

Package ‘ecodist’

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Title Dissimilarity-based functions for ecological analysis

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Description Dissimilarity-based analysis functions including ordination and Mantel test functions, intended for use with spatial and community data.

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ecodist-package	<i>Dissimilarity-based functions for ecological analysis</i>
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Description

Dissimilarity-based analysis functions including ordination and Mantel test functions with multiple partials, intended for use with spatial and community data.

Details

Package: ecodist
Version: 1.2.2
Date: 2011-04-02
Depends: stats
License: GPL version 2 or newer

Index:

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nmds.min	Find minimum stress configuration
pco	Principal coordinates analysis

plot.mgram	Plot a Mantel correlogram
plot.vf	Plot fitted vectors onto an ordination diagram
pmgram	Partial Mantel correlogram
vf	Vector fitting

Author(s)

Sarah Goslee and Dean Urban

Maintainer: Sarah Goslee <Sarah.Goslee@ars.usda.gov>

References

Goslee, S.C. and Urban, D.L. 2007. The ecodist package for dissimilarity-based analysis of ecological data. *Journal of Statistical Software* 22(7):1-19.

Examples

```
## Distance example using a subset of the built-in iris dataset
data(iris)
iris <- iris[seq(1, 150, by=3),]
iris.md <- distance(iris[,1:4], "mahal")
iris.bc <- bcdist(iris[,1:4])

# compare Mahalanobis and Bray-Curtis dissimilarities
plot(iris.md, iris.bc, xlab="Mahalanobis", ylab="Bray-Curtis")

## NMDS example
# Example of multivariate analysis using built-in iris dataset

# Minimum-stress 2-dimensional nonmetric multidimensional scaling configuration
# Uses only 3 starting configuration to increase speed of example.
# Use more for final analysis to make it more likely that you find the
# global minimum configuration.
iris.nmnds <- nmnds(iris.md, mindim=2, maxdim=2, nits=3)
iris.nmin <- nmnds.min(iris.nmnds)

# Plot NMDS result with symbols denoting species
plot(iris.nmin, pch=as.numeric(iris[,5]))

# Fit vectors for the main variables to the NMDS configuration
# Use more permutations for analysis and publication.
iris.vf <- vf(iris.nmin, iris[,1:4], nperm=50)
plot(iris.vf, col="blue")

## PCO example

iris.pco <- pco(iris.md)
# scatterplot of the first two dimensions
plot(iris.pco$vectors[,1], iris.pco$vectors[,2], pch=as.numeric(iris[,5]))
```

```
## Mantel test example

# Example of multivariate analysis using built-in iris dataset

# Create a model matrix for testing species differences
iris.model <- distance(as.numeric(iris[,5]), "eucl")
iris.model[iris.model > 0] <- 1

# Test whether samples within the same species are more similar than those not
# Use very low numbers of permutations to increase speed of example.
# Use many permutations for final analyses!
mantel(iris.md ~ iris.model, nperm=50, nboot=0)

## Mantel correlogram example

# generate a simple surface
x <- matrix(1:10, nrow=10, ncol=10, byrow=FALSE)
y <- matrix(1:10, nrow=10, ncol=10, byrow=TRUE)
z <- x + 3*y
image(z)

# analyze the pattern of z across space
space <- cbind(as.vector(x), as.vector(y))
z <- as.vector(z)
space.d <- distance(space, "eucl")
z.d <- distance(z, "eucl")
z.mgram <- mgram(z.d, space.d, nperm=0)
plot(z.mgram)

## Partial Mantel correlogram example
# generate a simple surface
x <- matrix(1:10, nrow=10, ncol=10, byrow=FALSE)
y <- matrix(1:10, nrow=10, ncol=10, byrow=TRUE)
z1 <- x + 3*y
z2 <- 2*x - y

# look at patterns
par(mfrow=c(1,2))
image(z1)
image(z2)

# analyze the pattern of z across space
z1 <- as.vector(z1)
z2 <- as.vector(z2)
z1.d <- distance(z1, "eucl")
z2.d <- distance(z2, "eucl")

space <- cbind(as.vector(x), as.vector(y))
space.d <- distance(space, "eucl")
```

```
# take partial correlogram of z2 on the residuals of z1 ~ space.d
z.pmgram <- pmgram(z1.d, space.d, z2.d, nperm=0)
par(mfrow=c(1,1))
plot(z.pmgram, pval=0.1)
```

bcdist	<i>Bray-Curtis distance</i>
--------	-----------------------------

Description

Returns the Bray-Curtis (also known as Sorenson, 1 - percent similarity) pairwise distances for the objects in the data. It has been superseded by `distance()` but remains for backward compatibility.

Usage

```
bcdist(x, rmzero = FALSE)
```

Arguments

<code>x</code>	matrix or data frame with rows as samples and columns as variables (such as species). Distances will be calculated for each pair of rows.
<code>rmzero</code>	If <code>rmzero=TRUE</code> , empty rows will be removed from the data before distances are calculated. Otherwise, the distance between two empty rows is assumed to be 0 (the default).

Value

This function returns a column-order lower-triangular distance matrix. The returned object has an attribute, `Size`, giving the number of objects, that is, `nrow(x)`. The length of the vector that is returned is `nrow(x)*(nrow(x)-1)/2`.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[dist](#)

Examples

```
data(iris)
iris.bc <- bcdist(iris[,1:4])
# equivalent to
iris.bc2 <- distance(iris[,1:4], "bray-curtis")
```

bump *Nine-bump spatial pattern*

Description

A two-dimensional artificial "landscape" illustrating the kind of spatial pattern that might be seen across mountain peaks.

Usage

```
data(bump)
```

Format

The format is: int [1:25, 1:25] 2 2 2 2 2 2 2 2 2 ... - attr(*, "dimnames")=List of 2 ..\$: chr [1:25] "1" "3" "5" "7"\$: chr [1:25] "V1" "V3" "V5" "V7" ...

Examples

```
data(bump)
image(bump)
## maybe str(bump) ; plot(bump) ...
```

cor2m *Generates a correlation table between the variables of 2 matrices*

Description

cor2m generates a correlation table between the variables of 2 matrices—by design, species and environment. It stores these in a table with species as columns and environmental variables as rows, so it's easy to scan. It also removes correlations less than a user-specified alpha (0.05 by default)

Usage

```
cor2m(x, y, trim = TRUE, alpha = 0.05)
```

Arguments

x	A matrix of species (or other) variables
y	A matrix of environmental (or other) variables matching the sites of x
trim	If trim is TRUE, set rho<critical value(alpha) to 0
alpha	alpha p-value to use with trim

Details

Correlate variables from 1 matrix with variables from another matrix. If TRIM, set rho<critical value(alpha) to 0. Computes this critical value as a t-test with n-2 df.

Value

Returns a correlation table between the variables of 2 matrices.

Author(s)

Dean Urban

Examples

```
# toy example
speciesdata <- matrix(runif(50), nrow=10)
colnames(speciesdata) <- c("spA", "spB", "spC", "spD", "spE")
envdata <- matrix(runif(30), nrow=10)
colnames(envdata) <- c("var1", "var2", "var3")
sppenv.cor <- cor2m(speciesdata, envdata)
```

corgen

Generate correlated data

Description

Generate correlated data vectors of a given length.

Usage

```
corgen(len, x, r, population = FALSE, epsilon = 0)
```

Arguments

len	Length of data vectors.
x	Independent data. If x is specified, the population parameter is automatically set to TRUE.
r	Correlation between data vectors.
population	TRUE for vectors drawn from two populations with correlation r, otherwise r is the sample correlation.
epsilon	If epsilon = 0, it has no effect, otherwise the sampling process is repeated until the sample correlation is within epsilon of r. This option allows the production of exactly-correlated data, within the limits of epsilon. Setting epsilon > 0 invalidates the population setting; data will be correlated within that range, rather than sampled from that population.

Details

Either `x` or `len` must be specified. This function has additional capabilities beyond the `corgen` function available in the `stats` package.

Value

`x` First data vector, either generated by `corgen` or given by the user.
`y` Second data vector.
 ...

Author(s)

Sarah Goslee

Examples

```
# create two random variables of length 100 with correlation
# of 0.10 +/- 0.01
xy <- corgen(len=100, r=.1, epsilon=0.01)

# create two random variables of length 100 drawn from a population with
# a correlation of -0.82
xy <- corgen(len=100, r=-0.82, population=TRUE)

# create a variable y within 0.01 of the given correlation to x
x <- 1:100
y <- corgen(x=x, r=.5, epsilon=.01)$y
```

crosstab

Data formatting

Description

Converts field data of the form site, species, observation into a matrix.

Usage

```
crosstab(rowlab, collab, values, type = "sum", data, allrows, allcols, ...)
```

Arguments

`rowlab` row labels, e.g. site names.
`collab` column labels, e.g. species names.
`values` data values.
`data` optional data argument to take `rowlab`, `collab` and/or values from.

type	function to use to combine data, one of "sum" (default), "min", "max", "mean", "count".
allows	optional, list of all desired row names that may not appear in rowlab.
allcols	optional, list of all desired column names that may not appear in collab.
...	optional arguments to the function specified in type, such as na.rm=TRUE

Details

Field data are often recorded as a separate row for each site-species combination. This function turns them into a matrix for further analysis based on unique row and column labels.

The three vectors should all be the same length (including duplicates). The three vectors may also be provided as names of columns in the data frame specified by the data argument.

If allows or allcols exists, rows and/or columns of zeros are inserted for any elements of allrows/allcols not present in rowlab/collab.

If values is missing the number of occurrences of combinations of rowlab and collab will be returned. Thus, crosstab(rowlab, collab) is equivalent to table(rowlab, collab).

If type is "count", the unique combinations of rowlab, collab and values will be returned.

Value

matrix with rowlab as row headings, collab as columns, and values as the data.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

Examples

```
# Make a random example
plotnames <- sort(rep(1:5, 6))
speciesnames <- rep(c("A", "B", "C"), 10)
freqdata <- runif(30)

# number of samples of each species and plot
crosstab(plotnames, speciesnames)

# mean frequency by species and plot
crosstab(plotnames, speciesnames, freqdata, type="mean")
```

distance	<i>Calculate dissimilarity/distance metrics</i>
----------	---

Description

This function calculates a variety of dissimilarity or distance metrics. Although it duplicates the functionality of `dist()` and `bcdist()`, it is written in such a way that new metrics can easily be added. `distance()` was written for extensibility and understandability, and is not an efficient choice for use with large matrices.

Usage

```
distance(x, method = "euclidean", sprange=NULL, spweight=NULL)
```

Arguments

x	matrix or data frame with rows as samples and columns as variables (such as species). Distances will be calculated for each pair of rows.
method	Currently 7 dissimilarity metrics can be calculated: "euclidean", "bray-curtis", "manhattan", "mahalanobis", "jaccard", "difference", "sorensen", "gower", "modgower10" (modified Gower, base 10), "modgower2" (modified Gower, base 2). Partial matching will work for selecting a method.
sprange	Gower dissimilarities offer the option of dividing by the species range. If <code>sprange=NULL</code> no range is used. If <code>sprange</code> is a vector of length <code>nrow(x)</code> it is used for standardizing the dissimilarities.
spweight	Euclidean, Manhattan, and Gower dissimilarities allow weighting. If <code>spweight=NULL</code> , no weighting is used. If <code>spweight="absence"</code> , then $W=0$ if both species are absent and 1 otherwise, thus deleting joint absences.

Value

Returns a lower-triangular distance matrix as an object of class "dist".

Author(s)

Sarah Goslee <Sarah.Goslee@ars.usda.gov>

See Also

[dist](#)

Examples

```
data(iris)
iris.md <- distance(iris[,1:4], "mahal")
```

`fixdmat`*Distance matrix conversion*

Description

Converts a row-order lower-triangular distance matrix to a full symmetric matrix.

Usage

```
fixdmat(v)
```

Arguments

`v` lower-triangular distance matrix in row order.

Details

R and S-Plus distance functions such as `dist` and `bcdist` return a lower triangular distance matrix in column order. Some other programs, such as `EUCLID`, return the lower triangular matrix in row order. To use this matrix in R/S-Plus functions, it must be converted from row order to column order.

Value

Full symmetric distance matrix.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[lower](#), [full](#), [dist](#), [bcdist](#)

`full`*Full symmetric matrix*

Description

Converts a column order distance matrix to a full symmetric matrix.

Usage

```
full(v)
```

Arguments

v lower-triangular column order distance matrix.

Details

Converts lower-triangular distance matrix as written by R/S-Plus functions into a symmetric matrix. Note that lower() used on a 1x1 matrix will return the single element, which may not be the correct behavior in all cases, while full() used on a single element will return a 2x2 matrix.

Value

a full symmetric matrix.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[lower](#)

graze

Site information and grazed vegetation data.

Description

These data contain site location, landscape context and dominant plant species abundances for 50 grazed pastures in the northeastern United States. Elements are the mean values for canopy cover for ten 0.5 x 2 m quadrats.

Usage

```
data(graze)
```

Format

A data frame with 50 observations on the following 67 variables.

sitelocation Site location along a geographic gradient.

forestpct Percentage forest cover within a 1-km radius.

ACMI2 percentage canopy cover

AGGI2 percentage canopy cover

AGROS2 percentage canopy cover

ANAR6 percentage canopy cover

ANOD percentage canopy cover

ARMI2 percentage canopy cover

ASSY percentage canopy cover
BRCA5 percentage canopy cover
BRIN2 percentage canopy cover
CEF02 percentage canopy cover
CIAR4 percentage canopy cover
CIIN percentage canopy cover
CIVU percentage canopy cover
DACA6 percentage canopy cover
DAGL percentage canopy cover
DRYOP percentage canopy cover
ELRE4 percentage canopy cover
ERAN percentage canopy cover
EUGRG percentage canopy cover
FERU2 percentage canopy cover
FRVI percentage canopy cover
GAM0 percentage canopy cover
HIERA percentage canopy cover
HOLA percentage canopy cover
HYPU percentage canopy cover
HYRA3 percentage canopy cover
JUEF percentage canopy cover
LEAU2 percentage canopy cover
LEVU percentage canopy cover
LOAR10 percentage canopy cover
LOC06 percentage canopy cover
LOPE percentage canopy cover
MOVE percentage canopy cover
OXST percentage canopy cover
PEGL2 percentage canopy cover
PHAR3 percentage canopy cover
PHPR3 percentage canopy cover
PLLA percentage canopy cover
PLMA2 percentage canopy cover
POAV percentage canopy cover
POCA17 percentage canopy cover
POLA4 percentage canopy cover
POLYG percentage canopy cover

PON03 percentage canopy cover
POPR percentage canopy cover
POSI2 percentage canopy cover
POTEN percentage canopy cover
PRVU percentage canopy cover
RAAC3 percentage canopy cover
RUBUS percentage canopy cover
RUCR percentage canopy cover
SIOF percentage canopy cover
SOLID percentage canopy cover
SORU2 percentage canopy cover
STGR percentage canopy cover
STME2 percentage canopy cover
SYLA4 percentage canopy cover
TAOF percentage canopy cover
THVU percentage canopy cover
TRAU2 percentage canopy cover
TRHY percentage canopy cover
TRPR2 percentage canopy cover
TRRE3 percentage canopy cover
VEOF2 percentage canopy cover
VICIA percentage canopy cover

Details

Site locations fall along a southwest-northeast transect through the northeastern United States. This is a synthetic gradient calculated from latitude and longitude. Forest landcover is taken from the USGS 1992 National Land Cover Dataset. All forest classes were combined, and the percentage within 1 km of the sample sites was calculated using a GIS. Species codes follow the USDA Plants database. Species with a maximum cover ≤ 1 have been omitted.

Source

Details of these data are available in Tracy and Sanderson (2000) and Goslee, Sanderson and Tracy (ms in review). The 1992 NLCD data can be obtained from <http://www.mrlc.gov/>. Species codes are from <http://plants.usda.gov>.

References

Tracy, B.F. and M.A. Sanderson. 2000. Patterns of plant species richness in pasture lands of the northeast United States. *Plant Ecology* 149:169-180.

Examples

```
data(graze)
```

lower	<i>Lower-triangular matrix</i>
-------	--------------------------------

Description

Converts a symmetric distance matrix to a column order lower triangular matrix.

Usage

```
lower(m)
```

Arguments

`m` a symmetric distance matrix.

Details

Converts a symmetric matrix, for example a dissimilarity matrix, into a lower- triangular matrix. This may be useful to format the input for certain clustering and ordination functions. Note that `lower()` used on a 1x1 matrix will return the single element, which may not be the correct behavior in all cases, while `full()` used on a single element will return a 2x2 matrix.

Value

a column order lower triangular matrix.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[full](#), [fixdmat](#),

mantel	<i>Mantel test</i>
--------	--------------------

Description

Simple and partial Mantel tests, with options for ranked data, permutation tests, and bootstrapped confidence limits.

Usage

```
mantel(formula = formula(data), data = sys.parent(), nperm = 1000,  
       mrank = FALSE, nboot = 500, pboot = 0.9, cboot = 0.95)
```

Arguments

formula	formula in R/S-Plus format describing the test to be conducted. For this test, $y \sim x$ will perform a simple Mantel test, while $y \sim x + z1 + z2 + z3$ will do a partial Mantel test of the relationship between x and y given $z1$, $z2$, $z3$. All variables can be either a distance matrix of class <code>dist</code> or vectors of dissimilarities.
data	an optional dataframe containing the variables in the model as columns of dissimilarities. By default the variables are taken from the current environment.
nperm	number of permutations to use. If set to 0, the permutation test will be omitted.
mrank	if this is set to FALSE (the default option), Pearson correlations will be used. If set to TRUE, the Spearman correlation (correlation ranked distances) will be used.
nboot	number of iterations to use for the bootstrapped confidence limits. If set to 0, the bootstrapping will be omitted.
pboot	the level at which to resample the data for the bootstrapping procedure.
cboot	the level of the confidence limits to estimate.

Details

If only one independent variable is given, the simple Mantel r (r_{12}) is calculated. If more than one independent variable is given, the partial Mantel r ($r_{y|x_1 \dots}$) is calculated by permuting one of the original dissimilarity matrices. The bootstrapping is actually resampling without replacement, because duplication of samples is not useful in a dissimilarity context (the dissimilarity of a sample with itself is zero). Resampling within dissimilarity values is inappropriate, just as for permutation.

Value

mantelr	Mantel coefficient.
pval1	one-tailed p-value (null hypothesis: $r \leq 0$).
pval2	one-tailed p-value (null hypothesis: $r \geq 0$).
pval3	two-tailed p-value (null hypothesis: $r = 0$).
llim	lower confidence limit.
ulim	upper confidence limit.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

References

- Mantel, N. 1967. The detection of disease clustering and a generalized regression approach. *Cancer Research* 27:209-220.
- Smouse, P.E., J.C. Long and R.R. Sokal. 1986. Multiple regression and correlation extensions of the Mantel test of matrix correspondence. *Systematic Zoology* 35:627-632.

See Also[mgram](#)**Examples**

```
## Not run:
# Example of multivariate analysis using built-in iris dataset
data(iris)
iris.md <- distance(iris[,1:4], "mahal")

# Create a model matrix for testing species differences
iris.model <- distance(as.numeric(iris[,5]), "eucl")
iris.model[iris.model > 0] <- 1

# Test whether samples within the same species are more similar than those not
mantel(iris.md ~ iris.model, nperm=10000)

## End(Not run)
```

mgram

*Mantel correlogram***Description**

Calculates simple and partial Mantel correlograms.

Usage

```
mgram(species.d, space.d, breaks, nclass, stepsize, nperm = 1000,
      mrank = FALSE, nboot = 500, pboot = 0.9, cboot = 0.95,
      alternative = "two.sided", trace = FALSE)
```

Arguments

species.d	lower-triangular dissimilarity matrix.
space.d	lower-triangular matrix of geographic distances.
breaks	locations of class breaks. If specified, overrides nclass and stepsize.
nclass	number of distance classes. If not specified, Sturge's rule will be used to determine an appropriate number of classes.
stepsize	width of each distance class. If not specified, nclass and the range of space.d will be used to calculate an appropriate default.
nperm	number of permutations to use. If set to 0, the permutation test will be omitted.
mrnk	if this is set to F (the default option), Pearson correlations will be used. If set to T, the Spearman correlation (correlation ranked distances) will be used.
nboot	number of iterations to use for the bootstrapped confidence limits. If set to 0, the bootstrapping will be omitted.

pboot	the level at which to resample the data for the bootstrapping procedure.
cboot	the level of the confidence limits to estimate.
alternative	default is "two.sided", and returns p-values for $H_0: r_M = 0$. The alternative is "one.sided", which returns p-values for $H_0: r_M \leq 0$.
trace	if TRUE, returns progress indicators.

Details

This function calculates Mantel correlograms. The Mantel correlogram is essentially a multivariate autocorrelation function. The Mantel r represents the dissimilarity in variable composition (often species composition) at a particular lag distance.

Value

Returns an object of class `mgram`, which is a list with two elements. `mgram` is a matrix with one row for each distance class and 6 columns:

lag	midpoint of the distance class.
ngroup	number of distances in that class.
mantelr	Mantel r value.
pval	p-value for the test chosen.
llim	lower bound of confidence limit for mantelr.
ulim	upper bound of confidence limit for mantelr.

resids is NA for objects calculated by `mgram()`.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

References

Legendre, P. and M. Fortin. 1989. Spatial pattern and ecological analysis. *Vegetatio* 80:107-138.

See Also

[mantel](#), [plot.mgram](#), [pmgram](#)

Examples

```
## Not run:

# generate a simple surface
x <- matrix(1:10, nrow=10, ncol=10, byrow=FALSE)
y <- matrix(1:10, nrow=10, ncol=10, byrow=TRUE)
z <- x + 3*y
image(z)

# analyze the pattern of z across space
```

```

space <- cbind(as.vector(x), as.vector(y))
z <- as.vector(z)
space.d <- distance(space, "eucl")
z.d <- distance(z, "eucl")
z.mgram <- mgram(z.d, space.d, nperm=0)
plot(z.mgram)

## End(Not run)

```

MRM

Multiple Regression on distance Matrices

Description

Multiple regression on distance matrices (MRM) using permutation tests of significance for regression coefficients and R-squared.

Usage

```
MRM(formula = formula(data), data = sys.parent(), nperm = 1000, mrank = FALSE)
```

Arguments

formula	formula in R/S-Plus format describing the test to be conducted.
data	an optional dataframe containing the variables in the model as columns of dissimilarities. By default the variables are taken from the current environment.
nperm	number of permutations to use. If set to 0, the permutation test will be omitted.
mrnk	if this is set to FALSE (the default option), Pearson correlations will be used. If set to TRUE, the Spearman correlation (correlation ranked distances) will be used.

Details

Performs multiple regression on distance matrices following the methods outlined in Legendre et al. 1994.

Value

coef	A matrix with regression coefficients and associated p-values from the permutation test (using the pseudo-t of Legendre et al. 1994).
r.squared	Regression R-squared and associated p-value from the permutation test.
F.test	F-statistic and p-value for overall F-test for lack of fit.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

References

Lichstein, J. 2007. Multiple regression on distance matrices: A multivariate spatial analysis tool. *Plant Ecology* 188: 117-131.

Legendre, P.; Lapointe, F. and Casgrain, P. 1994. Modeling brain evolution from behavior: A permutational regression approach. *Evolution* 48: 1487-1499.

See Also

[mantel](#)

Examples

```
data(graze)
LOAR10.mrm <- MRM(dist(LOAR10) ~ dist(sitelocation) + dist(forestpct), data=graze, nperm=100)
```

nmds

Non-metric multidimensional scaling

Description

Non-metric multidimensional scaling.

Usage

```
nmds(dmat, mindim = 1, maxdim = 2, nits = 10, iconf = 0, epsilon = 1e-12,
     maxit = 500, trace = FALSE, stresscalc="default")
```

Arguments

dmat	lower-triangular dissimilarity matrix.
mindim	optional, minimum number of dimensions to use.
maxdim	optional, maximum number of dimensions to use.
nits	optional, number of separate ordinations to use.
iconf	optional, initial configuration. If not specified, then a random configuration is used.
epsilon	optional, acceptable difference in stress.
maxit	optional, maximum number of iterations.
trace	if TRUE, will write progress indicator to the screen.
stresscalc	if "default" uses modified stress calculation; if "kruskal" uses unmodified Kruskal stress calculation.

Details

The goal of NMDS is to find a configuration in a given number of dimensions which preserves rank-order dissimilarities as closely as possible. The number of dimensions must be specified in advance. Because NMDS is prone to finding local minima, several random starts must be used. Stress is used as the measure of goodness of fit. A lower stress indicates a better match between dissimilarity and ordination.

Value

conf	list of configurations.
stress	list of final stress values.
r2	total variance explained by each configuration.

The first results are for the lowest number of dimensions (total number is (mindim - maxdim + 1) * nits).

Author(s)

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References

Kruskal, J.B. 1964. Multidimensional scaling by optimizing goodness of fit to a nonmetric hypothesis. *Psychometrika* 29:1-27.

Minchin, P.R. 1987. An evaluation of the relative robustness of techniques for ecological ordination. *Vegetatio* 96:89-108.

See Also

[nmds.min](#), [vf](#)

Examples

```
## Not run:
# Example of multivariate analysis using built-in iris dataset
data(iris)
iris <- iris[seq(1, 150, by=3),]
iris.md <- distance(iris[,1:4], "mahal")

# Minimum-stress 2-dimensional nonmetric multidimensional scaling configuration
# Uses small number of separate ordinations (5) to increase speed of example.
# Use more for final analysis.
iris.nmds <- nmds(iris.md, mindim=2, maxdim=2, nits=3)
iris.nmin <- nmds.min(iris.nmds)

# Plot NMDS result with symbols denoting species
plot(iris.nmin, pch=as.numeric(iris[,5]))

# Fit vectors for the main variables to the NMDS configuration
```

```
iris.vf <- vf(iris.nmin, iris[,1:4], nperm=10)
plot(iris.vf, col="blue")

## End(Not run)
```

nmds.min

Find minimum stress configuration

Description

Finds minimum stress configuration from output of nmds()

Usage

```
nmds.min(x, dims = 0)
```

Arguments

x	output from nmds()
dims	desired dimensionality of result. If dims == 0 (the default) then the overall minimum stress configuration is chosen.

Value

Configuration of minimum stress ordination (dataframe of coordinates). The stress and r² for the minimum stress configuration are stored as attributes.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[nmds](#)

Examples

```
## Not run:
# Example of multivariate analysis using built-in iris dataset
data(iris)
iris <- iris[seq(1, 150, by=3),]
iris.md <- distance(iris[,1:4], "mahal")

# Minimum-stress 2-dimensional nonmetric multidimensional scaling configuration
# Uses small number of separate ordinations (5) to increase speed of example.
# Use more for final analysis.
iris.nmds <- nmds(iris.md, mindim=2, maxdim=2, nits=3)
iris.nmin <- nmds.min(iris.nmds)
```

```
# Plot NMDS result with symbols denoting species
plot(iris.nmin, pch=as.numeric(iris[,5]))

# Fit vectors for the main variables to the NMDS configuration
iris.vf <- vf(iris.nmin, iris[,1:4], nperm=10)
plot(iris.vf, col="blue")

## End(Not run)
```

pco

Principal coordinates analysis

Description

Principal coordinates analysis (classical scaling).

Usage

```
pco(x, negvals = "zero", dround = 0)
```

Arguments

x	a lower-triangular dissimilarity matrix.
negvals	if = "zero" sets all negative eigenvalues to zero; if = "rm" corrects for negative eigenvalues using method 1 of Legendre and Anderson 1999.
dround	if greater than 0, attempts to correct for round-off error by rounding to that number of places.

Details

PCO (classical scaling, metric multidimensional scaling) is very similar to principal components analysis, but allows the use of any dissimilarity metric.

Value

values	eigenvalue for each component. This is a measure of the variance explained by each dimension.
vectors	eigenvectors. Each column contains the scores for that dimension.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[princomp](#), [nmds](#)

Examples

```
## Not run:
data(iris)
iris.md <- distance(iris[,1:4], "mahal")
iris.pco <- pco(iris.md)

# scatterplot of the first two dimensions
plot(iris.pco$vectors[,1], iris.pco$vectors[,2], pch=as.numeric(iris[,5]))

## End(Not run)
```

permresults	<i>Mantel test results for number of permutations, correlation and dataset size.</i>
-------------	--

Description

These results explore the variability in randomization tests of significance and in bootstrap confidence limits for different sample sizes, strengths of relationship and numbers of permutations.

Usage

```
data(permresults)
```

Format

A data frame with 15000 observations on the following 17 variables.

- n The number of samples.
- r The Mantel r of the dissimilarity vectors.
- p500 The p-values generated using 500 permutations.
- l100 The lower confidence limit generated using 100 subsamples.
- u100 The upper confidence limit generated using 100 subsamples.
- p1000 p-values with 1,000 permutations
- l500 lower CI with 500 permutations
- u500 upper CI with 500 permutations
- p10000 p-values with 10,000 permutations
- l1000 lower CI with 1,000 permutations
- u1000 upper CI with 1,000 permutations
- p100000 p-values with 100,000 permutations
- l10000 lower CI with 10,000 permutations
- u10000 upper CI with 10,000 permutations
- p1000000 p-values with 1,000,000 permutations
- l100000 lower CI with 100,000 permutations
- u100000 upper CI with 100,000 permutations

Details

These simulation results are included here because the entire simulation takes an extremely long time to run. The tests explored how the p-value and confidence limits varied with:

- number of samples (10 (45 dissimilarities), 25 (300), 50 (1225), 100 (4950))
- number of permutations: 500, 1000, 10000, 100000, 1000000
- number of bootstrap samples: 100, 500, 1000, 10000, 100000
- correlation of the original data: 0.01, 0.05, 0.10, 0.25, 0.50

The larger correlation values were omitted for the larger sample sizes because they were always highly significant, and an additional low value of 0.025 was added for the largest sample size.

The method used to generate the data is shown in the example section.

Examples

```
data(permresults)

## Not run:
# Simulation Method
# example for sample size 10, correlation of 0.01

permresults <- data.frame(matrix(0, nrow=1000, ncol=17))
colnames(permresults) <- c("n", "r", "p500", "l100", "u100", "p1000",
  "l500", "u500", "p10000", "l1000", "u1000", "p100000", "l10000", "u10000",
  "p1000000", "l100000", "u100000")

# generate correlated data
corxy.len010.r010 <- corgen(len= 10, r= 0.01, epsilon=0.0001)

all.nperm <- c(500, 1000, 10000, 100000, 1000000)
all.nboot <- c(100, 500, 1000, 10000, 100000)

for(i in 1:1000) {
  permresults[i, 1] <- 10
  permresults[i, 2] <- 0.01
  for(j in 1:5) {
    # record p-value, lower and upper confidence limits
    thismantel <- mantel(corxy.len010.r010$y ~ corxy.len010.r010$x,
      nperm=all.nperm[j], nboot=all.nboot[j])
    permresults[i, (3*j):(3*j + 2)] <- c(thismantel[c(2,5,6)])
  }
}

## End(Not run)
```

`plot.mgram`*Plot a Mantel correlogram*

Description

Plot a Mantel correlogram from an object of S3 class `mgram`, using solid symbols for significant values.

Usage

```
## S3 method for class 'mgram'  
plot(x, pval = 0.05, xlab = "Distance", ylab = "Mantel r", ...)
```

Arguments

<code>x</code>	an object of class <code>mgram</code>
<code>pval</code>	cut-off level for statistical significance.
<code>xlab</code>	x-axis label.
<code>ylab</code>	y-axis label.
<code>...</code>	optional, any additional graphics parameters.

Value

draws a plot (graphics device must be active).

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[mgram](#)

Examples

```
## Not run:  
# generate a simple surface  
x <- matrix(1:10, nrow=10, ncol=10, byrow=FALSE)  
y <- matrix(1:10, nrow=10, ncol=10, byrow=TRUE)  
z <- x + 3*y  
image(z)  
  
# analyze the pattern of z across space  
space <- cbind(as.vector(x), as.vector(y))  
z <- as.vector(z)  
space.d <- distance(space, "eucl")  
z.d <- distance(z, "eucl")  
z.mgram <- mgram(z.d, space.d, nperm=0)
```

```
plot(z.mgram)

## End(Not run)
```

plot.vf *Plots fitted vectors onto an ordination diagram*

Description

Add vector fitting arrows to an existing ordination plot.

Usage

```
## S3 method for class 'vf'
plot(x, pval = 1, cex = 0.8, ascale = 0.9, ...)
```

Arguments

x	an object of S3 class vf, created by vf()
pval	optional, critical p-value for choosing variables to plot
cex	text size
ascale	optional, proportion of plot area to use when calculating arrow length
...	optional, other graphics parameters

Value

Adds arrows to an existing ordination plot. Only arrows with a p-value less than pval are added. By default, all variables are shown.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[vf](#)

Examples

```
## Not run:
# Example of multivariate analysis using built-in iris dataset
data(iris)
iris <- iris[seq(1, 150, by=3),]
iris.md <- distance(iris[,1:4], "mahal")

# Minimum-stress 2-dimensional nonmetric multidimensional scaling configuration
# Uses small number of separate ordinations (5) to increase speed of example.
```

```

# Use more for final analysis.
iris.nmnds <- nmnds(iris.md, mindim=2, maxdim=2, nits=3)
iris.nmin <- nmnds.min(iris.nmnds)

# Plot NMDS result with symbols denoting species
plot(iris.nmin, pch=as.numeric(iris[,5]))

# Fit vectors for the main variables to the NMDS configuration
iris.vf <- vf(iris.nmin, iris[,1:4], nperm=10)
plot(iris.vf, col="blue")

## End(Not run)

```

pmgram

Partial Mantel correlogram

Description

This function calculates simple and partial multivariate correlograms.

Usage

```
pmgram(data, space, partial, breaks, nclass, stepsize, resids = FALSE, nperm = 1000)
```

Arguments

data	lower-triangular dissimilarity matrix. This can be either an object of class <code>dist</code> (treated as one column) or a matrix or data frame with one or two columns, each of which is an independent lower-triangular dissimilarity in vector form.
space	lower-triangular matrix of geographic distances.
partial	optional, lower-triangular dissimilarity matrix of ancillary data.
breaks	locations of class breaks. If specified, overrides <code>nclass</code> and <code>stepsize</code> .
nclass	number of distance classes. If not specified, Sturge's rule will be used to determine an appropriate number of classes.
stepsize	width of each distance class. If not specified, <code>nclass</code> and the range of <code>space.d</code> will be used to calculate an appropriate default.
resids	if <code>resids=TRUE</code> , will return the residuals for each distance class. Otherwise returns 0.
nperm	number of permutations to use. If set to 0, the permutation test will be omitted.

Details

This function does four different analyses: If data has 1 column and partial is missing, calculates a multivariate correlogram for data.

If data has 2 columns and partial is missing, calculates Mantel r between the two columns for each distance class.

If data has 1 column and partial exists, does a multivariate correlogram for the residuals, taking residuals over whole extent.

If data has 2 columns and partial exists, does a partial multivariate correlogram, calculating partial for each distance class separately.

Value

Returns a object of class mgram, which is a list containing two objects: mgram is a matrix with one row for each distance class and 4 columns:

lag	midpoint of the distance class.
ngroup	number of distances in that class.
piecer	Mantel r value.
pval	two-sided p-value.

resids is a vector of the residuals (if calculated) and can be accessed with the residuals() method.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[mgram](#), [mantel](#)

Examples

```
## Not run:
# generate a simple surface
x <- matrix(1:10, nrow=10, ncol=10, byrow=FALSE)
y <- matrix(1:10, nrow=10, ncol=10, byrow=TRUE)
z1 <- x + 3*y
z2 <- 2*x - y

# look at patterns
par(mfrow=c(1,2))
image(z1)
image(z2)

# analyze the pattern of z across space
z1 <- as.vector(z1)
z2 <- as.vector(z2)
z1.d <- distance(z1, "eucl")
```

```
z2.d <- distance(z2, "eucl")

space <- cbind(as.vector(x), as.vector(y))
space.d <- distance(space, "eucl")

# take partial correlogram of z2 on the residuals of z1 ~ space.d
z.pmgram <- pmgram(z1.d, space.d, z2.d, nperm=0)
par(mfrow=c(1,1))
plot(z.pmgram, pval=0.1)

## End(Not run)
```

residuals.mgram

Residuals of a Mantel correlogram

Description

Extracts residuals from an S3 object of class `mgram` (only relevant for objects created by `pmgram{ }`).

Usage

```
## S3 method for class 'mgram'
residuals(object, ...)
```

Arguments

<code>object</code>	an object of class <code>mgram</code>
<code>...</code>	additional arguments

Value

vector of residuals.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

See Also

[mgram](#)

Examples

```
## Not run:
z.pmgram <- pmgram(z.d, space.d, resids=TRUE)
residuals(z.pmgram)

## End(Not run)

# A full example is available in the Mantel correlogram
# section of the main help file for \link{ecodist}.
```

vf

Vector fitting

Description

Fits environmental variables to an ordination configuration.

Usage

```
vf(ord, vars, nperm = 100)
```

Arguments

ord	matrix containing a 2-dimensional ordination result with axes as columns.
vars	matrix with ancillary variables as columns.
nperm	number of permutation for the significance test. If nperm = 0, the test will be omitted.

Details

Vector fitting finds the maximum correlation of the individual variables with a configuration of samples in ordination space.

Value

an object of class `vf` containing matrix with the first 2 columns containing the scores for every variable in each of the 2 dimensions of the ordination space. `r` is the maximum correlation of the variable with the ordination space, and `pval` is the result of the permutation test.

Author(s)

Sarah Goslee, Sarah.Goslee@ars.usda.gov

References

Jongman, R.H.G., C.J.F. ter Braak and O.F.R. van Tongeren. 1995. Data analysis in community and landscape ecology. Cambridge University Press, New York.

See Also[plot.vf](#)**Examples**

```
## Not run:
# Example of multivariate analysis using built-in iris dataset
data(iris)
iris <- iris[seq(1, 150, by=3),]
iris.md <- distance(iris[,1:4], "mahal")

# Minimum-stress 2-dimensional nonmetric multidimensional scaling configuration
# Uses small number of separate ordinations (5) to increase speed of example.
# Use more for final analysis.
iris.nmds <- nmds(iris.md, mindim=2, maxdim=2, nits=3)
iris.nmin <- nmds.min(iris.nmds)

# Plot NMDS result with symbols denoting species
plot(iris.nmin, pch=as.numeric(iris[,5]))

# Fit vectors for the main variables to the NMDS configuration
iris.vf <- vf(iris.nmin, iris[,1:4], nperm=10)
plot(iris.vf, col="blue")

## End(Not run)
```


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