initial values usually produced by <a href="#">setInits</a>
jags.control	Object of class <a href="#">jagsControl</a> containing MCMC burn in, sample and thinning as well as relevant files for BUGS commands, inits and data
stem	text to be used as part of JAGS .cmd file name and other files
fix.one	Logical to fix the dosage of the observation closest to the centre of each component on the logit scale. This can greatly assist with convergence (Default: TRUE)
run.jags	object of class <a href="#">runJAGS</a> produced by running JAGS
mcmc.mixture	Object of type <a href="#">segratioMCMC</a> produced by coda usually by using <a href="#">readJags</a>
diagnostics	list containing various diagnostic summaries and statistics produced by coda
summary	summaries of posterior distributions of model parameters
doses	object of class <a href="#">dosagesMCMC</a> containing posterior probabilities of dosages for each marker dosage and allocated dosages
DIC	Deviance Information Critereon

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[setPriors](#) [setInits](#) [expected.segRatio](#) [segRatio](#) [setControl](#) [dumpData](#) [dumpInits](#) and [diagnosticsJagsMix](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)

## Not run:
## fit simple model in one hit

x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)
print(x.run)

## End(Not run)
```

---

 setControl

*Set up controls for a JAGS segregation ratio model run*


---

**Description**

Sets up directives for running JAGS which are subsequently put into a .cmd file. MCMC attributes such as the size of burn in, length of MCMC and thinning may be specified

**Usage**

```
setControl(model, stem = "test", burn.in = 2000, sample = 5000, thin = 1,
  bugs.file = paste(stem, ".bug", sep = ""),
  data.file = paste(stem, "-data.R", sep = ""),
  inits.file = paste(stem, "-inits.R", sep = ""),
  monitor.var = model$monitor.var, seed=1)
```

**Arguments**

model	object of class <code>modelSegratioMM</code> specifying model parameters, ploidy etc
stem	text to be used as part of JAGS .cmd file name
burn.in	size of MCMC burn in (Default: 2000)
sample	size of MCMC sample (default: 5000)
thin	thinning interval between consecutive observations. Thinning may be a scalar or specified for each variable set by specifying a vector (default: 1 or no thinning)
bugs.file	name of .bug file
data.file	name of R data file
inits.file	name of R inits file
monitor.var	which variables to be monitored (Default: as per model)
seed	seed for JAGS run for Windows only (for unix set seed in <code>setInits</code> )

**Value**

Returns an object of class `jagsControl` which is a list with components

jags.code	Text containing control statements for JAGS .cmd file
model	object of class <code>modelSegratioMM</code> specifying model parameters, ploidy etc
stem	text to be used as part of JAGS .cmd file name
burn.in	size of MCMC burn in (Default: 2000)
sample	size of MCMC sample (default: 5000)
thin	thinning interval between consecutive observations
bugs.file	name of .bug file
data.file	name of R data file
inits.file	name of R inits file
monitor.var	which variables to be monitored
call	function call



**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[setModel](#) [setInits](#) [expected.segRatio](#) [segRatio](#) [setControl](#) [dumpData](#) [dumpInits](#) or for an easier way to run a segregation ratio mixture model see [runSegratioMM](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## set up model with 3 components
x <- setModel(3,8)
x2 <- setPriors(x)

jc <- setControl(x)
print(jc)
```

---

setInits

*Set up and dump initial values given the model and prior*


---

**Description**

Given a model of class `modelSegratioMM` and priors of class `priorsSegratioMM`, initial values are computed using approximate expected values by `setInits` and then written to file by `dumpInits`

**Usage**

```
setInits(model, priors, seed = 1)

dumpInits(inits, stem = "test", inits.file = paste(stem, "-inits.R",
  sep = ""))
```

**Arguments**

<code>model</code>	Object of class <code>modelSegratioMM</code> providing model attributes like the number of components and ploidy level
<code>priors</code>	Object of class <code>priorsSegratioMM</code>
<code>seed</code>	Seed to be used for JAGS runs. If a number of chains are to be run a vector of starting values may be specified. However, see note below.
<code>inits</code>	A list of initial values usually produced by <code>setInits</code>
<code>stem</code>	File name stem for inits file (default "test")
<code>inits.file</code>	Inits file name which is automatically generated from <code>stem</code> if not specified

**Value**

Returns a list with the following initial values:

mu	Mean of dosage classes on logit scale: usually $c(0, NA, NA, \dots, NA)$
P	Initial value for proportion in each dosage class
tau	Precision of means which depends on whether priors are strong or weak
theta	Differences in means (for parameterisation employed for better convergence)
seed	Sets seed for each MCMC chain (Default: 1)
taub	If the model contains a random effect then sets initial value of precision of random effect $b$ which is normally distributed with mean 0 and precision tau

**Note**

*Warning:* If a number of chains are to be produced then several seeds may be specified. Currently, this is largely untested and so it is highly unlikely that this will actually work for all functions in this package.

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[setModel](#) [setPriors](#) [setControl](#) [dumpInits](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## set up model, priors, inits etc and write files for JAGS
x <- setModel(3,8)
x2 <- setPriors(x)
inits <- setInits(x,x2)
dumpInits(inits)
```

---

setModel

*Set characteristics of the Bayesian mixture model for dosages*

---

**Description**

Used to automatically set up Bayesian finite mixture models for dosage allocation of dominant markers in autopolyploids given the number of components and ploidy level

**Usage**

```
setModel(n.components, ploidy.level, random.effect = FALSE, seg.ratios =NULL,
ploidy.name = NULL, equal.variances=TRUE,
type.parents = c("heterogeneous", "homozygous"))
```

**Arguments**

n.components	number of components for mixture model (less than or equal to maximum number of possible dosages)
ploidy.level	the number of homologous chromosomes, either as numeric or as a character string
random.effect	Logical indicating whether model contains random effect (Default: FALSE)
seg.ratios	segregation proportions for each marker provided as S3 class segRatio
ploidy.name	Can override ploidy name here or allow it to be determined from ploidy.level
equal.variances	Logical indicating whether model contains separate or common variances for each component (Default: TRUE)
type.parents	"heterogeneous" if parental markers are 0,1 or "homogeneous" if parental markers are both 1

**Value**

Returns object of class modelSegratioMM with components

bugs.code	text to be used by JAGS in the .bug file but without statements pertaining to priors
n.components	number of components for mixture model
monitor.var	names of variables to be monitored in JAGS run
ploidy.level	ploidy level
random.effect	Logical indicating whether model contains random effect (Default: FALSE)
equal.variances	Logical indicating equal or separate variances for each component
E.segRatio	Expected segregation ratios
type.parents	"heterogeneous" if parental markers are 0,1 or "homogeneous" if parental markers are both 1
call	function call

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[setPriors](#) [setInits](#) [expected.segRatio](#) [segRatio](#) [setControl](#) [dumpData](#) [dumpInits](#) or for an easier way to run a segregation ratio mixture model see [runSegratioMM](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## set up model with 3 components
x <- setModel(3,8)
print(x)
```

---

setPriors	<i>Set prior distributions for parameters of Bayesian mixture model for dosages</i>
-----------	-------------------------------------------------------------------------------------

---

**Description**

May be used to automatically set up vague or strong priors or explicitly set them for Bayesian finite mixture model specified as an object of class `modelSegratioMM` using `setModel`

**Usage**

```
setPriors(model, type.prior = c("strong",
                              "vague", "strong.tau", "strong.s", "specified"),
          mean.vague = 0.1, prec.vague = 0.1, A.vague = 0.1, B.vague = 0.1,
          prec.strong=400, n.individuals=200, reffect.A = 44, reffect.B = 0.8,
          M.sd = 0.025, STRONG.PREC=c(0.025, 0.975), UPPER = 0.995, PREC.INT=0.2,
          params = NULL, segRatio = NULL)
```

**Arguments**

model	object of class <code>modelSegratioMM</code> specifying model parameters, ploidy etc
type.prior	The type of prior required being one of “strong”, “vague”, “strong.tau” “strong.s” or “specified”. The first four prior types will automatically set prior distributions whereas for the last, namely “specified”, the prior distribution parameters must be set explicitly. Note that strong priors get progressively stronger from “strong” to “strong.s”
mean.vague	The mean of Normal priors for a “vague” prior
prec.vague	The precision of Normal priors for a “vague” prior
A.vague	The shape parameter of the Gamma prior for the precision parameters for a “vague” prior
B.vague	The rate (scale) parameter of the Gamma prior for the precision parameters for a “vague” prior
prec.strong	Precision for Normal mean parameters when type.prior is “strong”. Note that on logit scale default is equivalent to having a 95%CI as +/- 0.1
n.individuals	Used for Binomial calculations to set prior precision parameters when type.prior is “strong”.

reffect.A	The shape parameter of the Gamma prior for the precision parameter of the random.effect for a “vague” prior
reffect.B	The rate (scale) parameter of the Gamma prior for the precision parameter of the random.effect for a “vague” prior
M.sd	Approximate standard deviation for the mean segregation ratios on raw probability scale - this is set to 0.025 which would give an approximate 95% interval of 0.1 for the segregation ratio
UPPER	Cutoff for guessing parameters on logit scale noting that $\text{logit}(1)$ is undefined
STRONG.PREC	Interval on raw probability scale used to set strong priors on the the precision distribution parameters of the segregation ratios by using a 95% interval on the theoretical distribution and equating this on the logit scale (Default: $c(0.025, 0.975)$ )
PREC.INT	Multiplier or setting prior for precision on logit scale corresponding to approx confidence region being $\text{precision} * (1 - \text{PREC.INT}, 1 + \text{PREC.INT})$ Default: 0.2
params	if type.prior is “specified” then a list of priors parameters must be set containing components M for means, A and B for gamma prior parameters and if the model contains a random.effect then reffect.A, and reffect.B for the gamma prior for the precision of random effect taub. Note that the lengths of M, prec, A and B should be equal to the number of components
segRatio	If specified, this value overrides the automatically generated value which is set as the expected segregation ratio given the ploidy level

### Value

Returns an object of class priorsSegratioMM which is a list with components

type	Type of prior: one of “vague”, “strong” or “specified”
bugs.code	Text containing prior statements for BUGS file
random.effect	Logical indicating whether model contains random effect (Default: FALSE)
equal.variances	Logical indicating equal or separate variances for each component
params	List containing Normal means on logit scale <code>logit.means</code> , precision on logit scale <code>logit.prec</code> , and Gamma parameters A and B and finally <code>reffect.A</code> and <code>reffect.B</code> if the model contains a random effect
call	function call

### Author(s)

Peter Baker <p.baker1@uq.edu.au>

### See Also

[setModel](#) [setInits](#) [expected.segRatio](#) [segRatio](#) [setControl](#) [dumpData](#) [dumpInits](#) or for an easier way to run a segregation ratio mixture model see [runSegratioMM](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## set up model with 3 components
x <- setModel(3,8)
x2 <- setPriors(x)
print(x2)

x2b <- setPriors(x, "strong")
print(x2b)
```

---

summary.segratioMCMC    *Summary statistics for an segratioMCMC object*

---

**Description**

Wrapper for `summary.mcmc` processing only mixture model parameters although markers may also easily be summarised. The mean, standard deviation, naive standard error of the mean (ignoring autocorrelation of the chain) and time-series standard error based on an estimate of the spectral density at 0. For details see `summary.mcmc`

**Usage**

```
## S3 method for class 'segratioMCMC'
summary(object, ..., row.index = c(1:10),
        var.index = NULL,
        marker.index = c(1:8))
```

**Arguments**

<code>object</code>	object of class <code>segratioMCMC</code>
<code>...</code>	extra options for <code>summary.mcmc</code>
<code>row.index</code>	which rows to print (Default: first 10)
<code>var.index</code>	which mixture model variable to summarise (Default: all)
<code>marker.index</code>	which markers to summarise (Default: 1:8)

**Value**

An object of class `summarySegratioMCMC` is returned which contains summary statistics for parameters and some markers. For details see [summary.mcmc](#)

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[summary.mcmc](#) [mcmc](#) [segratioMCMC](#) [readJags](#) [diagnosticsJagsMix](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
##print(a1)
sr <- segregationRatios(a1$markers)
x <- setModel(3,8)

## Not run:
## fit simple model in one hit and summarise

x.run <- runSegratioMM(sr, x, burn.in=200, sample=500)
print(summary(x.run$mcmc.mixture))
print(summary(x.run$mcmc.mixture, var.index=c(1:3), marker.index=c(1:4)))

## End(Not run)
```

---

writeControlFile	<i>Write JAGS .cmd file for running JAGS</i>
------------------	----------------------------------------------

---

**Description**

Write JAGS .cmd file to disk

**Usage**

```
writeControlFile(jags.control,
  file = paste(jags.control$stem, ".cmd", sep = ""))
```

**Arguments**

jags.control	Object of class <code>jagsControl</code> containing MCMC burn in, sample and thinning as well as relevant files for BUGS commands, inits and data
file	JAGS .cmd file name

**Value**

None.

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[setControl](#) [runJags](#)

**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)
sr <- segregationRatios(a1$markers)

## set up model with 3 components
x <- setModel(3,8)
x2 <- setPriors(x)
dumpData(sr, x)
inits <- setInits(x,x2)
dumpInits(inits)
##x.priors <- setPriors(x, "vague")
writeJagsFile(x, x2, stem="test")

small <- setControl(x, burn.in=20, sample=50)
writeControlFile(small)
```

---

writeJagsFile

*Writes BUGS file for processing by JAGS*


---

**Description**

Given the model and priors a file is written to disk for subsequent JAGS run. BUGS code contained in the model and priors objects is combined and altered if necessary

**Usage**

```
writeJagsFile(model, priors, stem = "test")
```

**Arguments**

model	object of class <code>modelSegratioMM</code> specifying model parameters, ploidy etc
priors	Object of class <code>priorsSegratioMM</code> specifying priors
stem	File name stem for BUGS file (default "test")

**Value**

None.

**Author(s)**

Peter Baker <p.baker1@uq.edu.au>

**See Also**

[segRatio dump](#)



**Examples**

```
## simulate small autooctaploid data set
a1 <- sim.autoMarkers(8,c(0.7,0.2,0.1),n.markers=100,n.individuals=50)

## compute segregation ratios
sr <- segregationRatios(a1$markers)

## set up model for 3 components of autooctoploid
x <- setModel(3,8)
x2 <- setPriors(x)

dumpData(sr, x)
inits <- setInits(x,x2)
dumpInits(inits)
##x.priors <- setPriors(x, "vague")
writeJagsFile(x, x2, stem="test")
```

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