

Package ‘ade4’

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License GPL (>= 2)

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ade4-package

The ade4 package

Description

This package is developed in the Biometry and Evolutionary Biology Lab (UMR 5558) - University Lyon 1. It contains Data Analysis functions to analyse Ecological and Environmental data in the framework of Euclidean Exploratory methods, hence the name ade4.

ade4 is characterized by (1) the implementation of graphical and statistical functions, (2) the availability of numerical data, (3) the redaction of technical and thematic documentation and (4) the inclusion of bibliographic references.

To cite ade4, please use `citation("ade4")`.

Author(s)

Daniel Chessel, Anne B Dufour, Stephane Dray with the contributions from J.R. Lobry, S. Ollier, S. Pavoine and J. Thioulouse

References

See ade4 website: <http://pbil.univ-lyon1.fr/ADE-4/>

See Also

ade4TkGUI, adegenet, adehabitat

abouheif.eg

Phylogenies and quantitative traits from Abouheif

Description

This data set gathers three phylogenies with three sets of traits as reported by Abouheif (1999).

Usage

```
data(abouheif.eg)
```


Format

`abouheif.eg` is a list containing the 6 following objects :

tre1 is a character string giving the first phylogenetic tree made up of 8 leaves.

vec1 is a numeric vector with 8 values.

tre2 is a character string giving the second phylogenetic tree made up of 7 leaves.

vec2 is a numeric vector with 7 values.

tre3 is a character string giving the third phylogenetic tree made up of 15 leaves.

vec3 is a numeric vector with 15 values.

Source

Data taken from the phylogenetic independence program developed by Ehab Abouheif starting from <http://ww2.mcgill.ca/biology/faculty/abouheif/programs.html>.

References

Abouheif, E. (1999) A method for testing the assumption of phylogenetic independence in comparative data. *Evolutionary Ecology Research*, **1**, 895–909.

Examples

```
data(abouheif.eg)
par(mfrow=c(2,2))
symbols.phylog(newick2phylog(abouheif.eg$tre1), abouheif.eg$vec1,
  sub = "Body Mass (kg)", csi = 2, csub = 2)
symbols.phylog(newick2phylog(abouheif.eg$tre2), abouheif.eg$vec2,
  sub = "Body Mass (kg)", csi = 2, csub = 2)
dotchart.phylog(newick2phylog(abouheif.eg$tre1), abouheif.eg$vec1,
  sub = "Body Mass (kg)", cdot = 2, cnod = 1, possub = "topleft",
  csub = 2, ceti = 1.5)
dotchart.phylog(newick2phylog(abouheif.eg$tre2), abouheif.eg$vec2,
  sub = "Body Mass (kg)", cdot = 2, cnod = 1, possub = "topleft",
  csub = 2, ceti = 1.5)
par(mfrow = c(1,1))

w.phy=newick2phylog(abouheif.eg$tre3)
dotchart.phylog(w.phy, abouheif.eg$vec3, clabel.n = 1)
```

Description

Counts of individuals of *Acacia ehrenbergiana* from five parallel transects of 32 quadrats.

Usage

```
data(acacia)
```

Format

acacia is a data frame with 15 variables :

se.T1, se.T2, se.T3, se.T4, se.T5 are five numeric vectors containing quadrats counts of seedlings from transects 1 to 5 respectively;

sm.T1, sm.T2, sm.T3, sm.T4, sm.T5 are five numeric vectors containing quadrats counts of small trees (crown < 1 m² in canopy) of transects 1 to 5 respectively;

la.T1, la.T2, la.T3, la.T4, la.T5 are five numeric vectors containing quadrats counts of trees with large crown (crown > 1 m² in canopy) of transects 1 to 5 respectively.

Source

Greig-Smith, P. and Chadwick, M.J. (1965) Data on pattern within plant communities. III. *Acacia-Capparis* semi-desert scrub in the Sudan. *Journal of Ecology*, **53**, 465–474.

References

Hill, M.O. (1973) The intensity of spatial pattern in plant communities. *Journal of Ecology*, **61**, 225–235.

Examples

```
data(acacia)
par(mfcol = c(5,3))
par(mar = c(2,2,2,2))
for(k in 1:15) {
  barplot(acacia[,k], ylim = c(0,20), col = grey(0.8))
  ade4:::scatterutil.sub(names(acacia)[k], 1.5, "topleft")
}
par(mfcol = c(1,1))
```

```
add.scatter
```

Add graphics to an existing plot

Description

add.scatter is a function which defines a new plot area within an existing plot and displays an additional graphic inside this area. The additional graphic is determined by a function which is the first argument taken by add.scatter. It can be used in various ways, for instance to add a screeplot to an ordination scatterplot (add.scatter.eig).

The function add.scatter.eig uses the following colors: black (represented axes), grey (axes retained in the analysis) and white (others).

Usage

```
add.scatter(func, posi = c("bottomleft", "bottomright", "topleft", "topright"),
            ratio = 0.2, inset = 0.01, bg.col = 'white')
add.scatter.eig(w, nf = NULL, xax, yax, posi = "bottomleft", ratio =
               .25, inset = 0.01, sub = "Eigenvalues", csub = 2 * ratio)
```

Arguments

func	an - evaluated - function producing a graphic
posi	a character vector (only its first element being considered) giving the position of the added graph. Possible values are "bottomleft" (= "bottom"), "bottomright", "topleft" (= "top"), "topright", and "none" (no plot).
ratio	the size of the added graph in proportion of the current plot region
inset	the inset from which the graph is drawn, in proportion of the whole plot region. Can be a vector of length 2, giving the inset in x and y. If atomic, same inset is used in x and y
bg.col	the color of the background of the added graph
w	numeric vector of eigenvalues
nf	the number of retained factors, NULL if not provided
xax	first represented axis
yax	second represented axis
sub	title of the screeplot
csub	size of the screeplot title

Details

add.scatter uses `par("plt")` to redefine the new plot region. As stated in `par` documentation, this produces to (sometimes surprising) interactions with other parameters such as "mar". In particular, such interactions are likely to reset the plot region by default which would cause the additional graphic to take the whole plot region. To avoid such inconvenient, add `par([other options], plt=par("plt"))` when using `par` in your graphical function (argument `func`).

Value

The matched call (invisible).

Author(s)

Thibaut Jombart <jombart@biomserv.univ-lyon1.fr>

See Also

[scatter](#)

Examples

```

par(mfrow=c(2,2))

f1 <- function(a){
  opar=par("mar","xaxt","yaxt","plt")
  on.exit(par(opar))
  par(mar=rep(.1,4),xaxt="n",yaxt="n",plt=par("plt"))

  hist(a,xlab="",ylab="",main="",col="white",proba=TRUE)
  lines(seq(-4,4,le=50),dnorm(seq(-4,4,le=50)),col="red")
}

a <- rnorm(100)
barplot(sort(a))
add.scatter(f1(a),posi="topleft",bg.col="grey")

a <- rnorm(100)
barplot(sort(a))
add.scatter(f1(a),posi="topleft",bg.col="grey",inset=c(.25,.01))

a <- rnorm(100)
barplot(sort(a))
add.scatter(f1(a),posi="topleft",bg.col="grey",inset=.25,ratio=.1)

a <- rnorm(100)
barplot(sort(a))
add.scatter(f1(a),posi="bottomright",bg.col="grey",ratio=.3)

par(mfrow=c(1,1))

data(microsatt)
w <- dudi.coa(data.frame(t(microsatt$tab)), scann = FALSE,nf=3)
s.label(w$co)
ade4:::add.scatter.eig(w$eig,w$nf,posi="bottomright",1,2)

```

ade4toR

Format Change Utility

Description

Exchange objects between R and ADE4 (classic version)
<http://pbil.univ-lyon1.fr/ADE-4>

Usage

```

ade4toR (fictab, ficcolnames = NULL, ficrownames = NULL)
Rtoade4 (x)

```

Arguments

fictab	a name of ADE4 text file. A data frame with the same name is created in the R environment.
ficcolnames	the column names label file
ficrownames	the row names label file
x	a data frame

Details

"xxx" is the name of object x ((deparse(substitute(x))))

For any table :

creates a file "xxx.txt"

creates a file "xxx_row_lab.txt" with row names

creates a file "xxx_col_lab.txt" with column names

if x has the 'col.blocks' attribute

creates a file "xxx_col_bloc_lab.txt" with blocks names

creates a file "xxx_col_bloc.txt" with blocks sizes

For a table which all columns are factors :

creates a file "xxx.txt"

creates a file "xxx_var_lab.txt" with row names

creates a file "xxx_moda_lab.txt" with categories names

Files are created in the current working directory.

Value

'ade4toR' gives data frames.

'Rtoade4' gives text files.

Examples

```
## Not run: data(tarentaise)
traits <- tarentaise$traits
ade4:::Rtoade4(traits)
# File creation traits.txt
# File creation traits_col_lab.txt
# File creation traits_row_lab.txt
# File creation traits_col_bloc.txt
# File creation traits_col_bloc_lab.txt

mil <- tarentaise$envir
ade4:::Rtoade4(mil)
#File creation mil.txt
#File creation mil_var_lab.txt
#File creation mil_moda_lab.txt

## End(Not run)
```

`aminoacyl`*Codon usage*

Description

`aminoacyl` is a list containing the codon counts of 36 genes encoding yeast aminoacyl-tRNA-synthetase(*S.Cerevisiae*).

Usage

```
data(aminoacyl)
```

Format

`aminoacyl` is a list containing the 5 following objects:

genes is a vector giving the gene names.

localisation is a vector giving the cellular localisation of the proteins (M = mitochondrial, C = cytoplasmic, I = indetermined, CI = cyto and mito).

codon is a vector containing the 64 triplets.

AA is a factor giving the amino acid names for each codon.

usage.codon is a dataframe containing the codon counts for each gene.

Source

Data prepared by D. Charif <charif@biomserv.univ-lyon1.fr> starting from:
<http://www.expasy.org/sprot/>

References

Chiapello H., Olivier E., Landes-Devauchelle C., Nitschké P. and Risler J.L (1999) Codon usage as a tool to predict the cellular localisation of eukariotic ribosomal proteins and aminoacyl-tRNA synthetases. *Nucleic Acids Res.*, **27**, 14, 2848–2851.

Examples

```
data(aminoacyl)
aminoacyl$genes
aminoacyl$usage.codon
dudi.coa(aminoacyl$usage.codon, scannf = FALSE)
```

amova

Analysis of molecular variance

Description

The analysis of molecular variance tests the differences among population and/or groups of populations in a way similar to ANOVA. It includes evolutionary distances among alleles.

Usage

```
amova(samples, distances, structures)
## S3 method for class 'amova'
print(x, full = FALSE, ...)
```

Arguments

samples	a data frame with haplotypes (or genotypes) as rows, populations as columns and abundance as entries
distances	an object of class <code>dist</code> computed from Euclidean distance. If <code>distances</code> is null, equidistances are used.
structures	a data frame containing, in the <i>j</i> th row and the <i>k</i> th column, the name of the group of level <i>k</i> to which the <i>j</i> th population belongs
x	an object of class <code>amova</code>
full	a logical value indicating whether the original data ('distances', 'samples', 'structures') should be printed
...	further arguments passed to or from other methods

Value

Returns a list of class `amova`

call	call
results	a data frame with the degrees of freedom, the sums of squares, and the mean squares. Rows represent levels of variability.
componentsofcovariance	a data frame containing the components of covariance and their contribution to the total covariance
statphi	a data frame containing the phi-statistics

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Excoffier, L., Smouse, P.E. and Quattro, J.M. (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics*, **131**, 479–491.

See Also

[randtest.amova](#)

Examples

```
data(humDNAm)
amovahum <- amova(humDNAm$samples, sqrt(humDNAm$distances), humDNAm$structures)
amovahum
```

apis108	<i>Allelic frequencies in ten honeybees populations at eight microsatellites loci</i>
---------	---

Description

This data set gives the occurrences for the allelic form on 8 loci in 10 populations of honeybees.

Usage

```
data(apis108)
```

Format

A data frame containing 180 rows (allelic forms on 8 loci) and 10 columns (populations of honeybees : El.Hermel, Al.Hoceima, Nimba, Celinda, Pretoria, Chalkidiki, Forli, Valenciennes, Umea and Seville).

Source

<http://www.montpellier.inra.fr/URLB/apis/libanfreq.pdf>

Franck P., Garnery L., Solignac M. and Cornuet J.M. (2000) Molecular confirmation of a fourth lineage in honeybees from the Near-East. *Apidologie*, **31**, 167–180.

Examples

```
data(apis108)
apis <- count2genet(as.data.frame(t(apis108)))
apis.pca <- dudi.pca(apis$tab, center = apis$center,
  scale = FALSE, scannf = FALSE, nf = 3)
```

apqe *Apportionment of Quadratic Entropy*

Description

The hierarchical apportionment of quadratic entropy defined by Rao (1982).

Usage

```
apqe(samples, dis = NULL, structures)
## S3 method for class 'apqe'
print(x, full = FALSE, ...)
```

Arguments

samples	a data frame with haplotypes (or genotypes) as rows, populations as columns and abundance or presence-absence as entries
dis	an object of class <code>dist</code> computed from Euclidean distance. If <code>dis</code> is null, equidistances are used.
structures	a data frame that contains, in the <code>j</code> th row and the <code>k</code> th column, the name of the group of level <code>k</code> to which the <code>j</code> th population belongs
x	an object of class <code>apqe</code>
full	a logical value that indicates whether the original data ('distances', 'samples', 'structures') should be printed
...	... further arguments passed to or from other methods

Value

Returns a list of class `apqe`

call	call
results	a data frame that contains the components of diversity.

Author(s)

Sandrine Pavoine <pavoine@mnhn.fr>

References

Rao, C.R. (1982) Diversity: its measurement, decomposition, apportionment and analysis. *Sankhya: The Indian Journal of Statistics*, **A44**, 1–22.

Pavoine S. and Dolédec S. (2005) The apportionment of quadratic entropy: a useful alternative for partitioning diversity in ecological data. *Environmental and Ecological Statistics*, **12**, 125–138.

Examples

```
data(ecomor)
ecomor.phylog <- taxo2phylog(ecomor$taxo)
apqe(ecomor$habitat, ecomor.phylog$Wdist)
```

aravo

Distribution of Alpine plants in Aravo (Valloire, France)

Description

This dataset describe the distribution of 82 species of Alpine plants in 75 sites. Species traits and environmental variables are also measured.

Usage

```
data(aravo)
```

Format

aravo is a list containing the following objects :

spe is a data.frame with the abundance values of 82 species (columns) in 75 sites (rows).

env is a data.frame with the measurements of 6 environmental variables for the sites.

traits is data.frame with the measurements of 8 traits for the species.

spe.names is a vector with full species names.

Details

The environmental variables are:

Aspect	Relative south aspect (opposite of the sine of aspect with flat coded 0)	/
Slope	Slope inclination (degrees)	/
Form	Microtopographic landform index	1 (convexity); 2 (convex slope)
Snow	Mean snowmelt date (Julian day) averaged over 1997-1999	/
PhysD	Physical disturbance, i.e., percentage of unvegetated soil due to physical processes	/
ZoogD	Zoogenic disturbance, i.e., quantity of unvegetated soil due to marmot activity	no; some; high

The species traits for the plants are:

Height	Vegetative height (cm)
Spread	Maximum lateral spread of clonal plants (cm)
Angle	Leaf elevation angle estimated at the middle of the lamina
Area	Area of a single leaf
Thick	Maximum thickness of a leaf cross section (avoiding the midrib)
SLA	Specific leaf area
Nmass	Mass-based leaf nitrogen content
Seed	Seed mass

Source

Choler, P. (2005) Consistent shifts in Alpine plant traits along a mesotopographical gradient. *Arctic, Antarctic, and Alpine Research*, **37**,444–453.

Examples

```
data(aravo)
coa1 <- dudi.coa(aravo$spe, scannf = FALSE, nf = 2)
dudienv <- dudi.hillsmith(aravo$env, scannf = FALSE, nf = 2, row.w = coa1$lw)
duditrait <- dudi.pca(aravo$traits, scannf = FALSE, nf = 2, row.w = coa1$cw)
rlq1 <- rlq(dudienv, coa1, duditrait, scannf = FALSE, nf = 2)
plot(rlq1)
```

ardeche

*Fauna Table with double (row and column) partitioning***Description**

This data set gives information about species of benthic macroinvertebrates in different sites and dates.

Usage

```
data(ardeche)
```

Format

`ardeche` is a list with 6 components.

tab is a data frame containing fauna table with 43 species (rows) and 35 samples (columns).

col.blocks is a vector containing the repartition of samples for the 6 dates : july 1982, august 1982, november 1982, february 1983, april 1983 and july 1983.

row.blocks is a vector containing the repartition of species in the 4 groups defining the species order.

dat.fac is a date factor for samples (6 dates).

sta.fac is a site factor for samples (6 sites).

esp.fac is a species order factor (Ephemeroptera, Plecoptera, Coleoptera, Trichoptera).

Details

The columns of the data frame `ardeche$tab` define the samples by a number between 1 and 6 (the date) and a letter between A and F (the site).

Source

Cazes, P., Chessel, D., and Dolédec, S. (1988) L'analyse des correspondances internes d'un tableau partitionné : son usage en hydrobiologie. *Revue de Statistique Appliquée*, **36**, 39–54.

Examples

```
data(ardeche)
dudi1 <- dudi.coa(ardeche$tab, scan = FALSE)
s.class(dudi1$co, ardeche$dat.fac)
s.label(dudi1$co, clab = 0.5, add.p = TRUE)
```

 area.plot

Graphical Display of Areas

Description

'area' is a data frame with three variables.

The first variable is a factor defining the polygons.

The second and third variables are the xy coordinates of the polygon vertices in the order where they are found.

area.plot : grey levels areas mapping

poly2area takes an object of class 'polylist' (maptools package) and returns a data frame of type area.

area2poly takes an object of type 'area' and returns a list of class 'polylist'

area2link takes an object of type 'area' and returns a proximity matrix which terms are given by the length of the frontier between two polygons.

area.util.contour, area.util.xy and area.util.class are three utility functions.

Usage

```
area.plot(x, center = NULL, values = NULL, graph = NULL, lwdgraph = 2,
nclasslegend = 8, clegend = 0.75, sub = "", csub = 1,
possub = "topleft", cpoint = 0, label = NULL, clabel = 0, ...)
```

```
area2poly(area)
poly2area(polys)
area2link(area)
area.util.contour(area)
area.util.xy(area)
```

Arguments

x	a data frame with three variables
center	a matrix with the same row number as x and two columns, the coordinates of polygone centers. If NULL, it is computed with area.util.xy
values	if not NULL, a vector which values will be mapped to grey levels. The values must be in the same order as the values in unique(x.area[,1])
graph	if not NULL, graph is a neighbouring graph (object of class "neig") between polygons
lwdgraph	a line width to draw the neighbouring graph

nclasslegend	if value not NULL, a number of classes for the legend
clegend	if not NULL, a character size for the legend, used with <code>par("cex")*clegend</code>
sub	a string of characters to be inserted as sub-title
csub	a character size for the sub-titles, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-titles position ("topleft", "topright", "bottomleft", "bottomright")
cpoint	if positive, a character size for drawing the polygons vertices (check up), used with <code>par("cex")*cpoint</code>
label	if not NULL, by default the levels of the factor that define the polygons are used as labels. To change this value, use label. These labels must be in the same order than <code>unique(x.area[,1])</code>
clabel	if not NULL, a character size for the polygon labels, used with <code>par("cex")*clabel</code>
polys	a list belonging to the 'polylist' class in the spdep package
area	a data frame of class 'area'
...	further arguments passed to or from other methods

Value

poly2area returns a data frame 'factor,x,y'.
 area2poly returns a list of class polylist.

Author(s)

Daniel Chessel

Examples

```
data(elec88)
par(mfrow = c(2,2))
area.plot(elec88$area, cpoint = 1)
area.plot(elec88$area, lab = elec88$lab$dep, clab = 0.75)
area.plot(elec88$area, clab = 0.75)
# elec88$neig <- neig(area = elec88$area)
area.plot(elec88$area, graph = elec88$neig,
sub = "Neighbourhood graph", possub = "topright")
par(mfrow = c(1,1))

## Not run:
par(mfrow = c(3,3))
for (i in 1:9) {
  x <- elec88$tab[,i]
  area.plot(elec88$area, val=x,
sub = names(elec88$tab)[i], csub = 3, cleg = 1.5)
}

par(mfrow = c(3,3))
```

```

for (i in 1:9) {
  x <- elec88$tab[,i]
  s.value(elec88$xy, elec88$tab[,i], contour = elec88$contour,
    meth = "greylevel", sub = names(elec88$tab)[i], csub = 3,
    cleg = 1.5, incl = FALSE)
}

data(irishdata)
par(mfrow = c(2,2))
w <- area.util.contour(irishdata$area)
xy <- area.util.xy(irishdata$area)
area.plot(irishdata$area, cpoint = 1)
apply(w, 1, function(x) segments(x[1],x[2],x[3],x[4], lwd = 3))
area.plot(irishdata$area, clabel = 1)
s.label(xy, area = irishdata$area, incl = FALSE, clab = 0,
  cpoi = 3, addax = FALSE, contour = w)
s.label(xy, area = irishdata$area, incl = FALSE,
  addax = FALSE, contour = w)
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  data(columbus)
  par(mfrow = c(2,2))
  plot(col.gal.nb, coords, pch = 20, cex = 2)
  col.gal.neig <- nb2neig(col.gal.nb)
  s.label(data.frame(coords), neig = col.gal.neig,
    inc = FALSE, addax = FALSE, clab = 0, cneig = 1, cpo = 2)
  maptools:::plot.polylist(polys,bbs)
  area.plot(poly2area(polys))

  # 1
  crime.f <- as.ordered(cut(columbus$CRIME,
    breaks = quantile(columbus$CRIME, probs = seq(0,1,0.2)),
    include.lowest = TRUE))
  colours <- c("salmon1", "salmon2", "red3", "brown", "black")
  plot(bbs[,1], bbs[,4], xlab = "", ylab = "", asp = 1, type = "n",
    xlim = range(c(bbs[,1], bbs[,3])), ylim = range(c(bbs[,2],
      bbs[,4])))
  for (i in 1:length(polys))
    polygon(polys[[i]], col = colours[unclass(crime.f[i])])
  legend(x = c(6, 7.75), y = c(13.5, 15), legend = levels(crime.f),
    fill = colours, cex = 0.7)
  title(sub = paste("Columbus OH: residential burglaries and ",
    "vehicle\nthefts", "per thousand households, 1980"))

  # 2
  area1 <- poly2area(polys)
  w <- area.util.contour(area1)
  wxy <- area.util.xy(area1)
  area.plot(area1, values = columbus$CRIME, sub = paste("Columbus ",
    "OH: residential burglaries and vehicle\nthefts",
    "per thousand households, 1980"))
  apply(w, 1, function(x) segments(x[1], x[2], x[3], x[4], lwd = 2))

  # 3

```

```

data(elec88)
fr.area <- elec88$area
fr.xy <- area.util.xy(fr.area)
fr.neig <- elec88$neig # neig(area = fr.area)

# 4
fr.poly <- area2poly(fr.area)
fr.nb <- neig2nb(fr.neig)
mapproj::plot.polylist(fr.poly, attr(fr.poly, "region.rect"), border = "grey")
plot(fr.nb, fr.xy, add = TRUE)
s.label(fr.xy, clab = 0, area = fr.area, neig = fr.neig,
cneig = 1, cpo = 2, inc = FALSE, addax = FALSE)
par(mfrow=c(1,1))
}
## End(Not run)

data(irishdata)
w <- irishdata$area[c(42:53,18:25),]
w
w$poly <- as.factor(as.character(w$poly))
area.plot(w, clab = 2)

points(68, 59, pch = 20, col = "red", cex = 3)
points(68, 35, pch = 20, col = "red", cex = 3)
points(45, 12, pch = 20, col = "red", cex = 3)
sqrt((59-35)^2)+sqrt((68-45)^2+(35-12)^2)
area2link(w)

```

arrival

Arrivals at an intensive care unit

Description

This data set gives arrival times of 254 patients at an intensive care unit during one day.

Usage

```
data(arrival)
```

Format

arrival is a list containing the 2 following objects :

times is a vector giving the arrival times in the form HH:MM

hours is a vector giving the number of arrivals per hour for the day considered

Source

Data taken from the logiciel Oriana developed by Warren L. Kovach <sales@kovcomp.com> starting from <http://www.kovcomp.com/oriana/index.html>.

References

Fisher, N. I. (1993) *Statistical Analysis of Circular Data*. Cambridge University Press.

Examples

```
data(arrival)
dotcircle(arrival$hours, pi/2 + pi/12)
```

as.taxo

Taxonomy

Description

The function `as.taxo` creates an object of class `taxo` that is a sub-class of `data.frame`. Each column of the data frame must be a factor corresponding to a level j of the taxonomy (genus, family, ...). The levels of factor j define some classes that must be completely included in classes of factor $j+1$.

A factor with exactly one level is not allowed. A factor with exactly one individual in each level is not allowed. The function `dist.taxo` compute taxonomic distances.

Usage

```
as.taxo(df)
dist.taxo(taxo)
```

Arguments

<code>df</code>	a data frame
<code>taxo</code>	a data frame of class <code>taxo</code>

Value

`as.taxo` returns a data frame of class `taxo`. `dist.taxo` returns a numeric of class `dist`.

Author(s)

Daniel Chessel
Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

See Also

[taxo2phylog](#) to transform an object of class `taxo` into an object of class `phylog`

Examples

```
data(taxo.eg)
tax <- as.taxo(taxo.eg[[1]])
tax.phy <- taxo2phylog(as.taxo(taxo.eg[[1]]),add.tools=TRUE)
par(mfrow = c(1,2))
plot(tax.phy, clabel.l = 1.25, clabel.n = 1.25, f = 0.75)
plot(taxo2phylog(as.taxo(taxo.eg[[1]][sample(15),)]),
      clabel.l = 1.25, clabel.n = 1.25, f = 0.75)
par(mfrow = c(1,1))
all(dist.taxo(tax)==tax.phy$Wdist)
```

atlas

Small Ecological Dataset

Description

atlas is a list containing three kinds of information about 23 regions (The French Alps) : geographical coordinates, meteorology and bird presences.

Usage

```
data(atlas)
```

Format

This list contains the following objects:

area is a convex hull of 23 geographical regions.

xy are the coordinates of the region centers and altitude (in meters).

names.district is a vector of region names.

meteo is a data frame with 7 variables: min and max temperature in january; min and max temperature in july; january, july and total rainfalls.

birds is a data frame with 15 variables (species).

alti is a data frame with 3 variables altitude in percentage [0,800],]800,1500] and]1500,5000].

Source

Extract from:

Lebreton, Ph. (1977) Les oiseaux nicheurs rhonalpins. *Atlas ornithologique Rhone-Alpes*. Centre Ornithologique Rhone-Alpes, Université Lyon 1, 69621 Villeurbanne. Direction de la Protection de la Nature, Ministère de la Qualité de la Vie. 1–354.

Examples

```

data(atlas)
op <- par(no.readonly = TRUE)
par(mfrow = c(2,2))
area.plot(atlas$area, cpoim = 1.5)
area.plot(atlas$area, lab = atlas$names.district, clab = 1)
x <- atlas$meteo$mini.jan

names(x) <- row.names(atlas$meteo)
area.plot(atlas$area, val = x, ncl = 12, sub = "Temp Mini January",
          csub = 2, cleg = 1)
s.corcircle((dudi.pca(atlas$meteo, scann = FALSE)$co),
            clab = 1)

area.plot(atlas$area, val = dudi.pca(atlas$meteo,scann=FALSE)$li[,1],
          ncl = 12, sub = "Principal Component Analysis analysis", csub = 1.5,
          cleg = 1)
birds.coa <- dudi.coa(atlas$birds, sca = FALSE, nf = 1)
x <- birds.coa$li$Axis1
area.plot(atlas$area, val = x, ncl = 12,
          sub = "Correspondence analysis", csub = 1.5, cleg = 1)

s.value(atlas$xy, x, contour = atlas$contour, csi = 2,
        incl = FALSE, addax = FALSE)
triangle.plot(atlas$alti)
par(op)
par(mfrow=c(1,1))

```

atya

Genetic variability of Cacadors

Description

This data set contains information about genetic variability of *Atya innocous* and *Atya scabra* in Guadeloupe (France).

Usage

```
data(atya)
```

Format

atya is a list with the following objects :

xy : a data frame with the coordinates of the 31 sites

gen : a data frame with 22 variables collected on 31 sites

neig : an object of class neig

Source

Fievet, E., Eppe, F. and Dolédec, S. (2001) Etude de la variabilité morphométrique et génétique des populations de Cacadors (*Atya innocous* et *Atya scabra*) de l'île de Basse-Terre. Direction Régionale de L'Environnement Guadeloupe, Laboratoire des hydrosystèmes fluviaux, Université Lyon 1.

Examples

```
## Not run:
data(atya)
if (require(pixmap, quiet = TRUE)) {
  atya.digi <- read.pnm(system.file("pictures/atyadigi.pnm",
    package = "ade4"))
  atya.carto <- read.pnm(system.file("pictures/atyacarto.pnm",
    package = "ade4"))
  par(mfrow = c(1,2))
  plot(atya.digi)
  plot(atya.carto)
  points(atya$xy, pch = 20, cex = 2)
}
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  plot(neig2nb(atya$neig), atya$xy, col = "red", add = TRUE, lwd = 2)
  par(mfrow = c(1,1))
}

## End(Not run)
```

 avijons

Bird species distribution

Description

This data set contains information about spatial distribution of bird species in a zone surrounding the river Rhône near Lyon (France).

Usage

```
data(avijons)
```

Format

avijons is a list with the following objects :

xy : a data frame with the coordinates of the sites

area : an object of class area

fau : a data frame with the abundance of 64 bird species in 91 sites

spe.names.fr : a vector of strings of character with the species names in french

Source

Bournaud, M., Amoros, C., Chessel, D., Coulet, M., Doledec, S., Michelot, J.L., Pautou, G., Rostan, J.C., Tachet, H. and Thioulouse, J. (1990) *Peuplements d'oiseaux et propriétés des écosystèmes de la plaine du Rhône : descripteurs de fonctionnement global et gestion des berges*. Rapport programme S.R.E.T.I.E., Ministère de l'Environnement CORA et URA CNRS 367, Univ. Lyon I.

References

Thioulouse, J., Chessel, D. and Champely, S. (1995) Multivariate analysis of spatial patterns: a unified approach to local and global structures. *Environmental and Ecological Statistics*, **2**, 1–14.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps051.pdf> (in French).

Examples

```
data(avijons)
w1=dudi.coa(avijons$fau,scannf=FALSE)$li
area.plot(avijons$area,center=avijons$xy,val=w1[,1],clab=0.75,sub="CA Axis 1",csub=3)
## Not run:
data(avijons)
if (require(pixmap,quiet=TRUE)) {
  pnm.eau <- read.pnm(system.file("pictures/avijonseau.pnm",
    package = "ade4"))
  pnm.rou <- read.pnm(system.file("pictures/avijonsrou.pnm",
    package = "ade4"))
  pnm.veg <- read.pnm(system.file("pictures/avijonsveg.pnm",
    package = "ade4"))
  pnm.vil <- read.pnm(system.file("pictures/avijonsvil.pnm",
    package = "ade4"))
  jons.coa <- dudi.coa(avijons$fau, scan = FALSE, nf = 4)
  par(mfcol = c(3,2))
  s.value(avijons$xy, jons.coa$li[,1], pixmap = pnm.rou, inclu = FALSE,
    grid = FALSE, addax = FALSE, cleg = 0, sub = "F1+ROADS", csub = 3)
  s.value(avijons$xy, jons.coa$li[,1], pixmap = pnm.veg, inclu = FALSE,
    grid = FALSE, addax = FALSE, cleg = 0, sub = "F1+TREES", csub = 3)
  s.value(avijons$xy, jons.coa$li[,1], pixmap = pnm.eau, inclu = FALSE,
    grid = FALSE, addax = FALSE, cleg = 0, sub = "F1+WATER", csub = 3)
  s.value(avijons$xy, jons.coa$li[,2], pixmap = pnm.rou, inclu = FALSE,
    grid = FALSE, addax = FALSE, cleg = 0, sub = "F2+ROADS", csub = 3)
  s.value(avijons$xy, jons.coa$li[,2], pixmap = pnm.veg, inclu = FALSE,
    grid = FALSE, addax = FALSE, cleg = 0, sub = "F2+TREES", csub = 3)
  s.value(avijons$xy, jons.coa$li[,2], pixmap = pnm.eau, inclu = FALSE,
    grid = FALSE, addax = FALSE, cleg = 0, sub = "F2+WATER", csub = 3)
  par(mfrow=c(1,1))}

if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)
  & require(pixmap, quiet = TRUE) ) {
  link1 <- area2link(avijons$area)
  lw1 <- apply(link1,1,function(x) x[x>0])
  neig1 <- neig(mat01=1*(link1>0))
  nb1 <- neig2nb(neig1)
  listw1 <- nb2listw(nb1,lw1)
```

```

jons.ms <- multispati(jons.coa, listw1, scan = FALSE, nfp = 3,
  nfn = 2)
summary(jons.ms)
par(mfrow = c(2,2))
barplot(jons.coa$eig)
barplot(jons.ms$eig)
s.corcircle(jons.ms$as)
plot(jons.coa$li[,1], jons.ms$li[,1])

par(mfcol=c(3,2))
s.value(avijons$xy, jons.ms$li[,1], pixmap = pnm.rou, inclu = FALSE,
  grid = FALSE, addax = FALSE, cleg = 0, sub = "F1+ROADS", csub = 3)
s.value(avijons$xy, jons.ms$li[,1], pixmap = pnm.veg, inclu = FALSE,
  grid = FALSE, addax = FALSE, cleg = 0, sub = "F1+TREES", csub = 3)
s.value(avijons$xy, jons.ms$li[,1], pixmap = pnm.eau, inclu = FALSE,
  grid = FALSE, addax = FALSE, cleg = 0, sub = "F1+WATER", csub = 3)
s.value(avijons$xy, jons.ms$li[,2], pixmap = pnm.rou, inclu = FALSE,
  grid = FALSE, addax = FALSE, cleg = 0, sub = "F2+ROADS", csub = 3)
s.value(avijons$xy, jons.ms$li[,2], pixmap = pnm.veg, inclu = FALSE,
  grid = FALSE, addax = FALSE, cleg = 0, sub = "F2+TREES", csub = 3)
s.value(avijons$xy, jons.ms$li[,2], pixmap = pnm.eau, inclu = FALSE,
  grid = FALSE, addax = FALSE, cleg = 0, sub = "F2+WATER", csub = 3)
par(mfrow=c(1,1))
}
## End(Not run)

```

avimedi

Fauna Table for Constrained Ordinations

Description

avimedi is a list containing the information about 302 sites :
frequencies of 51 bird species ; two factors (habitats and Mediterranean origin).

Usage

```
data(avimedi)
```

Format

This list contains the following objects:

fau is a data frame 302 sites - 51 bird species.

plan is a data frame 302 sites - 2 factors : reg with two levels Provence (Pr, South of France) and Corsica (Co) ; str with six levels describing the vegetation from a very low matorral (1) up to a mature forest of holm oaks (6).

nomesp is a vector 51 latin names.

Source

Blondel, J., Chessel, D., & Frochet, B. (1988) Bird species impoverishment, niche expansion, and density inflation in mediterranean island habitats. *Ecology*, **69**, 1899–1917.

Examples

```
## Not run:
data(avimedi)
par(mfrow = c(2,2))
coa1 <- dudi.coa(avimedi$fau, scan = FALSE, nf = 3)
s.class(coa1$li, avimedi$plan$str:avimedi$plan$reg,
        sub = "Correspondences Analysis")
bet1 <- bca(coa1, avimedi$plan$str, scan = FALSE)
s.class(bet1$ls, avimedi$plan$str,
        sub = "Between Analysis")
wit1 <- wca(coa1, avimedi$plan$reg, scan=FALSE)
s.class(wit1$li, avimedi$plan$str,
        sub = "Within Analysis")
pcaiv1 <- pcaiv(coa1, avimedi$plan, scan = FALSE)
s.match(pcaiv1$li, pcaiv1$ls, clab = 0,
        sub = "Canonical Correspondences Analysis")
s.class(pcaiv1$li, avimedi$plan$str:avimedi$plan$reg,
        add.plot = TRUE)
par(mfrow=c(1,1))

## End(Not run)
```

 aviurba

Ecological Tables Triplet

Description

This data set is a list of information about 51 sites : bird species and environmental variables.
A data frame contains biological traits for each species.

Usage

```
data(aviurba)
```

Format

This list contains the following objects:

fau is a data frame 51 sites 40 bird species.

mil is a data frame 51 sites 11 environmental variables (see details).

traits is a data frame 40 species 4 biological traits (see details).

species.names.fr is a vector of the species names in french.

species.names.la is a vector of the species names in latin.

species.family is a factor : the species families.

Details

aviurba\$mil contains for each site, 11 habitat attributes describing the degree of urbanization. The presence or absence of farms or villages, small buildings, high buildings, industry, fields, grassland, scrubby areas, deciduous woods, coniferous woods, noisy area are noticed. At least, the vegetation cover (variable 11) is a factor with 8 levels from a minimum cover (R5) up to a maximum (R100).

aviurba\$traits contains four factors : feeding habit (insectivor, granivore, omnivore), feeding stratum (ground, aerial, foliage and scrub), breeding stratum (ground, building, scrub, foliage) and migration strategy (resident, migrant).

Source

Dolédec, S., Chessel, D., Ter Braak, C. J. F. and Champely S. (1996) Matching species traits to environmental variables: a new three-table ordination method. *Environmental and Ecological Statistics*, **3**, 143–166.

Examples

```
data(aviurba)
a1 <- dudi.coa(aviurba$fau, scan = FALSE, nf=4)
a2 <- dudi.acm(aviurba$mil, row.w = a1$lw, scan = FALSE, nf = 4)
plot(coinertia(a1, a2, scan = FALSE))
```

bacteria

Genomes of 43 Bacteria

Description

bacteria is a list containing 43 species and genomic informations : codons, amino acid and bases.

Usage

```
data(bacteria)
```

Format

This list contains the following objects:

code is a factor with the amino acid names for each codon.

espcodon is a data frame 43 species 64 codons.

espa is a data frame 43 species 21 amino acid.

espbases is a data frame 43 species 4 bases.

Source

Data prepared by J. Lobry <lobry@biomserv.univ-lyon1.fr> starting from:
<http://www.tigr.org/tdb/mdb/mdbcomplete.html>

Examples

```

data(bacteria)
names(bacteria$espcodon)
names(bacteria$espaa)
names(bacteria$espbase)
sum(bacteria$espcodon) # 22,619,749 codons
scatter.coa(dudi.coa(bacteria$espcodon, scann = FALSE),
            posi = "bottom")

```

banque

Table of Factors

Description

banque gives the results of a bank survey onto 810 customers.

Usage

```
data(banque)
```

Format

This data frame contains the following columns:

1. csp: "Socio-professional categories" a factor with levels
 - agric Farmers
 - artis Craftsmen, Shopkeepers, Company directors
 - cadsu Executives and higher intellectual professions
 - inter Intermediate professions
 - emplo Other white-collar workers
 - ouvri Manual workers
 - retra Pensionners
 - inact Non working population
 - etudi Students
2. duree: "Time relations with the customer" a factor with levels
 - dm2 <2 years
 - d24 [2 years, 4 years[
 - d48 [4 years, 8 years[
 - d812 [8 years, 12 years[
 - dp12 >= 12 years
3. oppo: "Stopped a check ?" a factor with levels
 - non no
 - oui yes
4. age: "Customer's age" a factor with levels

- ai25 [18 years, 25 years[
 - ai35 [25 years, 35 years[
 - ai45 [35 years, 45 years[
 - ai55 [45 years, 55 years[
 - ai75 [55 years, 75 years[
5. sexe: "Customer's gender" a factor with levels
 - hom Male
 - fem Female
 6. interdit: "No checkbook allowed" a factor with levels
 - non no
 - oui yes
 7. cableue: "Possess a bank card ?" a factor with levels
 - non no
 - oui yes
 8. assurvi: "Contrat of life insurance ?" a factor with levels
 - non no

 - oui yes
 9. soldevu: "Balance of the current accounts" a factor with levels
 - p4 credit balance > 20000
 - p3 credit balance 12000-20000
 - p2 credit balance 4000-120000
 - p1 credit balance >0-4000
 - n1 debit balance 0-4000
 - n2 debit balance >4000
 10. eparlog: "Savings and loan association account amount" a factor with levels
 - for > 20000
 - fai >0 and <20000
 - nul nulle
 11. eparliv: "Savings bank amount" a factor with levels
 - for > 20000
 - fai >0 and <20000
 - nul nulle
 12. credhab: "Home loan owner" a factor with levels
 - non no
 - oui yes
 13. credcon: "Consumer credit amount" a factor with levels
 - nul none
 - fai >0 and <20000
 - for > 20000

14. versesp: "Check deposits" a factor with levels
 - oui yes
 - non no
15. retresp: "Cash withdrawals" a factor with levels
 - fai < 2000
 - moy 2000-5000
 - for > 5000
16. remiche: "Endorsed checks amount" a factor with levels
 - for >10000
 - moy 10000-5000
 - fai 1-5000
 - nul none
17. preltre: "Treasury Department tax deductions" a factor with levels
 - nul none
 - fai <1000
 - moy >1000
18. prelfm: "Financial institution deductions" a factor with levels
 - nul none
 - fai <1000
 - moy >1000
19. viredeb: "Debit transfer amount" a factor with levels
 - nul none
 - fai <2500
 - moy 2500-5000
 - for >5000
20. virecre: "Credit transfer amount" a factor with levels
 - for >10000
 - moy 10000-5000
 - fai <5000
 - nul aucun
21. porttit: "Securities portfolio estimations" a factor with levels
 - nul none
 - fai < 20000
 - moy 20000-100000
 - for >100000

Source

anonymous

Examples

```
data(banque)
banque.acm <- dudi.acm(banque, scann = FALSE, nf = 3)
apply(banque.acm$cr, 2, mean)
banque.acm$eig[1:banque.acm$nf] # the same thing
s.arrow(banque.acm$c1, clab = 0.75)
```

baran95

African Estuary Fishes

Description

This data set is a list containing relations between sites and fish species linked to dates.

Usage

```
data(baran95)
```

Format

This list contains the following objects:

fau is a data frame 95 seinings and 33 fish species.

plan is a data frame 2 factors : date and site. The date has 6 levels (april 1993, june 1993, august 1993, october 1993, december 1993 and february 1994) and the sites are defined by 4 distances to the Atlantic Ocean (km03, km17, km33 and km46).

species.names is a vector of species latin names.

Source

Baran, E. (1995) *Dynamique spatio-temporelle des peuplements de Poissons estuariens en Guinée (Afrique de l'Ouest)*. Thèse de Doctorat, Université de Bretagne Occidentale. Data collected by net fishing sampling in the Fatala river estuary.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps027.pdf> (in French).

Examples

```
data(baran95)
w <- dudi.pca(log(baran95$fau+1), scal = FALSE, scann = FALSE,
             nf = 3)
w1 <- wca(w, baran95$plan$date, scann = FALSE)
fatala <- ktab.within(w1)
stat1 <- stasis(fatala, scan = FALSE, nf = 3)
w1 <- split(stat1$C.Co, baran95$plan$date)
w2 <- split(baran95$plan$site, baran95$plan$date)
```

```

par(mfrow = c(3,2))
for (j in 1:6) {
  s.label(stat1$C.Co[,1:2], clab = 0,
  sub = tab.names(fatala)[j], csub = 3)
  s.class(w1[[j]][,1:2], w2[[j]], clab = 2, axese = FALSE,
  add.plot = TRUE)
}
par(mfrow = c(1,1))

kplot(stat1, arrow = FALSE, traj = FALSE, clab = 2, uni = TRUE,
class = baran95$plan$site) #simpler

mfa1 <- mfa(fatala, scan = FALSE, nf = 3)
w1 <- split(mfa1$co, baran95$plan$date)
w2 <- split(baran95$plan$site, baran95$plan$date)
par(mfrow = c(3,2))
for (j in 1:6) {
  s.label(mfa1$co[,1:2], clab = 0,
  sub = tab.names(fatala)[j], csub = 3)
  s.class(w1[[j]][,1:2], w2[[j]], clab = 2, axese=FALSE,
  add.plot = TRUE)
}
par(mfrow = c(1,1))

```

bca

Between-Class Analysis

Description

Performs a particular case of a Principal Component Analysis with respect to Instrumental Variables (pcaiv), in which there is only a single factor as explanatory variable.

Usage

```

between(dudi, fac, scannf = TRUE, nf = 2)
## S3 method for class 'dudi'
bca(x, fac, scannf = TRUE, nf = 2, ...)

```

Arguments

dudi	a duality diagram, object of class <code>dudi</code> obtained from the functions <code>dudi.coa</code> , <code>dudi.pca</code> ,...
x	a duality diagram, object of class <code>dudi</code> from one of the functions <code>dudi.coa</code> , <code>dudi.pca</code> ,...
fac	a factor partitioning the rows of <code>dudi\$tab</code> in classes
scannf	a logical value indicating whether the eigenvalues barplot should be displayed
nf	if <code>scannf</code> <code>FALSE</code> , a numeric value indicating the number of kept axes
...	further arguments passed to or from other methods

Value

Returns a list of class `dudi`, subclass 'between' containing

<code>tab</code>	a data frame class-variables containing the means per class for each variable
<code>cw</code>	a numeric vector of the column weights
<code>lw</code>	a numeric vector of the class weights
<code>eig</code>	a numeric vector with all the eigenvalues
<code>rank</code>	the rank of the analysis
<code>nf</code>	an integer value indicating the number of kept axes
<code>c1</code>	a data frame with the column normed scores
<code>l1</code>	a data frame with the class normed scores
<code>co</code>	a data frame with the column coordinates
<code>li</code>	a data frame with the class coordinates
<code>call</code>	the matching call
<code>ratio</code>	the between-class inertia percentage
<code>ls</code>	a data frame with the row coordinates
<code>as</code>	a data frame containing the projection of inertia axes onto between axes

Note

To avoid conflict names with the `base::within` function, the function `within` is now deprecated and will be removed. To be consistent, the `between` function is also deprecated and is replaced by the method `bca.dudi` of the new generic `bca` function.

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Dolédec, S. and Chessel, D. (1987) Rythmes saisonniers et composantes stationnelles en milieu aquatique I- Description d'un plan d'observations complet par projection de variables. *Acta Oecologica, Oecologia Generalis*, **8**, 3, 403–426.

Examples

```
data(meaudret)
par(mfrow = c(2,2))
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)
s.class(pca1$li, meaudret$design$site,
  sub = "Principal Component Analysis (env)", csub = 1.75)
pca2 <- dudi.pca(meaudret$spe, scal = FALSE, scan = FALSE, nf = 4)
s.class(pca2$li, meaudret$design$site,
  sub = "Principal Component Analysis (spe)", csub = 1.75)
bet1 <- bca(pca1, meaudret$design$site, scan = FALSE, nf = 2)
```

```
bet2 <- bca(pca2, meaudret$design$site, scan = FALSE, nf = 2)
s.class(bet1$ls, meaudret$design$site,
        sub = "Between sites PCA (env)", csub = 1.75)
s.class(bet2$ls, meaudret$design$site,
        sub = "Between sites PCA (spe)", csub = 1.75)

par(mfrow = c(1,1))
coib <- coinertia(bet1, bet2, scannf = FALSE)
plot(coib)
```

bca.coinertia

Between-class coinertia analysis

Description

Performs a between-class analysis after a coinertia analysis

Usage

```
betweencoinertia(obj, fac, scannf = TRUE, nf = 2)
## S3 method for class 'coinertia'
bca(x, fac, scannf = TRUE, nf = 2, ...)
```

Arguments

obj	a coinertia analysis (object of class coinertia) obtained by the function coinertia
x	a coinertia analysis (object of class coinertia) obtained by the function coinertia
fac	a factor partitioning the rows in classes
scannf	a logical value indicating whether the eigenvalues barplot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
...	further arguments passed to or from other methods

Details

This analysis is equivalent to do a between-class analysis on each initial dudi, and a coinertia analysis on the two between analyses. This function returns additional outputs for the interpretation.

Value

An object of the class `betcoi`. Outputs are described by the `print` function

Note

To avoid conflict names with the `base::within` function, the function `within` is now deprecated and will be removed. To be consistent, the `betweencoinertia` function is also deprecated and is replaced by the method `bca.coinertia` of the new generic `bca` function.

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr> and Jean Thioulouse <jean.thioulouse@univ-lyon1.fr>

References

Franquet E., Doledec S., and Chessel D. (1995) Using multivariate analyses for separating spatial and temporal effects within species-environment relationships. *Hydrobiologia*, **300**, 425–431.

See Also

[coinertia, between](#)

Examples

```
data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)
pca2 <- dudi.pca(meaudret$spe, scal = FALSE, scan = FALSE, nf = 4)

bet1 <- bca(pca1, meaudret$design$site, scan = FALSE, nf = 2)
bet2 <- bca(pca2, meaudret$design$site, scan = FALSE, nf = 2)
coib <- coinertia(bet1, bet2, scannf = FALSE)

coi <- coinertia(pca1,pca2,scannf = FALSE,nf = 3)
coi.b <- bca(coi,meaudret$design$site, scannf = FALSE)
## coib and coi.b are equivalent

plot(coi.b)
```

bca.rlq

Between-Class RLQ analysis

Description

Performs a particular RLQ analysis where a partition of sites (rows of R) is taken into account. The between-class RLQ analysis search for linear combinations of traits and environmental variables maximizing the covariances between the traits and the average environmental conditions of classes.

Usage

```
## S3 method for class 'rlq'
bca(x, fac, scannf = TRUE, nf = 2, ...)
## S3 method for class 'betrlq'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'betrlq'
print(x, ...)
```

Arguments

x	an object of class r1q (created by the r1q function) for the bca.r1q function. An object of class betr1q for the print and plot functions
fac	a factor partitioning the rows of R
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
xax	the column number for the x-axis
yax	the column number for the y-axis
...	further arguments passed to or from other methods

Value

The bca.r1q function returns an object of class 'betr1q' (sub-class of 'dudi'). See the outputs of the print function for more details.

Author(s)

Stephane Dray <stephane.drays@univ-lyon1.fr>

References

Wesuls, D., Oldeland, J. and Dray, S. (2012) Disentangling plant trait responses to livestock grazing from spatio-temporal variation: the partial RLQ approach. *Journal of Vegetation Science*, **23**, 98–113.

See Also

[r1q](#), [bca](#), [wca.r1q](#)

Examples

```
data(piosphere)
afcL <- dudi.coa(log(piosphere$veg + 1), scannf = FALSE)
acpR <- dudi.pca(piosphere$env, scannf = FALSE, row.w = afcL$lw)
acpQ <- dudi.hillsmith(piosphere$traits, scannf = FALSE, row.w =
  afcL$cw)
r1q1 <- r1q(acpR, afcL, acpQ, scannf = FALSE)

br1q1 <- bca(r1q1, fac = piosphere$habitat, scannf = FALSE)
br1q1
plot(br1q1)
```

between	<i>Between-Class Analysis</i>
---------	-------------------------------

Description

Outputs and graphical representations of the results of a between-class analysis.

Usage

```
## S3 method for class 'between'  
plot(x, xax = 1, yax = 2, ...)  
## S3 method for class 'between'  
print(x, ...)  
## S3 method for class 'betcoi'  
plot(x, xax = 1, yax = 2, ...)  
## S3 method for class 'betcoi'  
print(x, ...)
```

Arguments

x	an object of class between or betcoi
xax, yax	the column index of the x-axis and the y-axis
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>
Stephane Dray <stephane.dray@univ-lyon1.fr>

References

Dolédec, S. and Chessel, D. (1987) Rythmes saisonniers et composantes stationnelles en milieu aquatique I- Description d'un plan d'observations complet par projection de variables. *Acta Oecologica, Oecologia Generalis*, **8**, 3, 403–426.

See Also

[bca.dudi](#), [bca.coinertia](#)

Examples

```
data(meaudret)  
par(mfrow = c(2,2))  
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)  
s.class(pca1$li, meaudret$design$site,  
        sub = "Principal Component Analysis (env)", csub = 1.75)  
pca2 <- dudi.pca(meaudret$spe, scal = FALSE, scan = FALSE, nf = 4)
```

```

s.class(pca2$li, meaudret$design$site,
       sub = "Principal Component Analysis (spe)", csub = 1.75)
bet1 <- bca(pca1, meaudret$design$site, scan = FALSE, nf = 2)
bet2 <- bca(pca2, meaudret$design$site, scan = FALSE, nf = 2)
s.class(bet1$ls, meaudret$design$site,
       sub = "Between sites PCA (env)", csub = 1.75)
s.class(bet2$ls, meaudret$design$site,
       sub = "Between sites PCA (spe)", csub = 1.75)

par(mfrow = c(1,1))
coib <- coinertia(bet1, bet2, scann = FALSE)
plot(coib)

```

bf88

*Cubic Ecological Data***Description**

bf88 is a list of 6 data frames corresponding to 6 stages of vegetation. Each data frame gives some bird species informations for 4 counties.

Usage

```
data(bf88)
```

Format

A list of six data frames with 79 rows (bird species) and 4 columns (counties). The 6 arrays (S1 to S6) are the 6 stages of vegetation. The attribut 'nomesp' of this list is a vector of species French names.

Source

Blondel, J. and Farre, H. (1988) The convergent trajectories of bird communities along ecological successions in european forests. *Oecologia* (Berlin), **75**, 83–93.

Examples

```

data(bf88)
fou1 <- foucart(bf88, scann = FALSE, nf = 3)
fou1
par(mfrow = c(2,2))
scatter(fou1)
s.traject(fou1$Tco, fou1$TC[,1])
s.traject(fou1$Tco, fou1$TC[,2])
s.label(fou1$Tco)
s.label(fou1$co, add.p = TRUE, clab = 2)
par(mfrow = c(1,1))
kplot(fou1, clab.c = 2, clab.r = 0, csub = 3)

```

bicenter.wt	<i>Double Weighted Centring</i>
-------------	---------------------------------

Description

This function creates a doubly centred matrix.

Usage

```
bicenter.wt(X, row.wt = rep(1, nrow(X)), col.wt = rep(1, ncol(X)))
```

Arguments

X	a matrix with n rows and p columns
row.wt	a vector of positive or null weights of length n
col.wt	a vector of positive or null weights of length p

Value

returns a doubly centred matrix

Author(s)

Daniel Chessel

Examples

```
w <- matrix(1:6, 3, 2)
bicenter.wt(w, c(0.2,0.6,0.2), c(0.3,0.7))
```

```
w <- matrix(1:20, 5, 4)
sum(bicenter.wt(w, runif(5), runif(4))^2)
```

bordeaux	<i>Wine Tasting</i>
----------	---------------------

Description

The bordeaux data frame gives the opinions of 200 judges in a blind tasting of five different types of claret (red wine from the Bordeaux area in the south western parts of France).

Usage

```
data(bordeaux)
```

Format

This data frame has 5 rows (the wines) and 4 columns (the judgements) divided in excellent, good, mediocre and boring.

Source

van Rijckevorsel, J. (1987) *The application of fuzzy coding and horseshoes in multiple correspondence analysis*. DSWO Press, Leiden (p. 32)

Examples

```
data(bordeaux)
bordeaux
score(dudi.coa(bordeaux, scan = FALSE))
```

bsetal97

Ecological and Biological Traits

Description

This data set gives ecological and biological characteristics of 131 species of aquatic insects.

Usage

```
data(bsetal97)
```

Format

bsetal97 is a list of 8 components.

species.names is a vector of the names of aquatic insects.

taxo is a data frame containing the taxonomy of species: genus, family and order.

biol is a data frame containing 10 biological traits for a total of 41 modalities.

biol.blo is a vector of the numbers of items for each biological trait.

biol.blo.names is a vector of the names of the biological traits.

ecol is a data frame with 7 ecological traits for a total of 34 modalities.

ecol.blo is a vector of the numbers of items for each ecological trait.

ecol.blo.names is a vector of the names of the ecological traits.

Details

The 10 variables of the data frame `bsetal97$biol` are called in `bsetal97$biol.blo.names` and the number of modalities per variable given in `bsetal97$biol.blo`. The variables are: female size - the body length from the front of the head to the end of the abdomen (7 length modalities), egg length - the egg size (6 modalities), egg number - count of eggs actually oviposited, generations per year (3 modalities: ≤ 1 , 2, > 2), oviposition period - the length of time during which oviposition occurred (3 modalities: ≤ 2 months, between 2 and 5 months, > 5 months), incubation time - the time between oviposition and hatching of the larvae (3 modalities: ≤ 4 weeks, between 4 and 12 weeks, > 12 weeks), egg shape (1-spherical, 2-oval, 3-cylindrical), egg attachment - physiological feature of the egg and of the female (4 modalities), clutch structure (1-single eggs, 2-grouped eggs, 3-egg masses), clutch number (3 modalities : 1, 2, > 2).

The 7 variables of the data frame `bsetal97$ecol` are called in `bsetal97$ecol.blo.names` and the number of modalities per variable given in `bsetal97$ecol.blo`. The variables are: oviposition site - position relative to the water (7 modalities), substratum type for eggs - the substratum to which the eggs are definitely attached (6 modalities), egg deposition - the position of the eggs during the oviposition process (4 modalities), gross habitat - the general habitat use of the species such as temporary waters or estuaries (8 modalities), saturation variance - the exposure of eggs to the risk of dessication (2 modalities), time of day (1-morning, 2-day, 3-evening, 4-night), season - time of the year (1-Spring, 2-Summer, 3-Autumn).

Source

Statzner, B., Hoppenhaus, K., Arens, M.-F. and Richoux, P. (1997) Reproductive traits, habitat use and templet theory: a synthesis of world-wide data on aquatic insects. *Freshwater Biology*, **38**, 109–135.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps029.pdf> (in French).

Examples

```
data(bsetal97)
X <- prep.fuzzy.var(bsetal97$biol, bsetal97$biol.blo)
Y <- prep.fuzzy.var(bsetal97$ecol, bsetal97$ecol.blo)
plot(coinertia(dudi.fca(X, scan = FALSE),
              dudi.fca(Y, scan = FALSE), scan = FALSE))
```

buech

Buech basin

Description

This data set contains informations about Buech basin characteristics.

Usage

```
data(buech)
```

Format

buech is a list with the following components :

tab1 : a data frame with 10 environmental variables collected on 31 sites in Juin (1984)

tab2 : a data frame with 10 environmental variables collected on 31 sites in September (1984)

xy : a data frame with the coordinates of the sites

neig : an object of class neig

contour : a data frame for background map

Details

Variables of buech\$tab1 and buech\$tab2 are the following ones :

pH ; Conductivity (μ S/cm) ; Carbonate (water hardness (mg/l CaCO₃)) ; hardness (total water hardness (mg/l CaCO₃)) ; Bicarbonate (alcalinity (mg/l HCO₃⁻)) ; Chloride (alcalinity (mg/l Cl⁻)) ; Suspens (particles in suspension (mg/l)) ; Organic (organic particles (mg/l)) ; Nitrate (nitrate rate (mg/l NO₃⁻)) ; Ammonia (amoniac rate (mg/l NH₄⁻))

Source

Vespini, F. (1985) *Contribution à l'étude hydrobiologique du Buech, rivière non aménagée de Haute-Provence*. Thèse de troisième cycle, Université de Provence.

Vespini, F., Légier, P. and Champeau, A. (1987) Ecologie d'une rivière non aménagée des Alpes du Sud : Le Buëch (France) I. Evolution longitudinale des descripteurs physiques et chimiques. *Annales de Limnologie*, **23**, 151–164.

Examples

```
data(buech)
par(mfrow = c(1,2))
s.label(buech$xy, contour = buech$contour, neig = buech$neig)
s.value (buech$xy, buech$tab2$Suspens-buech$tab1$Suspens,
        contour = buech$contour, neig = buech$neig, csi = 3)
par(mfrow = c(1,1))
```

butterfly

Genetics-Ecology-Environment Triple

Description

This data set contains environmental and genetics informations about 16 *Euphydryas editha* butterfly colonies studied in California and Oregon.

Usage

```
data(butterfly)
```

Format

`butterfly` is a list with 4 components.

`xy` is a data frame with the two coordinates of the 16 *Euphydryas editha* butterfly colonies.

`envir` is an environmental data frame of 16 sites - 4 variables.

`genet` is a genetics data frame of 16 sites - 6 allele frequencies.

`contour` is a data frame for background map (California map).

Source

McKechnie, S.W., Ehrlich, P.R. and White, R.R. (1975) Population genetics of *Euphydryas* butterflies. I. Genetic variation and the neutrality hypothesis. *Genetics*, **81**, 571–594.

References

Manly, B.F. (1994) *Multivariate Statistical Methods. A primer*. Second edition. Chapman & Hall, London. 1–215.

Examples

```
data(butterfly)
par(mfrow = c(2,2))
s.label(butterfly$xy, contour = butterfly$contour, inc = FALSE)
table.dist(dist(butterfly$xy), labels = row.names(butterfly$xy)) # depends of mva
s.value(butterfly$xy, dudi.pca(butterfly$envir, scan = FALSE)$li[,1],
        contour = butterfly$contour, inc = FALSE, csi = 3)
plot(mantel.randtest(dist(butterfly$xy), dist(butterfly$gen), 99),
     main = "genetic/spatial")
par(mfrow = c(1,1))
```

cailliez

Transformation to make Euclidean a distance matrix

Description

This function computes the smallest positive constant that makes Euclidean a distance matrix and applies it.

Usage

```
cailliez(distmat, print = FALSE, tol = 1e-07, cor.zero = TRUE)
```

Arguments

<code>distmat</code>	an object of class <code>dist</code>
<code>print</code>	if <code>TRUE</code> , prints the eigenvalues of the matrix
<code>tol</code>	a tolerance threshold for zero
<code>cor.zero</code>	if <code>TRUE</code> , zero distances are not modified

Value

an object of class `dist` containing a Euclidean distance matrix.

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

Cailliez, F. (1983) The analytical solution of the additive constant problem. *Psychometrika*, **48**, 305–310.

Legendre, P. and Anderson, M.J. (1999) Distance-based redundancy analysis: testing multispecies responses in multifactorial ecological experiments. *Ecological Monographs*, **69**, 1–24.

Legendre, P., and Legendre, L. (1998) *Numerical ecology*, 2nd English edition edition. Elsevier Science BV, Amsterdam.

From the DistPCoA program of P. Legendre et M.J. Anderson
<http://www.fas.umontreal.ca/BIOL/Casgrain/en/labo/distpcoa.html>

Examples

```
data(capitales)
d0 <- capitales$dist
is.euclid(d0) # FALSE
d1 <- cailliez(d0, TRUE)
# Cailliez constant = 2429.87867
is.euclid(d1) # TRUE
plot(d0, d1)
abline(lm(unclass(d1)~unclass(d0)))
print(coefficients(lm(unclass(d1)~unclass(d0))), dig = 8) # d1 = d + Cte
is.euclid(d0 + 2428) # FALSE
is.euclid(d0 + 2430) # TRUE the smallest constant
```

capitales

Road Distances

Description

This data set gives the road distances between 15 European capitals and their coordinates.

Usage

```
data(capitales)
```


Format

This list contains the following objects:

dist is dist object the road distances between 15 European capitals.

xy is a data frame containing the coordinates of capitals.

area is a data frame containing three variables, designed to be used in area.plot function.

logo is a list of pixmap objects, each one symbolizing a capital

Source

<http://www.euro.gouv.fr/jeunes/eurocollege/tableaucarte.htm>

Examples

```
if (require(pixmap, quiet = TRUE)) {
  data(capitales)
  attr(capitales$dist, "Labels")
  index <- pmatch(tolower(attr(capitales$dist, "Labels")), names(capitales$logo))
  w1 <- capitales$area
  par(mfrow=c(2,2))
  s.label(capitales$xy, lab = attr(capitales$dist, "Labels"))
  area.plot(w1)
  rect(min(w1$x), min(w1$y), max(w1$x), max(w1$y), col = "lightblue")
  invisible(lapply(split(w1, w1$id), function(x) polygon(x[, -1], col = "white")))
  s.logo(capitales$xy, capitales$logo, klogo = index, add.plot =
TRUE, clogo = 0.5) # depends on pixmap
  table.dist(capitales$dist, lab = attr(capitales$dist, "Labels")) # depends on mva
  s.logo(pcoscaled(lingoes(capitales$dist)), capitales$logo,
klogo = index, clogo = 0.5) # depends on pixmap
}
```

carni19

Phylogeny and quantitative trait of carnivora

Description

This data set describes the phylogeny of carnivora as reported by Diniz-Filho et al. (1998). It also gives the body mass of these 19 species.

Usage

```
data(carni19)
```

Format

carni19 is a list containing the 2 following objects :

tre is a character string giving the phylogenetic tree in Newick format.

bm is a numeric vector which values correspond to the body mass of the 19 species (log scale).

Source

Diniz-Filho, J. A. F., de Sant'Ana, C.E.R. and Bini, L.M. (1998) An eigenvector method for estimating phylogenetic inertia. *Evolution*, **52**, 1247–1262.

Examples

```
data(carni19)
carni19.phy <- newick2phylog(carni19$tre)
par(mfrow = c(1,2))
symbols.phylog(carni19.phy, carn19$bm-mean(carni19$bm))
dotchart.phylog(carni19.phy, carn19$bm, clabel.l=0.75)
par(mfrow = c(1,1))
```

carni70

Phylogeny and quantitative traits of carnivora

Description

This data set describes the phylogeny of 70 carnivora as reported by Diniz-Filho and Torres (2002). It also gives the geographic range size and body size corresponding to these 70 species.

Usage

```
data(carni70)
```

Format

carni70 is a list containing the 2 following objects:

tre is a character string giving the phylogenetic tree in Newick format. Branch lengths are expressed as divergence times (millions of years)

tab is a data frame with 70 species and two traits: size (body size (kg)) ; range (geographic range size (km)).

Source

Diniz-Filho, J. A. F., and N. M. Tôrres. (2002) Phylogenetic comparative methods and the geographic range size-body size relationship in new world terrestrial carnivora. *Evolutionary Ecology*, **16**, 351–367.

Examples

```
## Not run:
data(carni70)
carni70.phy <- newick2phylog(carni70$tre)
plot.phylog(carni70.phy)

size <- scalewt(log(carni70$tab))[,1]
```

```

names(size) <- row.names(carni70$stab)
symbols.phylog(carni70.phy,size)
orthogram(size, phylog = carni70.phy)

yrange <- scalewt(carni70$stab[,2])
names(yrange) <- row.names(carni70$stab)
symbols.phylog(carni70.phy,yrange)
orthogram(yrange, phylog = carni70.phy)

s.hist(cbind.data.frame(size, yrange), clabel = 0)

## End(Not run)

```

carniherbi49	<i>Taxonomy, phylogenies and quantitative traits of carnivora and herbivora</i>
--------------	---

Description

This data set describes the taxonomic and phylogenetic relationships of 49 carnivora and herbivora species as reported by Garland and Janis (1993) and Garland et al. (1993). It also gives seven traits corresponding to these 49 species.

Usage

```
data(carniherbi49)
```

Format

carniherbi49 is a list containing the 5 following objects :

- taxo** is a data frame with 49 species and 2 columns : 'fam', a factor family with 14 levels and 'ord', a factor order with 3 levels.
- tre1** is a character string giving the phylogenetic tree in Newick format as reported by Garland et al. (1993).
- tre2** is a character string giving the phylogenetic tree in Newick format as reported by Garland and Janis (1993).
- tab1** is a data frame with 49 species and 2 traits: 'bodymass' (body mass (kg)) and 'homerange' (home range (km)).
- tab2** is a data frame with 49 species and 5 traits: 'clade' (dietary with two levels Carnivore and Herbivore), 'runningspeed' (maximal sprint running speed (km/h)), 'bodymass' (body mass (kg)), 'hindlength' (hind limb length (cm)) and 'mtfratio' (metatarsal/femur ratio).

Source

- Garland, T., Dickerman, A. W., Janis, C. M. and Jones, J. A. (1993) Phylogenetic analysis of covariance by computer simulation. *Systematics Biology*, **42**, 265–292.
- Garland, T. J. and Janis, C.M. (1993) Does metatarsal-femur ratio predict maximal running speed in cursorial mammals? *Journal of Zoology*, **229**, 133–151.

Examples

```
## Not run:
data(carniherbi49)
par(mfrow=c(1,3))
plot(newick2phylog(carniherbi49$tre1), clabel.leaves = 0,
     f.phylog = 2, sub ="article 1")
plot(newick2phylog(carniherbi49$tre2), clabel.leaves = 0,
     f.phylog = 2, sub = "article 2")
taxo <- as.taxo(carniherbi49$taxo)
plot(taxo2phylog(taxo), clabel.nodes = 1.2, clabel.leaves = 1.2)
par(mfrow = c(1,1))

## End(Not run)
```

casitas

Enzymatic polymorphism in Mus musculus

Description

This data set is a data frame with 74 rows (mice) and 15 columns (loci enzymatic polymorphism of the DNA mitochondrial). Each value contains 6 characters coding for two alleles. The missing values are coding by '000000'.

Usage

```
data(casitas)
```

Format

The 74 individuals of casitas belong to 4 groups:

- 1 24 mice of the sub-species *Mus musculus domesticus*
- 2 11 mice of the sub-species *Mus musculus castaneus*
- 3 9 mice of the sub-species *Mus musculus musculus*
- 4 30 mice from a population of the lake Casitas (California)

Source

Exemple du logiciel GENETIX. Belkhir k. et al. GENETIX, logiciel sous Windows™ pour la génétique des populations. Laboratoire Génome, Populations, Interactions CNRS UMR 5000, Université de Montpellier II, Montpellier (France).

<http://www.univ-montp2.fr/~genetix/genetix/genetix.htm>

References

Orth, A., T. Adama, W. Din and F. Bonhomme. (1998) Hybridation naturelle entre deux sous espèces de souris domestique *Mus musculus domesticus* et *Mus musculus castaneus* près de Lake Casitas (Californie). *Genome*, **41**, 104–110.

Examples

```
data(casitas)
casitas.pop <- as.factor(rep(c("dome", "cast", "musc", "casi"),
  c(24,11,9,30)))
table(casitas.pop,casitas[,1])
casi.genet <- char2genet(casitas, casitas.pop)
names(casi.genet)
```

cca

Canonical Correspondence Analysis

Description

Performs a Canonical Correspondence Analysis.

Usage

```
cca(sitspe, sitenv, scannf = TRUE, nf = 2)
```

Arguments

sitspe	a data frame for correspondence analysis, typically a sites x species table
sitenv	a data frame containing variables, typically a sites x environmental variables table
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Value

returns an object of class `pcaiv`. See [pcaiv](#)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Ter Braak, C. J. F. (1986) Canonical correspondence analysis : a new eigenvector technique for multivariate direct gradient analysis. *Ecology*, **67**, 1167–1179.

Ter Braak, C. J. F. (1987) The analysis of vegetation-environment relationships by canonical correspondence analysis. *Vegetatio*, **69**, 69–77.

Chessel, D., Lebreton J. D. and Yoccoz N. (1987) Propriétés de l'analyse canonique des correspondances. Une utilisation en hydrobiologie. *Revue de Statistique Appliquée*, **35**, 55–72.

See Also

[cca](#) in the package `vegan`

Examples

```

data(rpjdl)
millog <- log(rpjdl$mil + 1)
iv1 <- cca(rpjdl$fau, millog, scan = FALSE)
plot(iv1)

# analysis with c1 - as - li -ls
# projections of inertia axes on PCAIV axes
s.corcircle(iv1$as)

# Species positions
s.label(iv1$c1, 2, 1, clab = 0.5, xlim = c(-4,4))
# Sites positions at the weighted mean of present species
s.label(iv1$ls, 2, 1, clab = 0, cpoi = 1, add.p = TRUE)

# Prediction of the positions by regression on environmental variables
s.match(iv1$ls, iv1$li, 2, 1, clab = 0.5)

# analysis with fa - l1 - co -cor
# canonical weights giving unit variance combinations
s.arrow(iv1$fa)

# sites position by environmental variables combinations
# position of species by averaging
s.label(iv1$l1, 2, 1, clab = 0, cpoi = 1.5)
s.label(iv1$co, 2, 1, add.plot = TRUE)

s.distri(iv1$l1, rpjdl$fau, 2, 1, cell = 0, csta = 0.33)
s.label(iv1$co, 2, 1, clab = 0.75, add.plot = TRUE)

# coherence between weights and correlations
par(mfrow = c(1,2))
s.corcircle(iv1$cor, 2, 1)
s.arrow(iv1$fa, 2, 1)
par(mfrow = c(1,1))

```

Description

This data set gives the age, the fecundity and the number of litters for 26 groups of cats.

Usage

```
data(chatcat)
```

Format

chatcat is a list of two objects :

tab is a data frame with 3 factors (age, feco, nport).

eff is a vector of numbers.

Details

One row of tab corresponds to one group of cats.

The value in eff is the number of cats in this group.

Source

Pontier, D. (1984) *Contribution à la biologie et à la génétique des populations de chats domestiques (Felis catus)*. Thèse de 3ème cycle. Université Lyon 1, p. 67.

Examples

```
data(chatcat)
summary(chatcat$tab)
w <- acm.disjonctif(chatcat$tab) # Disjonctive table
names(w) <- c(paste("A", 1:5, sep = ""), paste("B", 1:5, sep = ""),
             paste("C", 1:2, sep = ""))
w <- t(w*chatcat$num)
w <- data.frame(w)
w # BURT table
```

chats

Pair of Variables

Description

This data set is a contingency table of age classes and fecundity classes of cats *Felis catus*.

Usage

```
data(chats)
```

Format

chats is a data frame with 8 rows and 8 columns.\ The 8 rows are age classes (age1, ..., age8)\ The 8 columns are fecundity classes (f0, f12, f34, ..., fcd)\ The values are cats numbers (contingency table).

Source

Legay, J.M. and Pontier, D. (1985) Relation âge-fécondité dans les populations de Chats domestiques, *Felis catus*. *Mammalia*, **49**, 395–402.

Examples

```
data(chats)
chatsw <- data.frame(t(chats))
chatscoa <- dudi.coa(chatsw, scann = FALSE)
par(mfrow = c(2,2))
table.cont(chatsw, abmean.x = TRUE, csi = 2, abline.x = TRUE,
  clabel.r = 1.5, clabel.c = 1.5)
table.cont(chatsw, abmean.y = TRUE, csi = 2, abline.y = TRUE,
  clabel.r = 1.5, clabel.c = 1.5)
table.cont(chatsw, x = chatscoa$c1[,1], y = chatscoa$l1[,1],
  abmean.x = TRUE, csi = 2, abline.x = TRUE, clabel.r = 1.5,
  clabel.c = 1.5)
table.cont(chatsw, x = chatscoa$c1[,1], y = chatscoa$l1[,1],
  abmean.y = TRUE, csi = 2, abline.y = TRUE, clabel.r = 1.5,
  clabel.c = 1.5)
par(mfrow = c(1,1))
```

chazeb

Charolais-Zebus

Description

This data set gives six different weights of 23 charolais and zebu oxen.

Usage

```
data(chazeb)
```

Format

chazeb is a list of 2 components.

tab is a data frame with 23 rows and 6 columns.

cla is a factor with two levels "cha" and "zeb".

Source

Tomassone, R., Danzard, M., Daudin, J. J. and Masson J. P. (1988) *Discrimination et classement*, Masson, Paris. p. 43

Examples

```
data(chazeb)
plot(discrimin(dudi.pca(chazeb$tab, scan = FALSE),
  chazeb$cla, scan = FALSE))
```


chevaine

*Enzymatic polymorphism in Leuciscus cephalus***Description**

This data set contains a list of three components: spatial map, allelic profiles and sample sizes.

Usage

```
data(chevaine)
```

Format

This data set is a list of three components:

tab a data frame with 27 populations and 9 allelic frequencies (4 locus)

coo a list containing all the elements to build a spatial map

eff a numeric containing the numbers of fish samples per station

References

Guinand B., Bouvet Y. and Brohon B. (1996) Spatial aspects of genetic differentiation of the European chub in the Rhone River basin. *Journal of Fish Biology*, **49**, 714–726.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps054.pdf> (in French).

Examples

```
data(chevaine)
'fun.chevaine' <- function(label=TRUE) {
  opar <- par(mar = par("mar"))
  on.exit(par(opar))
  par(mar = c(0.1, 0.1, 0.1, 0.1))
  s.label(chevaine$coo$poi, xlim = c(-20,400), clab = 0, cpoi = 0)
  invisible(lapply(chevaine$coo$lac, polygon, col = "blue", lwd = 2))
  invisible(lapply(chevaine$coo$riv, points, col = "blue", type = "l", lwd = 2))
  if (label){
    s.label(chevaine$coo$poi, clab = 0.75, add.p = TRUE)
    s.label(chevaine$coo$sta, add.p = TRUE, clab = 0.5)
  }
  arrows(200,100,300,100, code = 3, angle = 15, length = 0.2)
  text(250,125,"50 Km")
}

fun.chevaine()

che.genet <- freq2genet(chevaine$tab)
che.pca <- dudi.pca(che.genet$tab, center = che.genet$center, scannf = FALSE, nf = 3)

par(mfrow = c(1,2))
```

```

fun.chevaine(FALSE)
s.value(chevaine$coo$sta, che.pca$li[,1], csi = 2, add.p = TRUE)
fun.chevaine(FALSE)
s.value(chevaine$coo$sta, che.pca$li[,2], csi = 2, add.p = TRUE)

w = prep.fuzzy.var (che.genet$stab, che.genet$loc.blocks)
che.fca <- dudi.fca(w, scannf = FALSE, nf = 3)

fun.chevaine(FALSE)
s.value(chevaine$coo$sta, che.fca$li[,1], csi = 1.5, add.p = TRUE)
fun.chevaine(FALSE)
s.value(chevaine$coo$sta, che.fca$li[,2], csi = 1.5, add.p = TRUE)

```

clementines

Fruit Production

Description

The clementines is a data set containing the fruit production of 20 clementine trees during 15 years.

Usage

```
data(clementines)
```

Format

A data frame with 15 rows and 20 columns

Source

Tisné-Agostini, D. (1988) *Description par analyse en composantes principales de l'évolution de la production du clémentinier en association avec 12 types de porte-greffe*. Rapport technique, DEA Analyse et modélisation des systèmes biologiques, Université Lyon 1.

Examples

```

data(clementines)
op <- par(no.readonly = TRUE)
par(mfrow = c(5,4)) ; par(mar = c(2,2,1,1))
for (i in 1:20) {
  w0 <- 1:15 ; plot(w0, clementines[,i], type = "b")
  abline(lm(clementines[,i] ~ w0))
}
par(op)

pca1 <- dudi.pca(clementines, scan = FALSE)
s.corcircle(pca1$co, clab = 0.75)

```

```

barplot(pca1$li[,1])

op <- par(no.readonly = TRUE)
par(mfrow = c(5,4)) ; par(mar = c(2,2,1,1))
clem0 <- pca1$stab
croi <- 1:15
alter <- c(rep(c(1,-1),7),1)
for (i in 1:20) {
  y <- clem0[,i]
  plot(w0, y, type = "b", ylim = c(-2,2))
  z <- predict(lm(clem0[,i] ~ croi*alter))
  points(w0, z, pch = 20, cex = 2)
  for(j in 1:15) segments (j,y[j],j,z[j])
}
par(op)
par(mfrow = c(1,1))

```

cnc2003

Frequenting movie theaters in France in 2003

Description

cnc2003 is a data frame with 94 rows (94 departments from continental Metropolitan France) and 12 variables.

Usage

```
data(cnc2003)
```

Format

This data frame contains the following variables:

popu is the population department in million inhabitants.

entr is the number of movie theater visitors in million.

rece is the takings from ticket offices.

sean is the number of proposed shows in thousands.

comm is the number of equipped communes in movie theaters (units).

etab is the number of active movie theaters (units).

salle is the number of active screens.

faut is the number of proposed seats.

artes is the number of movie theaters offering "Art and Essay" movies.

multi is the number of active multiplexes.

depart is the name of the department.

reg is the administrative region of the department.

Source

National Center of Cinematography (CNC), september 2003
<http://www.cnc.fr/cncinfo/288/index.html>

See Also

This dataset is compatible with `elec88` and `presid2002`

Examples

```
data(cnc2003)
sco.quant(cnc2003$popu, cnc2003[,2:10], abline = TRUE, csub = 3)
```

coinertia

Coinertia Analysis

Description

The `coinertia` analysis performs a double inertia analysis of two tables.

Usage

```
coinertia(dudiX, dudiY, scannf = TRUE, nf = 2)
## S3 method for class 'coinertia'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'coinertia'
print(x, ...)
## S3 method for class 'coinertia'
summary(object, ...)
```

Arguments

<code>dudiX</code>	a duality diagram providing from one of the functions <code>dudi.coa</code> , <code>dudi.pca</code> , ...
<code>dudiY</code>	a duality diagram providing from one of the functions <code>dudi.coa</code> , <code>dudi.pca</code> , ...
<code>scannf</code>	a logical value indicating whether the eigenvalues bar plot should be displayed
<code>nf</code>	if <code>scannf</code> <code>FALSE</code> , an integer indicating the number of kept axes
<code>x</code> , <code>object</code>	an object of class <code>'coinertia'</code>
<code>xax</code> , <code>yax</code>	the numbers of the x-axis and the y-axis
...	further arguments passed to or from other methods

Value

Returns a list of class 'coinertia', sub-class 'dudi' containing:

call	call
rank	rank
nf	a numeric value indicating the number of kept axes
RV	a numeric value, the RV coefficient
eig	a numeric vector with all the eigenvalues
lw	a numeric vector with the rows weights (crossed table)
cw	a numeric vector with the columns weights (crossed table)
tab	a crossed table (CT)
li	CT row scores (cols of dudiY)
l1	Principal components (loadings for cols of dudiY)
co	CT col scores (cols of dudiX)
c1	Principal axes (cols of dudiX)
lX	Row scores (rows of dudiX)
mX	Normed row scores (rows of dudiX)
lY	Row scores (rows of dudiY)
mY	Normed row scores (rows of dudiY)
aX	Correlations between dudiX axes and coinertia axes
aY	Correlations between dudiY axes and coinertia axes

WARNING

IMPORTANT : dudi1 and dudi2 must have identical row weights.

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Dolédec, S. and Chessel, D. (1994) Co-inertia analysis: an alternative method for studying species-environment relationships. *Freshwater Biology*, **31**, 277–294.

Dray, S., Chessel, D. and J. Thioulouse (2003) Co-inertia analysis and the linking of the ecological data tables. *Ecology*, **84**, 11, 3078–3089.

Examples

```

data(doubs)
dudi1 <- dudi.pca(doubs$env, scale = TRUE, scan = FALSE, nf = 3)
dudi2 <- dudi.pca(doubs$fish, scale = FALSE, scan = FALSE, nf = 2)
coin1 <- coinertia(dudi1,dudi2, scan = FALSE, nf = 2)
s.arrow(coin1$l1, clab = 0.7)

s.arrow(coin1$c1, clab = 0.7)

par(mfrow = c(1,2))
s.corcircle(coin1$aX)
s.corcircle(coin1$aY)
par(mfrow = c(1,1))

coin1
summary(coin1)
plot(coin1)

```

 coleo

Table of Fuzzy Biological Traits

Description

This data set coleo (coleoptera) is a fuzzy biological traits table.

Usage

```
data(coleo)
```

Format

coleo is a list of 5 components.

tab is a data frame with 110 rows (species) and 32 columns (categories).

species.names is a vector of species names.

moda.names is a vector of fuzzy variables names.

families is a factor species family.

col.blocks is a vector containing the number of categories of each trait.

Source

Bournaud, M., Richoux, P. and Usseglio-Polatera, P. (1992) An approach to the synthesis of qualitative ecological information from aquatic coleoptera communities. *Regulated rivers: Research and Management*, **7**, 165–180.

Examples

```

data(coleo)
op <- par(no.readonly = TRUE)
coleo.fuzzy <- prep.fuzzy.var(coleo$tab, coleo$col.blocks)
fca1 <- dudi.fca(coleo.fuzzy, sca = FALSE, nf = 3)
par(mfrow = c(3,3))
indica <- factor(rep(names(coleo$col), coleo$col))
for (j in levels(indica)) s.distri (fca1$11,
  coleo$tab[,which(indica==j)], clab = 1.5, sub = as.character(j),
  cell = 0, csta = 0.5, csub = 3,
  label = coleo$moda.names[which(indica == j)])
par(op)
par(mfrow = c(1,1))

```

combine.4thcorner

Functions to combine and adjust the outputs 3-table methods

Description

Functions to combine and adjust the outputs of the `fourthcorner` and `randtest.rlq` functions created using permutational models 2 and 4 (sequential approach).

Usage

```

combine.randtest.rlq(obj1, obj2)
combine.4thcorner(four1, four2)
p.adjust.4thcorner(x, p.adjust.method.G = p.adjust.methods,
p.adjust.method.D = p.adjust.methods, p.adjust.D = c("global",
"levels"))

```

Arguments

<code>four1</code>	an object of the class <code>4thcorner</code> created with <code>modeltype = 2</code> (or 4)
<code>four2</code>	an object of the class <code>4thcorner</code> created with <code>modeltype = 4</code> (or 2)
<code>obj1</code>	an object created with <code>randtest.rlq</code> and <code>modeltype = 2</code> (or 4)
<code>obj2</code>	an object created with <code>randtest.rlq</code> and <code>modeltype = 4</code> (or 2)
<code>x</code>	an object of the class <code>4thcorner</code>
<code>p.adjust.method.G</code>	a string indicating a method for multiple adjustment used for output <code>tabG</code> , see p.adjust.methods for possible choices
<code>p.adjust.method.D</code>	a string indicating a method for multiple adjustment used for output <code>tabD/tabD2</code> , see p.adjust.methods for possible choices
<code>p.adjust.D</code>	a string indicating if multiple adjustment for <code>tabD/tabD2</code> should be done globally or only between levels of a factor ("levels", as in the original paper of Legendre et al. 1997)

Details

The functions combines the outputs of two objects (created by `fourthcorner` and `randtest.rlq` functions) as described in Dray and Legendre (2008) and ter Braak et al (2012).

Value

The functions return objects of the same class than their argument. They simply create a new object where pvalues are equal to the maximum of pvalues of the two arguments.

Author(s)

Stephane Dray <stephane.drays@univ-lyon1.fr>

References

- Dray, S. and Legendre, P. (2008) Testing the species traits-environment relationships: the fourth-corner problem revisited. *Ecology*, **89**, 3400–3412.
- ter Braak, C., Cormont, A., and Dray, S. (2012) Improved testing of species traits-environment relationships in the fourth corner problem. *Ecology*, **93**, 1525–1526.

See Also

[rlq](#), [fourthcorner](#), [p.adjust.methods](#)

Examples

```
data(aravo)
four2 <- fourthcorner(aravo$env, aravo$spe, aravo$traits, nrepet=99,modeltype=2)
four4 <- fourthcorner(aravo$env, aravo$spe, aravo$traits, nrepet=99,modeltype=4)
four.comb <- combine.4thcorner(four2,four4)
## or directly :
## four.comb <- fourthcorner(aravo$env, aravo$spe, aravo$traits, nrepet=99,modeltype=6)
summary(four.comb)
plot(four.comb, stat = "G")
```

Description

The `mantelkdist` and `RVkdist` functions apply to blocks of distance matrices the `mantel.rtest` and `RV.rtest` functions.

Usage

```

mantelkdist (kd, nrepet = 999)
RVkdist (kd, nrepet = 999)
## S3 method for class 'corkdist'
plot(x, whichinrow = NULL, whichincol = NULL,
     gap = 4, nclass = 10, coeff = 1,...)

```

Arguments

kd	a list of class kdist
nrepet	the number of permutations
x	an objet of class corkdist, coming from RVkdist or mantelkdist
whichinrow	a vector of integers to select the graphs in rows (if NULL all the graphs are computed)
whichincol	a vector of integers to select the graphs in columns (if NULL all the graphs are computed)
gap	an integer to determinate the space between two graphs
nclass	a number of intervals for the histogram
coeff	an integer to fit the magnitude of the graph
...	further arguments passed to or from other methods

Details

The corkdist class has some generic functions `print`, `plot` and `summary`. The plot shows bivariate scatterplots between semi-matrices of distances or histograms of simulated values with an error position.

Value

a list of class corkdist containing for each pair of distances an object of class randtest (permutation tests).

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```

data(friday87)
fri.w <- ktab.data.frame(friday87$fau, friday87$fau.blo,
                       tabnames = friday87$tab.names)
fri.kc <- lapply(1:10, function(x) dist.binary(fri.w[[x]],10))
names(fri.kc) <- substr(friday87$tab.names,1,4)
fri.kd <- kdist(fri.kc)
fri.mantel = mantelkdist(kd = fri.kd, nrepet = 999)
plot(fri.mantel,1:5,1:5)

```

```

plot(fri.mantel,1:5,6:10)
plot(fri.mantel,6:10,1:5)
plot(fri.mantel,6:10,6:10)
s.corcircle (dudi.pca(as.data.frame(fri.kd), scan = FALSE)$co)
plot(RVkdust(fri.kd),1:5,1:5)

data(yanomama)
m1 <- mantelkdist(kdist(yanomama),999)
m1
summary(m1)
plot(m1)

```

corvus

Corvus morphology

Description

This data set gives a morphological description of 28 species of the genus *Corvus* split in two habitat types and phylogeographic stocks.

Usage

```
data(corvus)
```

Format

corvus is data frame with 28 observations (the species) and 4 variables :

wing : wing length (mm)

bill : bill length (mm)

habitat : habitat with two levels clos and open

phylog : phylogeographic stock with three levels amer(America), orien(Oriental-Australian), pale(Paleoartic-African)

References

Laiolo, P. and Rolando, A. (2003) The evolution of vocalisations in the genus *Corvus*: effects of phylogeny, morphology and habitat. *Evolutionary Ecology*, **17**, 111–123.

Examples

```

data(corvus)
plot(corvus[,1:2])
s.class(corvus[,1:2], corvus[,4]:corvus[,3], add.p = TRUE)

```

costatis	<i>STATIS and Co-Inertia : Analysis of a series of paired ecological tables</i>
----------	---

Description

Does the analysis of a series of pairs of ecological tables. This function uses Partial Triadic Analysis ([pta](#)) and [coinertia](#) to do the computations.

Usage

```
costatis(KTX, KTY, scannf = TRUE)
```

Arguments

KTX	an objet of class ktab
KTY	an objet of class ktab
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed

Details

This function takes 2 ktabs. It does a PTA (partial triadic analysis: [pta](#)) on each ktab, and does a coinertia analysis ([coinertia](#)) on the compromises of the two PTAs.

Value

a list of class coinertia, subclass dudi. See [coinertia](#)

WARNING

IMPORTANT : KTX and KTY must have the same k-tables structure, the same number of columns, and the same column weights.

Author(s)

Jean Thioulouse <jean.thioulouse@univ-lyon1.fr>

References

Thioulouse J., Simier M. and Chessel D. (2004). Simultaneous analysis of a sequence of paired ecological tables. *Ecology* **85**, 272-283..

Simier, M., Blanc L., Pellegrin F., and Nandris D. (1999). Approche simultanée de K couples de tableaux : Application a l'étude des relations pathologie végétale - environnement. *Revue de Statistique Appliquée*, **47**, 31-46.

Examples

```

data(meau)
wit1 <- withinpca(meau$env, meau$design$season, scan = FALSE, scal = "total")
pcaspe <- dudi.pca(meau$spe, scale = FALSE, scan = FALSE, nf = 2)
wit2 <- wca(pcaspe, meau$design$season, scan = FALSE, nf = 2)
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5", "S6"), 4))
kta2 <- ktab.within(wit2, colnames = rep(c("S1", "S2", "S3", "S4", "S5", "S6"), 4))
costatis1 <- costatis(kta1, kta2, scan = FALSE)
plot(costatis1)

```

Deprecated functions *Deprecated functions in ade4*

Description

The functions/data listed below are deprecated:

- orthogram: replaced by orthogram in adephylo.

deug *Exam marks for some students*

Description

This data set gives the exam results of 104 students in the second year of a French University onto 9 subjects.

Usage

```
data(deug)
```

Format

deug is a list of three components.

tab is a data frame with 104 students and 9 subjects : Algebra, Analysis, Proba, Informatic, Economy, Option1, Option2, English, Sport.

result is a factor of 104 components giving the final exam levels (A+, A, B, B-, C-, D).

cent is a vector of required marks by subject to get exactly 10/20 with a coefficient.

Source

University of Lyon 1

Examples

```
data(deug)
# decentred PCA
pca1 <- dudi.pca(deug$tab, scal = FALSE, center = deug$cent,
  scan = FALSE)
s.class(pca1$li, deug$result)
s.arrow(40 * pca1$c1, add.plot = TRUE)
```

disc *Rao's dissimilarity coefficient*

Description

Calculates the root square of Rao's dissimilarity coefficient between samples.

Usage

```
disc(samples, dis = NULL, structures = NULL)
```

Arguments

samples	a data frame with elements as rows, samples as columns, and abundance, presence-absence or frequencies as entries
dis	an object of class <code>dist</code> containing distances or dissimilarities among elements. If <code>dis</code> is <code>NULL</code> , equidistances are used.
structures	a data frame containing, in the <i>j</i> th row and the <i>k</i> th column, the name of the group of level <i>k</i> to which the <i>j</i> th population belongs.

Value

Returns a list of objects of class `dist`

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Rao, C.R. (1982) Diversity and dissimilarity coefficients: a unified approach. *Theoretical Population Biology*, **21**, 24–43.

Examples

```

data(humDNAm)
humDNA.dist <- disc(humDNAm$samples, sqrt(humDNAm$distances), humDNAm$structures)
humDNA.dist
is.euclid(humDNA.dist$samples)
is.euclid(humDNA.dist$regions)

## Not run:
data(ecomor)
dtaxo <- dist.taxo(ecomor$taxo)
ecomor.dist <- disc(ecomor$habitat, dtaxo)
ecomor.dist
is.euclid(ecomor.dist)

## End(Not run)

```

discrimin

Linear Discriminant Analysis (descriptive statistic)

Description

performs a linear discriminant analysis.

Usage

```

discrimin(dudi, fac, scannf = TRUE, nf = 2)
## S3 method for class 'discrimin'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'discrimin'
print(x, ...)

```

Arguments

dudi	a duality diagram, object of class dudi
fac	a factor defining the classes of discriminant analysis
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
x	an object of class 'discrimin'
xax	the column number of the x-axis
yax	the column number of the y-axis
...	further arguments passed to or from other methods

Value

returns a list of class 'discrimin' containing :

nf	a numeric value indicating the number of kept axes
eig	a numeric vector with all the eigenvalues
fa	a matrix with the loadings: the canonical weights
li	a data frame which gives the canonical scores
va	a matrix which gives the cosines between the variables and the canonical scores
cp	a matrix which gives the cosines between the components and the canonical scores
gc	a data frame which gives the class scores

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

See Also

lda in package MASS

Examples

```
data(chazeb)
dis1 <- discrimin(dudi.pca(chazeb$tab, scan = FALSE), chazeb$cla,
  scan = FALSE)
dis1
plot(dis1)

data(skulls)
plot(discrimin(dudi.pca(skulls, scan = FALSE), g1(5,30),
  scan = FALSE))
```

discrimin.coa

Discriminant Correspondence Analysis

Description

performs a discriminant correspondence analysis.

Usage

```
discrimin.coa(df, fac, scannf = TRUE, nf = 2)
```

Arguments

df	a data frame containing positive or null values
fac	a factor defining the classes of discriminant analysis
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Value

a list of class discrimin. See [discrimin](#)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Perriere, G., Lobry, J. R. and Thioulouse J. (1996) Correspondence discriminant analysis: a multi-variate method for comparing classes of protein and nucleic acid sequences. *CABIOS*, **12**, 519–524.

Perriere, G. and Thioulouse, J. (2003) Use of Correspondence Discriminant Analysis to predict the subcellular location of bacterial proteins. *Computer Methods and Programs in Biomedicine*, **70**, 2, 99–105.

Examples

```
data(perthi02)
plot(discrimin.coa(perthi02$tab, perthi02$cla, scan = FALSE))
```

dist.binary

Computation of Distance Matrices for Binary Data

Description

computes for binary data some distance matrice.

Usage

```
dist.binary(df, method = NULL, diag = FALSE, upper = FALSE)
```


Arguments

df	a matrix or a data frame with positive or null numeric values. Used with <code>as.matrix(1 * (df > 0))</code>
method	an integer between 1 and 10 . If NULL the choice is made with a console message. See details
diag	a logical value indicating whether the diagonal of the distance matrix should be printed by 'print.dist'
upper	a logical value indicating whether the upper triangle of the distance matrix should be printed by 'print.dist'

Details

Let be the contingency table of binary data such as $n_{11} = a$, $n_{10} = b$, $n_{01} = c$ and $n_{00} = d$. All these distances are of type $d = \sqrt{1 - s}$ with s a similarity coefficient.

1 = Jaccard index (1901) S3 coefficient of Gower & Legendre $s_1 = \frac{a}{a+b+c}$

2 = Simple matching coefficient of Sokal & Michener (1958) S4 coefficient of Gower & Legendre $s_2 = \frac{a+d}{a+b+c+d}$

3 = Sokal & Sneath(1963) S5 coefficient of Gower & Legendre $s_3 = \frac{a}{a+2(b+c)}$

4 = Rogers & Tanimoto (1960) S6 coefficient of Gower & Legendre $s_4 = \frac{a+d}{(a+2(b+c)+d)}$

5 = Dice (1945) or Sorensen (1948) S7 coefficient of Gower & Legendre $s_5 = \frac{2a}{2a+b+c}$

6 = Hamann coefficient S9 index of Gower & Legendre (1986) $s_6 = \frac{a-(b+c)+d}{a+b+c+d}$

7 = Ochiai (1957) S12 coefficient of Gower & Legendre $s_7 = \frac{a}{\sqrt{(a+b)(a+c)}}$

8 = Sokal & Sneath (1963) S13 coefficient of Gower & Legendre $s_8 = \frac{ad}{\sqrt{(a+b)(a+c)(d+b)(d+c)}}$

9 = Phi of Pearson S14 coefficient of Gower & Legendre $s_9 = \frac{ad-bc}{\sqrt{(a+b)(a+c)(b+d)(d+c)}}$

10 = S2 coefficient of Gower & Legendre $s_{10} = \frac{a}{a+b+c+d}$

Value

returns a distance matrix of class `dist` between the rows of the data frame

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

Gower, J.C. and Legendre, P. (1986) Metric and Euclidean properties of dissimilarity coefficients. *Journal of Classification*, **3**, 5–48.

Examples

```
data(aviurba)
for (i in 1:10) {
  d <- dist.binary(aviurba$fau, method = i)
  cat(attr(d, "method"), is.euclid(d), "\n")}
```

`dist.dudi`*Computation of the Distance Matrix from a Statistical Triplet*

Description

computes for a statistical triplet a distance matrix.

Usage

```
dist.dudi(dudi, amongrow = TRUE)
```

Arguments

<code>dudi</code>	a duality diagram, object of class <code>dudi</code>
<code>amongrow</code>	a logical value computing the distance if TRUE, between rows, if FALSE between columns.

Value

an object of class `dist`

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE)
sum((dist(scalewt(meaudret$env)) - dist.dudi(pca1))^2)
#[1] 4.045e-29 the same thing
```

dist.genet

*Genetic distances from gene frequencies***Description**

This program computes any one of five measures of genetic distance from a set of gene frequencies in different populations with several loci.

Usage

```
dist.genet(genet, method = 1, diag = FALSE, upper = FALSE)
```

Arguments

genet	a list of class genet
method	an integer between 1 and 5. See details
diag	a logical value indicating whether the diagonal of the distance matrix should be printed by print.dist
upper	a logical value indicating whether the upper triangle of the distance matrix should be printed by print.dist

Details

Let **A** a table containing allelic frequencies with t populations (rows) and m alleles (columns).

Let ν the number of loci. The locus j gets $m(j)$ alleles. $m = \sum_{j=1}^{\nu} m(j)$

For the row i and the modality k of the variable j , notice the value a_{ij}^k ($1 \leq i \leq t$, $1 \leq j \leq \nu$, $1 \leq k \leq m(j)$) the value of the initial table.

$$a_{ij}^+ = \sum_{k=1}^{m(j)} a_{ij}^k \text{ and } p_{ij}^k = \frac{a_{ij}^k}{a_{ij}^+}$$

Let **P** the table of general term p_{ij}^k

$$p_{ij}^+ = \sum_{k=1}^{m(j)} p_{ij}^k = 1, p_{i+}^+ = \sum_{j=1}^{\nu} p_{ij}^+ = \nu, p_{++}^+ = \sum_{j=1}^{\nu} p_{i+}^+ = t\nu$$

The option method computes the distance matrices between populations using the frequencies p_{ij}^k .

1. Nei's distance:

$$D_1(a, b) = -\ln\left(\frac{\sum_{k=1}^{\nu} \sum_{j=1}^{m(k)} p_{aj}^k p_{bj}^k}{\sqrt{\sum_{k=1}^{\nu} \sum_{j=1}^{m(k)} (p_{aj}^k)^2} \sqrt{\sum_{k=1}^{\nu} \sum_{j=1}^{m(k)} (p_{bj}^k)^2}}\right)$$

2. Angular distance or Edwards' distance:

$$D_2(a, b) = \sqrt{1 - \frac{1}{\nu} \sum_{k=1}^{\nu} \sum_{j=1}^{m(k)} \sqrt{p_{aj}^k p_{bj}^k}}$$

3. Coancestrality coefficient or Reynolds' distance:

$$D_3(a, b) = \sqrt{\frac{\sum_{k=1}^{\nu} \sum_{j=1}^{m(k)} (p_{aj}^k - p_{bj}^k)^2}{2 \sum_{k=1}^{\nu} (1 - \sum_{j=1}^{m(k)} p_{aj}^k p_{bj}^k)}}$$

4. Classical Euclidean distance or Rogers' distance:

$$D_4(a, b) = \frac{1}{\nu} \sum_{k=1}^{\nu} \sqrt{\frac{1}{2} \sum_{j=1}^{m(k)} (p_{aj}^k - p_{bj}^k)^2}$$

5. Absolute genetics distance or Provesti 's distance:

$$D_5(a, b) = \frac{1}{2\nu} \sum_{k=1}^{\nu} \sum_{j=1}^{m(k)} |p_{aj}^k - p_{bj}^k|$$

Value

returns a distance matrix of class `dist` between the rows of the data frame

Author(s)

Daniel Chessel

Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

To complete informations about distances:

Distance 1:

Nei, M. (1972) Genetic distances between populations. *American Naturalist*, **106**, 283–292.

Nei M. (1978) Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics*, **23**, 341–369.

Avise, J. C. (1994) Molecular markers, natural history and evolution. Chapman & Hall, London.

Distance 2:

Edwards, A.W.F. (1971) Distance between populations on the basis of gene frequencies. *Biometrics*, **27**, 873–881.

Cavalli-Sforza L.L. and Edwards A.W.F. (1967) Phylogenetic analysis: models and estimation procedures. *Evolution*, **32**, 550–570.

Hartl, D.L. and Clark, A.G. (1989) Principles of population genetics. Sinauer Associates, Sunderland, Massachusetts (p. 303).

Distance 3:

Reynolds, J. B., B. S. Weir, and C. C. Cockerham. (1983) Estimation of the coancestry coefficient: basis for a short-term genetic distance. *Genetics*, **105**, 767–779.

Distance 4:

Rogers, J.S. (1972) Measures of genetic similarity and genetic distances. *Studies in Genetics*, Univ. Texas Publ., **7213**, 145–153.

Avise, J. C. (1994) Molecular markers, natural history and evolution. Chapman & Hall, London.

Distance 5:

Prevosti A. (1974) La distancia genética entre poblaciones. *Miscellanea Alcobé*, **68**, 109–118.

Prevosti A., Ocaña J. and Alonso G. (1975) Distances between populations of *Drosophila subobscura*, based on chromosome arrangements frequencies. *Theoretical and Applied Genetics*, **45**,

231–241.

To find some useful explanations:

Sanchez-Mazas A. (2003) Cours de Génétique Moléculaire des Populations. Cours VIII Distances génétiques - Représentation des populations.

http://anthro.unige.ch/GMDP/Alicia/GMDP_dist.htm

Examples

```
data(casitas)
casi.genet <- char2genet(casitas,
  as.factor(rep(c("dome", "cast", "musc", "casi"), c(24,11,9,30))))
ldist <- lapply(1:5, function(method) dist.genet(casi.genet,method))
ldist
unlist(lapply(ldist, is.euclid))
kdist(ldist)
```

dist.ktab

Mixed-variables coefficient of distance

Description

The mixed-variables coefficient of distance generalizes Gower's general coefficient of distance to allow the treatment of various statistical types of variables when calculating distances. This is especially important when measuring functional diversity. Indeed, most of the indices that measure functional diversity depend on variables (traits) that have various statistical types (e.g. circular, fuzzy, ordinal) and that go through a matrix of distances among species.

Usage

```
dist.ktab(x, type, option = c("scaledBYrange", "scaledBYsd", "noscale"),
  scann = FALSE, tol = 1e-8)
ldist.ktab(x, type, option = c("scaledBYrange", "scaledBYsd",
  "noscale"), scann = FALSE, tol = 1e-8)
kdist.cor(x, type, option = c("scaledBYrange", "scaledBYsd", "noscale"),
  scann = FALSE, tol = 1e-8, squared = TRUE)
prep.fuzzy(df, col.blocks, row.w = rep(1, nrow(df)), labels = paste("F",
  1:length(col.blocks), sep = ""))
prep.binary(df, col.blocks, labels = paste("B", 1:length(col.blocks), sep = ""))
prep.circular(df, rangemin = apply(df, 2, min, na.rm = TRUE), rangemax =
  apply(df, 2, max, na.rm = TRUE))
```

Arguments

x Object of class ktab (see details)

type	Vector that provide the type of each table in x. The possible types are "Q" (quantitative), "O" (ordinal), "N" (nominal), "D" (dichotomous), "F" (fuzzy, or expressed as a proportion), "B" (multichoice nominal variables, coded by binary columns), "C" (circular). Values in type must be in the same order as in x.
option	A string that can have three values: either "scaledBYrange" if the quantitative variables must be scaled by their range, or "scaledBYsd" if they must be scaled by their standard deviation, or "noscale" if they should not be scaled. This last option can be useful if the the values have already been normalized by the known range of the whole population instead of the observed range measured on the sample. If x contains data from various types, then the option "scaledBYsd" is not suitable (a warning will appear if the option selected with that condition).
scann	A logical. If TRUE, then the user will have to choose among several possible functions of distances for the quantitative, ordinal, fuzzy and binary variables.
tol	A tolerance threshold: a value less than tol is considered as null.
squared	A logical, if TRUE, the squared distances are considered.
df	Objet of class data.frame
col.blocks	A vector that contains the number of levels per variable (in the same order as in df)
row.w	A vector of row weigths
labels	the names of the traits
rangemin	A numeric corresponding to the smallest level where the loop starts
rangemax	A numeric corresponding to the highest level where the loop closes

Value

The functions provide the following results:

dist.ktab	returns an object of class dist;
ldist.ktab	returns a list of objects of class dist that correspond to the distances between species calculated per trait;
kdist.cor	returns a list of three objects: "paircov" provides the covariance between traits in terms of (squared) distances between species; "paircor" provides the correlations between traits in terms of (squared) distances between species; "glocor" provides the correlations between the (squared) distances obtained for each trait and the global (squared) distances obtained by mixing all the traits (= contributions of traits to the global distances);
prep.binary and prep.fuzzy	returns a data frame with the following attributes: col.blocks specifies the number of columns per fuzzy variable; col.num specifies which variable each column belongs to;
prep.circular	returns a data frame with the following attributes: max specifies the number of levels in each circular variable.

Author(s)

Sandrine Pavoine <pavoine@mnhn.fr>

References

Pavoine S., Vallet, J., Dufour, A.-B., Gachet, S. and Daniel, H. (2009) On the challenge of treating various types of variables: Application for improving the measurement of functional diversity. *Oikos*, **118**, 391–402.

See Also

[daisy](#) in the `cluster` package in the case of ratio-scale (quantitative) and nominal variables; and [woangers](#) for an application.

Examples

```
# With fuzzy variables
data(bsetal97)

w <- prep.fuzzy(bsetal97$biol, bsetal97$biol.blo)
w[1:6, 1:10]
ktab1 <- ktab.list.df(list(w))
dis <- dist.ktab(ktab1, type = "F")
as.matrix(dis)[1:5, 1:5]

## Not run:
# With ratio-scale and multichoice variables
data(ecomor)

wM <- log(ecomor$morpho + 1) # Quantitative variables
wD <- ecomor$diet
# wD is a data frame containing a multichoice nominal variable
# (diet habit), with 8 modalities (Granivorous, etc)
# We must prepare it by prep.binary
head(wD)
wD <- prep.binary(wD, col.blocks = 8, label = "diet")
wF <- ecomor$forsub
# wF is also a data frame containing a multichoice nominal variable
# (foraging substrat), with 6 modalities (Foliage, etc)
# We must prepare it by prep.binary
head(wF)
wF <- prep.binary(wF, col.blocks = 6, label = "foraging")
# Another possibility is to combine the two last data frames wD and wF as
# they contain the same type of variables
wB <- cbind.data.frame(ecomor$diet, ecomor$forsub)
head(wB)
wB <- prep.binary(wB, col.blocks = c(8, 6), label = c("diet", "foraging"))
# The results given by the two alternatives are identical
ktab2 <- ktab.list.df(list(wM, wD, wF))
disecomor <- dist.ktab(ktab2, type= c("Q", "B", "B"))
as.matrix(disecomor)[1:5, 1:5]
contrib2 <- kdist.cor(ktab2, type= c("Q", "B", "B"))
contrib2

ktab3 <- ktab.list.df(list(wM, wB))
disecomor2 <- dist.ktab(ktab3, type= c("Q", "B"))
```

```

as.matrix(disecomor2)[1:5, 1:5]
contrib3 <- kdist.cor(ktab3, type= c("Q", "B"))
contrib3

# With a range of variables
data(woangers)

traits <- woangers$traits
# Nominal variables 'li', 'pr', 'lp' and 'le'
# (see table 1 in the main text for the codes of the variables)
tabN <- traits[,c(1:2, 7, 8)]
# Circular variable 'fo'
tabC <- traits[3]
tabCp <- prep.circular(tabC, 1, 12)
# The levels of the variable lie between 1 (January) and 12 (December).
# Ordinal variables 'he', 'ae' and 'un'
tabO <- traits[, 4:6]
# Fuzzy variables 'mp', 'pe' and 'di'
tabF <- traits[, 9:19]
tabFp <- prep.fuzzy(tabF, c(3, 3, 5), labels = c("mp", "pe", "di"))
# 'mp' has 3 levels, 'pe' has 3 levels and 'di' has 5 levels.
# Quantitative variables 'lo' and 'lf'
tabQ <- traits[, 20:21]
ktab1 <- ktab.list.df(list(tabN, tabCp, tabO, tabFp, tabQ))
distrat <- dist.ktab(ktab1, c("N", "C", "O", "F", "Q"))
is.euclid(distrat)
contrib <- kdist.cor(ktab1, type = c("N", "C", "O", "F", "Q"))
contrib
dotchart(sort(contrib$glocor), labels = rownames(contrib$glocor)[order(contrib$glocor[, 1])])

## End(Not run)

```

dist.neig

Computation of the Distance Matrix associated to a Neighbouring Graph

Description

This distance matrix between two points is the length of the shortest path between these points.

Usage

```
dist.neig(neig)
```

Arguments

neig a neighbouring graph, object of class neig

Value

returns a distance matrix, object of class dist

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
data(elec88)
d0 <- dist.neig(elec88$neig)
plot(dist(elec88$xy),d0)
```

 dist.prop

Computation of Distance Matrices of Percentage Data

Description

computes for percentage data some distance matrices.

Usage

```
dist.prop(df, method = NULL, diag = FALSE, upper = FALSE)
```

Arguments

df a data frame containing only positive or null values, used as row percentages

method an integer between 1 and 5. If NULL the choice is made with a console message. See details

diag a logical value indicating whether the diagonal of the distance matrix should be printed by 'print.dist'

upper a logical value indicating whether the upper triangle of the distance matrix should be printed by 'print.dist'

Details

$$1 = \text{Manly } d_1 = \frac{1}{2} \sum_{i=1}^K |p_i - q_i|$$

$$2 = \text{Overlap index Manly } d_2 = 1 - \frac{\sum_{i=1}^K p_i q_i}{\sqrt{\sum_{i=1}^K p_i^2} \sqrt{\sum_{i=1}^K q_i^2}}$$

$$3 = \text{Rogers 1972 (one locus) } d_3 = \sqrt{\frac{1}{2} \sum_{i=1}^K (p_i - q_i)^2}$$

$$4 = \text{Nei 1972 (one locus) } d_4 = \ln \frac{\sum_{i=1}^K p_i q_i}{\sqrt{\sum_{i=1}^K p_i^2} \sqrt{\sum_{i=1}^K q_i^2}}$$

$$5 = \text{Edwards 1971 (one locus) } d_5 = \sqrt{1 - \sum_{i=1}^K \sqrt{p_i q_i}}$$

Value

returns a distance matrix, object of class `dist`

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

- Edwards, A. W. F. (1971) Distance between populations on the basis of gene frequencies. *Biometrics*, **27**, 873–881.
- Manly, B. F. (1994) *Multivariate Statistical Methods. A primer*, Second edition. Chapman & Hall, London.
- Nei, M. (1972) Genetic distances between populations. *The American Naturalist*, **106**, 283–292.

Examples

```
data(microsatt)
w <- microsatt$tab[1:microsatt$loci.eff[1]]
par(mfrow = c(2,2))
scatter(dudi.pco(lingoes(dist.prop(w,1)), scann = FALSE))
scatter(dudi.pco(lingoes(dist.prop(w,2)), scann = FALSE))
scatter(dudi.pco(dist.prop(w,3), scann = FALSE))
scatter(dudi.pco(lingoes(dist.prop(w,4)), scann = FALSE))
par(mfrow = c(1,1))
```

dist.quant

Computation of Distance Matrices on Quantitative Variables

Description

computes on quantitative variables, some distance matrices as canonical, Joreskog and Mahalanobis.

Usage

```
dist.quant(df, method = NULL, diag = FALSE, upper = FALSE,
           tol = 1e-07)
```

Arguments

- `df` a data frame containing only quantitative variables
- `method` an integer between 1 and 3. If NULL the choice is made with a console message. See details
- `diag` a logical value indicating whether the diagonal of the distance matrix should be printed by 'print.dist'

upper	a logical value indicating whether the upper triangle of the distance matrix should be printed by 'print.dist'
tol	used in case 3 of method as a tolerance threshold for null eigenvalues

Details

All the distances are of type $d = \|x - y\|_A = \sqrt{(x - y)^t A (x - y)}$

1 = Canonical $A = \text{Identity}$

2 = Joreskog $A = \frac{1}{\text{diag}(\text{cov})}$

3 = Mahalanobis $A = \text{inv}(\text{cov})$

Value

an object of class `dist`

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
data(ecomor)
par(mfrow = c(2,2))
scatter(dudi.pco(dist.quant(ecomor$morpho,3), scan = FALSE))
scatter(dudi.pco(dist.quant(ecomor$morpho,2), scan = FALSE))
scatter(dudi.pco(dist(scalewt(ecomor$morpho)), scan = FALSE))
scatter(dudi.pco(dist.quant(ecomor$morpho,1), scan = FALSE))
par(mfrow = c(1,1))
```

divc	<i>Rao's diversity coefficient also called quadratic entropy</i>
------	--

Description

Calculates Rao's diversity coefficient within samples.

Usage

```
divc(df, dis, scale)
```

Arguments

df	a data frame with elements as rows, samples as columns, and abundance, presence-absence or frequencies as entries
dis	an object of class <code>dist</code> containing distances or dissimilarities among elements. If <code>dis</code> is <code>NULL</code> , Gini-Simpson index is performed.
scale	a logical value indicating whether or not the diversity coefficient should be scaled by its maximal value over all frequency distributions.

Value

Returns a data frame with samples as rows and the diversity coefficient within samples as columns

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Rao, C.R. (1982) Diversity and dissimilarity coefficients: a unified approach. *Theoretical Population Biology*, **21**, 24–43.

Gini, C. (1912) Variabilit'a e mutabilit'a. *Universite di Cagliari III*, Parte II.

Simpson, E.H. (1949) Measurement of diversity. *Nature*, **163**, 688.

Champely, S. and Chessel, D. (2002) Measuring biological diversity using Euclidean metrics. *Environmental and Ecological Statistics*, **9**, 167–177.

Examples

```
data(ecomor)
dtaxo <- dist.taxo(ecomor$taxo)
divc(ecomor$habitat, dtaxo)

data(humDNAm)
divc(humDNAm$samples, sqrt(humDNAm$distances))
```

divcmax	<i>Maximal value of Rao's diversity coefficient also called quadratic entropy</i>
---------	---

Description

For a given dissimilarity matrix, this function calculates the maximal value of Rao's diversity coefficient over all frequency distribution. It uses an optimization technique based on Rosen's projection gradient algorithm and is verified using the Kuhn-Tucker conditions.

Usage

```
divcmax(dis, epsilon, comment)
```

Arguments

dis	an object of class <code>dist</code> containing distances or dissimilarities among elements.
epsilon	a tolerance threshold : a frequency is non null if it is higher than epsilon.
comment	a logical value indicating whether or not comments on the optimization technique should be printed.

Value

Returns a list

value	the maximal value of Rao's diversity coefficient.
vectors	a data frame containing four frequency distributions : sim is a simple distribution which is equal to $\frac{D1}{1^t D1}$, pro is equal to $\frac{z}{1^t z1}$, where z is the nonnegative eigenvector of the matrix containing the squared dissimilarities among the elements, met is equal to z^2 , num is a frequency vector maximizing Rao's diversity coefficient.

Author(s)

Stéphane Champely <Stephane.Champely@univ-lyon1.fr>
 Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

- Rao, C.R. (1982) Diversity and dissimilarity coefficients: a unified approach. *Theoretical Population Biology*, **21**, 24–43.
- Gini, C. (1912) Variabilit'a e mutabilit'a. *Universite di Cagliari III*, Parte II.
- Simpson, E.H. (1949) Measurement of diversity. *Nature*, **163**, 688.
- Champely, S. and Chessel, D. (2002) Measuring biological diversity using Euclidean metrics. *Environmental and Ecological Statistics*, **9**, 167–177.
- Pavoine, S., Ollier, S. and Pontier, D. (2005) Measuring diversity from dissimilarities with Rao's quadratic entropy: are any dissimilarities suitable? *Theoretical Population Biology*, **67**, 231–239.

Examples

```
par.safe <- par()$mar
data(elec88)
par(mar = c(0.1, 0.1, 0.1, 0.1))
# Departments of France.
area.plot(elec88$area)

# Dissimilarity matrix.
d0 <- dist(elec88$xy)

# Frequency distribution maximizing spatial diversity in France
# according to Rao's quadratic entropy.
France.m <- divcmax(d0)
w0 <- France.m$vectors$num
v0 <- France.m$value
(1:94) [w0 > 0]

# Smallest circle including all the 94 departments.
# The squared radius of that circle is the maximal value of the
# spatial diversity.
w1 = elec88$xy[c(6, 28, 66), ]
w.c = apply(w1 * w0[c(6, 28, 66)], 2, sum)
```

```

symbols(w.c[1], w.c[2], circles = sqrt(v0), inc = FALSE, add = TRUE)
s.value(elec88$xy, w0, add.plot = TRUE)
par(mar = par.safe)

## Not run:
# Maximisation of Rao's diversity coefficient
# with ultrametric dissimilarities.
data(microsatt)
mic.genet <- count2genet(microsatt$tab)
mic.dist <- dist.genet(mic.genet, 1)
mic.phylog <- hclust2phylog(hclust(mic.dist))
plot.phylog(mic.phylog)
mic.maxpond <- divcmax(mic.phylog$Wdist)$vectors$num
dotchart.phylog(mic.phylog, mic.maxpond)

## End(Not run)

```

dotchart.phylog	<i>Representation of many quantitative variables in front of a phylogenetic tree</i>
-----------------	--

Description

dotchart.phylog represents the phylogenetic tree and draws Cleveland dot plot of each variable.

Usage

```

dotchart.phylog(phylog, values, y = NULL, scaling = TRUE, ranging =
TRUE, yranging = NULL, joining = TRUE, yjoining = NULL, ceti = 1, cdot =
1, csub = 1, f.phylog = 1/(1 + ncol(values)), ...)

```

Arguments

phylog	an object of class phylog
values	a vector or a data frame giving the variables
y	a vector which values correspond to leaves positions
scaling	if TRUE, data are scaled
ranging	if TRUE, dotplots are drawn with the same horizontal limits
yranging	a vector with two values giving the horizontal limits. If NULL, horizontal limits are defined by lower and upper values of data
joining	if TRUE, segments join each point to a central value
yjoining	a vector with the central value. If NULL, the central value equals 0
ceti	a character size for editing horizontal limits, used with par("cex")*ceti
cdot	a character size for plotting the points of the dot plot, used with par("cex")*cdot

csub	a character size for editing the names of variables, used with <code>par("cex")*csub</code>
f.phylog	a size coefficient for tree size (a parameter to draw the tree in proportion to leaves labels)
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel
Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

See Also

[symbols.phylog](#) and [table.phylog](#)

Examples

```
# one variable
tre <- c("((A,B),(C,D));")
phy <- newick2phylog(tre)
x <- 1:4
par(mfrow = c(2,2))
dotchart.phylog(phy, x, scaling = FALSE)
dotchart.phylog(phy, x)
dotchart.phylog(phy, x, joining = FALSE)
dotchart.phylog(phy, x, scaling = FALSE,
  yjoining = 0, y-ranging = c(-1, 5))
par(mfrow = c(1,1))

# many variables
data(mjrochet)
phy <- newick2phylog(mjrochet$tre)
tab <- data.frame(log(mjrochet$tab))
dotchart.phylog(phy, tab, ceti = 0.5, csub = 0.6,
  cleaves = 0, c-dot = 0.6)
par(mfrow=c(1,1))
```

dotcircle

Representation of n values on a circle

Description

This function represents n values on a circle. The n points are shared out regularly over the circle and put on the radius according to the value attributed to that measure.

Usage

```
dotcircle(z, alpha0 = pi/2, xlim = range(pretty(z)),
  labels = names(z), clabel = 1, cleg = 1)
```

Arguments

`z` : a numeric vector
`alpha0` : polar angle to put the first value
`xlim` : the ranges to be encompassed by the circle radius
`labels` : a vector of strings of characters for the angle labels
`clabel` : a character size for the labels, used with `par("cex")*clabel`
`cleg` : a character size for the ranges, used with `par("cex")*cleg`

Author(s)

Daniel Chessel

See Also

[circ.plot](#)

Examples

```
w <- scores.neig(neig(n.cir = 24))
par(mfrow = c(4,4))
for (k in 1:16) dotcircle(w[,k],labels = 1:24)
par(mfrow = c(1,1))
```

doubts

Pair of Ecological Tables

Description

This data set gives environmental variables, fish species and spatial coordinates for 30 sites.

Usage

```
data(doubts)
```

Format

`doubts` is a list with 4 components.

env is a data frame with 30 rows (sites) and 11 environmental variables.

fish is a data frame with 30 rows (sites) and 27 fish species.

xy is a data frame with 30 rows (sites) and 2 spatial coordinates.

species is a data frame with 27 rows (species) and 4 columns (names).

Details

The rows of `doubs$env`, `doubs$fish` and `doubs$xy` are 30 sites along the Doubs, a French and Switzerland river.

`doubs$env` contains the following variables: `dfs` - distance from the source (km * 10), `alt` - altitude (m), `slo` ($\ln(x + 1)$ where x is the slope (per mil * 100)), `flo` - minimum average stream flow (m³/s * 100), `pH` (* 10), `har` - total hardness of water (mg/l of Calcium), `pho` - phosphates (mg/l * 100), `nit` - nitrates (mg/l * 100), `amm` - ammonia nitrogen (mg/l * 100), `oxy` - dissolved oxygen (mg/l * 10), `bdo` - biological demand for oxygen (mg/l * 10).

`doubs$fish` contains the abundance of the following fish species: *Cottus gobio* (Cogo), *Salmo trutta fario* (Satr), *Phoxinus phoxinus* (Phph), *Nemacheilus barbatulus* (Neba), *Thymallus thymallus* (Thth), *Telestes soufia agassizi* (Teso), *Chondrostoma nasus* (Chna), *Chondrostoma toxostoma* (Chto), *Leuciscus leuciscus* (Lele), *Leuciscus cephalus cephalus* (Lece), *Barbus barbus* (Baba), *Spirulinus bipunctatus* (Spbi), *Gobio gobio* (Gogo), *Esox lucius* (Eslu), *Perca fluviatilis* (Pefl), *Rhodeus amarus* (Rham), *Lepomis gibbosus* (Legi), *Scardinius erythrophthalmus* (Scer), *Cyprinus carpio* (Cyca), *Tinca tinca* (Titi), *Abramis brama* (Abbr), *Ictalurus melas* (Icme), *Acerina cernua* (Acce), *Rutilus rutilus* (Ruru), *Blicca bjoerkna* (Blbj), *Alburnus alburnus* (Alal), *Anguilla anguilla* (Anan).

`doubs$species` contains the names of the 27 fish species. The four columns correspond to: 1 = scientific name (Genus species), 2 = French common name, 3 = English common name, 4 = Four character code.

Source

Verneaux, J. (1973) *Cours d'eau de Franche-Comté (Massif du Jura). Recherches écologiques sur le réseau hydrographique du Doubs. Essai de biotypologie*. Thèse d'état, Besançon. 1–257.

References

See a French description of fish species at <http://pbil.univ-lyon1.fr/R/articles/arti049.pdf>.

Chesse, D., Lebreton, J.D. and Yoccoz, N.G. (1987) Propriétés de l'analyse canonique des correspondances. Une illustration en hydrobiologie. *Revue de Statistique Appliquée*, **35**, 4, 55–72.

Examples

```
data(doubs)
pca1 <- dudi.pca(doubs$env, scan = FALSE)
pca2 <- dudi.pca(doubs$fish, scale = FALSE, scan = FALSE)
coiner1 <- coinertia(pca1, pca2, scan = FALSE)
par(mfrow = c(3,3))
s.corcircle(coiner1$aX)
s.value(doubs$xy, coiner1$lX[,1])
s.value(doubs$xy, coiner1$lX[,2])
s.arrow(coiner1$c1)
s.match(coiner1$mX, coiner1$mY)
s.corcircle(coiner1$aY)
s.arrow(coiner1$l1)
s.value(doubs$xy, coiner1$lY[,1])
s.value(doubs$xy, coiner1$lY[,2])
```

```
par(mfrow = c(1,1))
```

dpcoa

Double principal coordinate analysis

Description

Performs a double principal coordinate analysis

Usage

```
dpcoa (df, dis = NULL, scannf = TRUE, nf = 2, full = FALSE, tol = 1e-07)
## S3 method for class 'dpcoa'
plot(x, xax = 1, yax = 2, option = 1:4, csize = 2, ...)
## S3 method for class 'dpcoa'
print(x, ...)
```

Arguments

df	a data frame with elements as rows, samples as columns and abundance or presence-absence as entries
dis	an object of class <code>dist</code> containing the distances between the elements.
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if <code>scannf</code> is <code>FALSE</code> , an integer indicating the number of kept axes
full	a logical value indicating whether all non null eigenvalues should be kept
tol	a tolerance threshold for null eigenvalues (a value less than <code>tol</code> times the first one is considered as null)
x	an object of class <code>dpcoa</code>
xax	the column number for the x-axis
yax	the column number for the y-axis
option	the function <code>plot.dpcoa</code> produces four graphs, <code>option</code> allows us to choose only some of them
csize	a size coefficient for symbols
...	... further arguments passed to or from other methods

Value

Returns a list of class `dpcoa` containing:

call	call
nf	a numeric value indicating the number of kept axes
w1	a numeric vector containing the weights of the elements
w2	a numeric vector containing the weights of the samples

eig	a numeric vector with all the eigenvalues
RaoDiv	a numeric vector containing diversities within samples
RaoDis	an object of class <code>dist</code> containing the dissimilarities between samples
RaoDecodiv	a data frame with the decomposition of the diversity
l1	a data frame with the coordinates of the elements
l2	a data frame with the coordinates of the samples
c1	a data frame with the scores of the principal axes of the elements

Author(s)

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 Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Pavoine, S., Dufour, A.B. and Chessel, D. (2004) From dissimilarities among species to dissimilarities among communities: a double principal coordinate analysis. *Journal of Theoretical Biology*, **228**, 523–537.

Examples

```
data(humDNAm)
dpcoahum <- dpcoa(humDNAm$samples, sqrt(humDNAm$distances), scan = FALSE, nf = 2)
dpcoahum
plot(dpcoahum, csize = 1.5)
## Not run:
data(ecomor)
dtaxo <- dist.taxo(ecomor$taxo)
dpcoaeco <- dpcoa(ecomor$habitat, dtaxo, scan = FALSE, nf = 2)
dpcoaeco
plot(dpcoaeco, csize = 1.5)
## End(Not run)
```

dudi

Duality Diagram

Description

`as.dudi` is called by many functions (`dudi.pca`, `dudi.coa`, `dudi.acm`, ...) and not directly by the user. It creates duality diagrams.

`t.dudi` returns an object of class 'dudi' where the rows are the columns and the columns are the rows of the initial `dudi`.

`is.dudi` returns TRUE if the object is of class `dudi`

`redo.dudi` computes again an analysis, eventually changing the number of kept axes. Used by other functions.

Usage

```

as.dudi(df, col.w, row.w, scannf, nf, call, type, tol = 1e-07,
        full = FALSE)
## S3 method for class 'dudi'
print(x, ...)
is.dudi(x)
redo.dudi(dudi, newnf = 2)
## S3 method for class 'dudi'
t(x)
## S3 method for class 'dudi'
summary(object, ...)

```

Arguments

df	a data frame with n rows and p columns
col.w	a numeric vector containing the row weights
row.w	a numeric vector containing the column weights
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
call	generally <code>match.call()</code>
type	a string of characters : the returned list will be of class <code>c(type, "dudi")</code>
tol	a tolerance threshold for null eigenvalues (a value less than <code>tol</code> times the first one is considered as null)
full	a logical value indicating whether all non null eigenvalues should be kept
x, dudi, object	objects of class <code>dudi</code>
...	further arguments passed to or from other methods
newnf	an integer indicating the number of kept axes

Value

`as.dudi` and all the functions that use it return a list with the following components :

tab	a data frame with n rows and p columns
cw	column weights, a vector with n components
lw	row (lines) weights, a vector with p components
eig	eigenvalues, a vector with $\min(n,p)$ components
nf	integer, number of kept axes
c1	principal axes, data frame with p rows and nf columns
l1	principal components, data frame with n rows and nf columns
co	column coordinates, data frame with p rows and nf columns
li	row coordinates, data frame with n rows and nf columns
call	original call

Author(s)

Daniel Chessel
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 Stephane Dray <stephane.dray@univ-lyon1.fr>

References

Escoufier, Y. (1987) The duality diagram : a means of better practical applications In *Development in numerical ecology*, Legendre, P. & Legendre, L. (Eds.) NATO advanced Institute, Serie G. Springer Verlag, Berlin, 139–156.

Examples

```
data(deug)
dd1 <- dudi.pca(deug$tab, scannf = FALSE)
dd1
t(dd1)
is.dudi(dd1)
redo.dudi(dd1,3)
summary(dd1)
```

dudi.acm

Multiple Correspondence Analysis

Description

dudi.acm performs the multiple correspondence analysis of a factor table.
 acm.burt an utility giving the crossed Burt table of two factors table.
 acm.disjonctif an utility giving the complete disjunctive table of a factor table.
 boxplot.acm a graphic utility to interpret axes.

Usage

```
dudi.acm (df, row.w = rep(1, nrow(df)), scannf = TRUE, nf = 2)
acm.burt (df1, df2, counts = rep(1, nrow(df1)))
acm.disjonctif (df)
## S3 method for class 'acm'
boxplot(x, xax = 1, ...)
```

Arguments

df, df1, df2 data frames containing only factors
 row.w, counts vector of row weights, by default, uniform weighting
 scannf a logical value indicating whether the eigenvalues bar plot should be displayed
 nf if scannf FALSE, an integer indicating the number of kept axes

x an object of class acm
 xax the number of factor to display
 ... further arguments passed to or from other methods

Value

dudi.acm returns a list of class acm and dudi (see [dudi](#)) containing

cr a data frame which rows are the variables, columns are the kept scores and the values are the correlation ratios

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Tenenhaus, M. & Young, F.W. (1985) An analysis and synthesis of multiple correspondence analysis, optimal scaling, dual scaling, homogeneity analysis and other methods for quantifying categorical multivariate data. *Psychometrika*, **50**, 1, 91-119.

Lebart, L., A. Morineau, and M. Piron. 1995. Statistique exploratoire multidimensionnelle. Dunod, Paris.

See Also

[s.chull](#), [s.class](#)

Examples

```
data(ours)
summary(ours)
boxplot(dudi.acm(ours, scan = FALSE))
## Not run:
data(banque)
banque.acm <- dudi.acm(banque, scann = FALSE, nf = 3)
scatter.dudi(banque.acm)

apply(banque.acm$cr, 2, mean)
banque.acm$eig[1:banque.acm$nf] # the same thing
boxplot.acm(banque.acm)

scatter(banque.acm)

s.value(banque.acm$li, banque.acm$li[,3])

bb <- acm.burt(banque, banque)
bbcoa <- dudi.coa(bb, scann = FALSE)
plot(banque.acm$c1[,1], bbcoa$c1[,1])
# mca and coa of Burt table. Lebart & coll. section 1.4
```

```
bd <- acm.disjonctif(banque)
bdcoa <- dudi.coa(bd, scann = FALSE)
plot(banque.acm$li[,1], bdcoa$li[,1])
# mca and coa of disjonctive table. Lebart & coll. section 1.4
plot(banque.acm$co[,1], dudi.coa(bd, scann = FALSE)$co[,1])

## End(Not run)
```

dudi.coa

Correspondence Analysis

Description

performs a correspondence analysis.

Usage

```
dudi.coa(df, scannf = TRUE, nf = 2)
```

Arguments

df a data frame containing positive or null values
scannf a logical value indicating whether the eigenvalues bar plot should be displayed
nf if scannf FALSE, an integer indicating the number of kept axes

Value

returns a list of class coa and dudi (see [dudi](#)) containing

N the sum of all the values of the initial table

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Benzécri, J.P. and Coll. (1973) *L'analyse des données. II L'analyse des correspondances*, Bordas, Paris. 1–620.

Greenacre, M. J. (1984) *Theory and applications of correspondence analysis*, Academic Press, London.

Examples

```

data(rpjdl)
chisq.test(rpjdl$fau)$statistic
rpjdl.coa <- dudi.coa(rpjdl$fau, scannf = FALSE, nf = 4)
sum(rpjdl.coa$eig)*rpjdl.coa$N # the same

par(mfrow = c(1,2))
s.label(rpjdl.coa$co, clab = 0.6, lab = rpjdl$frlab)
s.label(rpjdl.coa$li, clab = 0.6)
par(mfrow = c(1,1))

data(bordeaux)
db <- dudi.coa(bordeaux, scan = FALSE)
db
score(db)

```

dudi.dec

Decentred Correspondence Analysis

Description

performs a decentred correspondence analysis.

Usage

```
dudi.dec(df, eff, scannf = TRUE, nf = 2)
```

Arguments

df	a data frame containing positive or null values
eff	a vector containing the reference distribution. Its length is equal to the number of rows of df
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Value

Returns a list of class dec and dudi (see [dudi](#)) containing also

R	sum of all the values of the initial table
---	--

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Dolédec, S., Chessel, D. and Olivier J. M. (1995) L'analyse des correspondances décentrée: application aux peuplements ichtyologiques du haut-Rhône. *Bulletin Français de la Pêche et de la Pisciculture*, **336**, 29–40.

Examples

```
data(ichtyo)
dudi1 <- dudi.dec(ichtyo$tab, ichtyo$eff, scan = FALSE)
sum(apply(ichtyo$tab, 2, function(x)
  chisq.test(x, p = ichtyo$eff/sum(ichtyo$eff))$statistic))
sum(dudi1$eig) * sum(ichtyo$eff) # the same

s.class(dudi1$li, ichtyo$dat, wt = ichtyo$eff/sum(ichtyo$eff))
```

dudi.fca	<i>Fuzzy Correspondence Analysis and Fuzzy Principal Components Analysis</i>
----------	--

Description

These functions analyse a table of fuzzy variables.

A fuzzy variable takes values of type $a = (a_1, \dots, a_k)$ giving the importance of k categories.

A missing data is denoted $(0, \dots, 0)$.

Only the profile $a/\text{sum}(a)$ is used, and missing data are replaced by the mean profile of the others in the function `prep.fuzzy.var`. See ref. for details.

Usage

```
prep.fuzzy.var (df, col.blocks, row.w = rep(1, nrow(df)))
dudi.fca(df, scannf = TRUE, nf = 2)
dudi.fpca(df, scannf = TRUE, nf = 2)
```

Arguments

df	a data frame containing positive or null values
col.blocks	a vector containing the number of categories for each fuzzy variable
row.w	a vector of row weights
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if <code>scannf</code> FALSE, an integer indicating the number of kept axes

Value

The function `prep.fuzzy.var` returns a data frame with the attribute `col.blocks`. The function `dudi.fca` returns a list of class `fca` and `dudi` (see [dudi](#)) containing also

`cr` a data frame which rows are the blocs, columns are the kept axes, and values are the correlation ratios.

The function `dudi.fpca` returns a list of class `pca` and `dudi` (see [dudi](#)) containing also

1. cent
2. norm
3. blo
4. indica
5. FST
6. inertia

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Chevenet, F., Dolédec, S. and Chessel, D. (1994) A fuzzy coding approach for the analysis of long-term ecological data. *Freshwater Biology*, **31**, 295–309.

Examples

```
w1 <- matrix(c(1,0,0,2,1,1,0,2,2,0,1,0,1,1,1,0,1,3,1,0), 4, 5)
w1 <- data.frame(w1)
w2 <- prep.fuzzy.var(w1, c(2,3))
w1
w2
attributes(w2)

data(bsetal97)
w <- prep.fuzzy.var(bsetal97$biol, bsetal97$biol.blo)
scatter(dudi.fca(w, scann = FALSE, nf = 3), csub = 3, clab.moda = 1.5)
scatter(dudi.fpca(w, scann = FALSE, nf = 3), csub = 3, clab.moda = 1.5)

## Not run:
w1 <- prep.fuzzy.var(bsetal97$biol, bsetal97$biol.blo)
w2 <- prep.fuzzy.var(bsetal97$ecol, bsetal97$ecol.blo)
d1 <- dudi.fca(w1, scann = FALSE, nf = 3)
d2 <- dudi.fca(w2, scann = FALSE, nf = 3)
plot(coinertia(d1, d2, scann = FALSE))

## End(Not run)
```

dudi.hillsmith *Ordination of Tables mixing quantitative variables and factors*

Description

performs a multivariate analysis with mixed quantitative variables and factors.

Usage

```
dudi.hillsmith(df, row.w = rep(1, nrow(df))/nrow(df),  
              scannf = TRUE, nf = 2)
```

Arguments

df	a data frame with mixed type variables (quantitative and factor)
row.w	a vector of row weights, by default uniform row weights are used
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Details

If df contains only quantitative variables, this is equivalent to a normed PCA.

If df contains only factors, this is equivalent to a MCA.

This analysis is the Hill and Smith method and is very similar to dudi.mix function. The differences are that dudi.hillsmith allow to use various row weights, while dudi.mix deals with ordered variables.

The principal components of this analysis are centered and normed vectors maximizing the sum of
:

squared correlation coefficients with quantitative variables
correlation ratios with factors

Value

Returns a list of class mix and dudi (see [dudi](#)) containing also

index	a factor giving the type of each variable : f = factor, q = quantitative
assign	a factor indicating the initial variable for each column of the transformed table
cr	a data frame giving for each variable and each score: the squared correlation coefficients if it is a quantitative variable the correlation ratios if it is a factor

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Hill, M. O., and A. J. E. Smith. 1976. Principal component analysis of taxonomic data with multi-state discrete characters. *Taxon*, **25**, 249-255.

See Also

dudi.mix

Examples

```
data(dunedata)
attributes(dunedata$envir$use)$class <- "factor" # use dudi.mix for ordered data
dd1 <- dudi.hillsmith(dunedata$envir, scannf = FALSE)
scatter.dudi(dd1, clab.r = 1, clab.c = 1.5)
```

dudi.mix

Ordination of Tables mixing quantitative variables and factors

Description

performs a multivariate analysis with mixed quantitative variables and factors.

Usage

```
dudi.mix(df, add.square = FALSE, scannf = TRUE, nf = 2)
```

Arguments

df	a data frame with mixed type variables (quantitative, factor and ordered)
add.square	a logical value indicating whether the squares of quantitative variables should be added
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Details

If df contains only quantitative variables, this is equivalent to a normed PCA.

If df contains only factors, this is equivalent to a MCA.

Ordered factors are replaced by `poly(x, deg=2)`.

This analysis generalizes the Hill and Smith method.

The principal components of this analysis are centered and normed vectors maximizing the sum of the:

squared correlation coefficients with quantitative variables

squared multiple correlation coefficients with polynoms

correlation ratios with factors.

Value

Returns a list of class `mix` and `dudi` (see [dudi](#)) containing also

<code>index</code>	a factor giving the type of each variable : f = factor, o = ordered, q = quantitative
<code>assign</code>	a factor indicating the initial variable for each column of the transformed table
<code>cr</code>	a data frame giving for each variable and each score: the squared correlation coefficients if it is a quantitative variable the correlation ratios if it is a factor the squared multiple correlation coefficients if it is ordered

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Hill, M. O., and A. J. E. Smith. 1976. Principal component analysis of taxonomic data with multi-state discrete characters. *Taxon*, **25**, 249-255.

De Leeuw, J., J. van Rijkevorsel, and . 1980. HOMALS and PRINCALS - Some generalizations of principal components analysis. Pages 231-242 in E. Diday and Coll., editors. *Data Analysis and Informatics II*. Elsevier Science Publisher, North Holland, Amsterdam.

Kiers, H. A. L. 1994. Simple structure in component analysis techniques for mixtures of qualitative and quantitative variables. *Psychometrika*, **56**, 197-212.

Examples

```
data(dunedata)
dd1 <- dudi.mix(dunedata$envir, scann = FALSE)
scatter.dudi(dd1, clab.r = 1, clab.c = 1.5)

dd2 <- dudi.mix(dunedata$envir, scann = FALSE, add = TRUE)
scatter.dudi(dd2, clab.r = 1, clab.c = 1.5)
```

dudi.nsc

Non symmetric correspondence analysis

Description

performs a non symmetric correspondence analysis.

Usage

```
dudi.nsc(df, scannf = TRUE, nf = 2)
```

Arguments

df a data frame containing positive or null values
scannf a logical value indicating whether the eigenvalues bar plot should be displayed
nf if scannf FALSE, an integer indicating the number of kept axes

Value

Returns a list of class nsc and dudi (see [dudi](#)) containing also

N sum of the values of the initial table

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Kroonenberg, P. M., and Lombardo R. (1999) Nonsymmetric correspondence analysis: a tool for analysing contingency tables with a dependence structure. *Multivariate Behavioral Research*, **34**, 367–396.

Examples

```
data(housetasks)
nsc1 <- dudi.nsc(housetasks, scan = FALSE)
s.label(nsc1$c1, clab = 1.25)
s.arrow(nsc1$li, add.pl = TRUE, clab = 0.75) # see ref p.383
```

dudi.pca *Principal Component Analysis*

Description

dudi.pca performs a principal component analysis of a data frame and returns the results as objects of class pca and dudi.

Usage

```
dudi.pca(df, row.w = rep(1, nrow(df))/nrow(df),
         col.w = rep(1, ncol(df)), center = TRUE, scale = TRUE,
         scannf = TRUE, nf = 2)
```

Arguments

df	a data frame with n rows (individuals) and p columns (numeric variables)
row.w	an optional row weights (by default, uniform row weights)
col.w	an optional column weights (by default, unit column weights)
center	a logical or numeric value, centring option if TRUE, centring by the mean if FALSE no centring if a numeric vector, its length must be equal to the number of columns of the data frame df and gives the decentring
scale	a logical value indicating whether the column vectors should be normed for the row.w weighting
scannf	a logical value indicating whether the screeplot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Value

Returns a list of classes `pca` and `dudi` (see [dudi](#)) containing the used information for computing the principal component analysis :

tab	the data frame to be analyzed depending of the transformation arguments (center and scale)
cw	the column weights
lw	the row weights
eig	the eigenvalues
rank	the rank of the analyzed matrice
nf	the number of kept factors
c1	the column normed scores i.e. the principal axes
l1	the row normed scores
co	the column coordinates
li	the row coordinates i.e. the principal components
call	the call function
cent	the p vector containing the means for variables (Note that if center = F, the vector contains $p 0$)
norm	the p vector containing the standard deviations for variables i.e. the root of the sum of squares deviations of the values from their means divided by n (Note that if norm = F, the vector contains $p 1$)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

See Also

prcomp, princomp in the mva library

Examples

```

data(deug)
deug.dudi <- dudi.pca(deug$stab, center = deug$cent,
  scale = FALSE, scan = FALSE)
par(mfrow = c(2,2))
s.class(deug.dudi$li, deug$result, cpoint = 1)
s.arrow(deug.dudi$c1, lab = names(deug$stab))
deug.dudi1 <- dudi.pca(deug$stab, center = TRUE,
  scale = TRUE, scan = FALSE)
s.class(deug.dudi1$li, deug$result, cpoint = 1)
s.corcircle(deug.dudi1$co, lab = names(deug$stab),
  full = FALSE, box = TRUE)
par(mfrow = c(1,1))

# for interpretations
par(mfrow = c(3,3))
par(mar = c(2.1,2.1,2.1,1.1))
for(i in 1:9) {
hist(deug.dudi$stab[,i], xlim = c(-40,40), breaks = seq(-45, 35, by = 5),
  prob = TRUE, right = FALSE, main = names(deug$stab)[i], xlab = "",
  ylim = c(0,0.10))
abline(v = 0, lwd = 3)
}
par(mfrow = c(1,1))

```

dudi.pco

Principal Coordinates Analysis

Description

dudi.pco performs a principal coordinates analysis of a Euclidean distance matrix and returns the results as objects of class pco and dudi.

Usage

```

dudi.pco(d, row.w = "uniform", scannf = TRUE, nf = 2,
  full = FALSE, tol = 1e-07)
## S3 method for class 'pco'
scatter(x, xax = 1, yax = 2, clab.row = 1, posieig = "top",
  sub = NULL, csub = 2, ...)

```


Arguments

d	an object of class <code>dist</code> containing a Euclidean distance matrix.
row.w	an optional distance matrix row weights. If not <code>NULL</code> , must be a vector of positive numbers with length equal to the size of the distance matrix
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if <code>scannf</code> <code>FALSE</code> , an integer indicating the number of kept axes
full	a logical value indicating whether all the axes should be kept
tol	a tolerance threshold to test whether the distance matrix is Euclidean : an eigenvalue is considered positive if it is larger than $-tol * \lambda_1$ where λ_1 is the largest eigenvalue.
x	an object of class <code>pco</code>
xax	the column number for the x-axis
yax	the column number for the y-axis
clab.row	a character size for the row labels
posieig	if "top" the eigenvalues bar plot is upside, if "bottom" it is downside, if "none" no plot
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex") * csub</code>
...	further arguments passed to or from other methods

Value

`dudi.pco` returns a list of class `pco` and `dudi`. See [dudi](#)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Gower, J. C. (1966) Some distance properties of latent root and vector methods used in multivariate analysis. *Biometrika*, **53**, 325–338.

Examples

```
data(yanomama)
gen <- quasieuclid(as.dist(yanomama$gen))
geo <- quasieuclid(as.dist(yanomama$geo))
ant <- quasieuclid(as.dist(yanomama$ant))
geo1 <- dudi.pco(geo, scann = FALSE, nf = 3)
gen1 <- dudi.pco(gen, scann = FALSE, nf = 3)
ant1 <- dudi.pco(ant, scann = FALSE, nf = 3)
plot(coinertia(ant1, gen1, scann = FALSE))
```

dunedata

Dune Meadow Data

Description

dunedata is a data set containing for 20 sites, environmental variables and plant species.

Usage

```
data(dunedata)
```

Format

dunedata is a list with 2 components.

envir is a data frame with 20 rows (sites) 5 columns (environmental variables).

veg is a data frame with 20 rows (sites) 30 columns (plant species).

Source

Jongman, R. H., ter Braak, C. J. F. and van Tongeren, O. F. R. (1987) *Data analysis in community and landscape ecology*, Pudoc, Wageningen.

Examples

```
data(dunedata)
summary(dunedata$envir)
is.ordered(dunedata$envir$use)
score(dudi.mix(dunedata$envir, scan = FALSE))
```

ecg

Electrocardiogram data

Description

These data were measured during the normal sinus rhythm of a patient who occasionally experiences arrhythmia. There are 2048 observations measured in units of millivolts and collected at a rate of 180 samples per second. This time series is a good candidate for a multiresolution analysis because its components are on different scales. For example, the large scale (low frequency) fluctuations, known as baseline drift, are due to the patient respiration, while the prominent short scale (high frequency) intermittent fluctuations between 3 and 4 seconds are evidently due to patient movement. Heart rhythm determines most of the remaining features in the series. The large spikes occurring about 0.7 seconds apart the R waves of normal heart rhythm; the smaller, but sharp peak coming just prior to an R wave is known as a P wave; and the broader peak that comes after a R wave is a T wave.

Usage

```
data(ecg)
```

Format

A vector of class `ts` containing 2048 observations.

Source

Gust Bardy and Per Reinhall, University of Washington

References

Percival, D. B., and Walden, A.T. (2000) *Wavelet Methods for Time Series Analysis*, Cambridge University Press.

Examples

```
## Not run:
# figure 130 in Percival and Walden (2000)
if (require(waveslim) == TRUE) {
  data(ecg)
  ecg.level <- haar2level(ecg)
  ecg.haar <- orthobasis.haar(length(ecg))
  ecg.mld <- mld(ecg, ecg.haar, ecg.level, plot = FALSE)
  res <- cbind.data.frame(apply(ecg.mld[,1:5],1,sum), ecg.mld[,6:11])
  par(mfrow = c(8,1))
  par(mar = c(2, 5, 1.5, 0.6))
  plot(as.ts(ecg), ylab = "ECG")
  apply(res, 2, function(x) plot(as.ts(x), ylim = range(res),
    ylab = ""))
  par(mfrow = c(1,1))
}
## End(Not run)
```

Description

This data set gives ecomorphological informations about 129 bird species.

Usage

```
data(ecomor)
```

Format

ecomor is a list of 7 components.

forsub is a data frame with 129 species, 6 variables (the feeding place classes): foliage, ground, twig, bush, trunk and aerial feeders. These dummy variables indicate the use (1) or no use (0) of a given feeding place by a species.

diet is a data frame with 129 species and 8 variables (diet types): Gr (granivorous: seeds), Fr (frugivorous: berries, acorns, drupes), Ne (frugivorous: nectar), Fo (folivorous: leaves), In (invertebrate feeder: insects, spiders, myriapods, isopods, snails, worms), Ca (carnivorous: flesh of small vertebrates), Li (limnivorous: invertebrates in fresh water), and Ch (carrion feeder). These dummy variables indicate the use (1) or no use (0) of a given diet type by a species.

habitat is a data frame with 129 species, 16 dummy variables (the habitats). These variables indicate the species presence (1) or the species absence (0) in a given habitat.

morpho is a data frame with 129 species and 8 morphological variables: wingl (Wing length, mm), taill (Tail length, mm), culml (Culmen length, mm), bilh (Bill height, mm), bilw (Bill width, mm), tarsl (Tarsus length, mm), midtl (Middle toe length, mm) and weig (Weight, g).

taxo is a data frame with 129 species and 3 factors: Genus, Family and Order. It is a data frame of class 'taxo': the variables are factors giving nested classifications.

labels is a data frame with vectors of the names of species (complete and in abbreviated form).

categ is a data frame with 129 species, 2 factors: 'forsub' summarizing the feeding place and 'diet' the diet type.

Source

Blondel, J., Vuilleumier, F., Marcus, L.F., and Terouanne, E. (1984). Is there ecomorphological convergence among mediterranean bird communities of Chile, California, and France. In *Evolutionary Biology* (eds M.K. Hecht, B. Wallace and R.J. MacIntyre), 141–213, **18**. Plenum Press, New York.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps023.pdf> (in French).

Examples

```
data(ecomor)
ric <- apply(ecomor$habitat, 2, sum)
s.corcircle(dudi.pca(log(ecomor$morpho), scan = FALSE)$co)

forsub <- data.frame(t(apply(ecomor$forsub, 1,
  function(x) x/sum(x))))
pca1 <- dudi.pca(forsub, scan = FALSE, scale = FALSE)
s.arrow(pca1$c1)
w <- as.matrix(forsub)
s.label(w, clab = 0, add.p = TRUE, cpoi = 2)

diet <- data.frame(t(apply(ecomor$diet, 1,
```

```

function (x) x/sum(x)))
pca2 <- dudi.pca(diet, scan = FALSE, scale = FALSE)
s.arrow(pca2$c1)
w <- as.matrix(diet)
s.label(w, clab = 0, add.p = TRUE, cpoi = 2)
## Not run:
dmorpho <- dist.quant(log(ecomor$morpho), 3)
dhabitat <- dist.binary(ecomor$habitat, 1)
dtaxo <- dist.taxo(ecomor$taxo)

mantel.randtest(dmorpho, dhabitat)
RV.rtest(pcoscaled(dmorpho), pcoscaled(dhabitat), 999)
procuste.randtest(pcoscaled(dmorpho), pcoscaled(dhabitat))

ecophy <- taxo2phylog(ecomor$taxo, add.tools=TRUE)
table.phylog(ecomor$habitat, ecophy, clabel.n = 0.5, f = 0.6,
             clabel.c = 0.75, clabel.r = 0.5, csi = 0.75, cleg = 0)
plot.phylog(ecophy, clabel.n = 0.75, clabel.l = 0.75,
            labels.l = ecomor$labels[,"latin"])
mantel.randtest(dmorpho, dtaxo)
mantel.randtest(dhabitat, dtaxo)

## End(Not run)

```

EH

Amount of Evolutionary History

Description

computes the sum of branch lengths on an ultrametric phylogenetic tree.

Usage

```
EH(phy1, select = NULL)
```

Arguments

phy1	an object of class phylog
select	a vector containing the numbers of the leaves (species) which must be considered in the computation of the amount of Evolutionary History. This parameter allows the calculation of the amount of Evolutionary History for a subset of species.

Value

returns a real value.

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Nee, S. and May, R.M. (1997) Extinction and the loss of evolutionary history. *Science*, **278**, 692–694.

Examples

```
data(carni70)
carni70.phy <- newick2phylog(carni70$tre)
EH(carni70.phy)
EH(carni70.phy, select = 1:15) # Felidae
```

elec88

Electoral Data

Description

This data set gives the results of the presidential election in France in 1988 for each department and all the candidates.

Usage

```
data(elec88)
```

Format

elec88 is a list of 7 components.

tab is a data frame with 94 rows (departments) and 9 variables (candidates)

res is the global result of the election all-over the country.

lab is a data frame with three variables: elec88\$lab\$dep a vector containing the names of the 94 french departments, elec88\$lab\$reg a vector containing the names of the 21 French administrative regions. and, elec88\$lab\$reg.fac a factor with 21 levels defining the French administrative regions.

area is the data frame of 3 variables returning the boundary lines of each department. The first variable is a factor. The levels of this one are the row.names of tab. The second and third variables return the coordinates (x,y) of the points of the boundary line.

contour is a data frame with 4 variables (x1,y1,x2,y2)for the contour display of France

xy is a data frame with two variables (x,y) giving the position of the center for each department

neig is the neighbouring graph between departments, object of the class neig

Source

Public data

See Also

This dataset is compatible with presid2002 and cnc2003

Examples

```

data(elec88)
apply(elec88$tab, 2, mean)
summary(elec88$res)

par(mfrow = c(2,2))
plot(elec88$area[,2:3], type = "n", asp = 1)
lpoly <- split(elec88$area[,2:3], elec88$area[,1])
lapply(lpoly, function(x) {points (x,type = "l");invisible()})
polygon(elec88$area[elec88$area$V1=="D25", 2:3], col = 1)
area.plot(elec88$area, graph = elec88$neig, lwdg = 1)
polygon(elec88$area[elec88$area$V1=="D25", 2:3], col = 1)
pca1 <- dudi.pca(elec88$tab, scal = FALSE, scan = FALSE)
area.plot(elec88$area, val = elec88$xy[,1] + elec88$xy[,2])
area.plot(elec88$area, val = pca1$li[,1], sub = "F1 PCA",
          csub = 2, cleg = 1.5)
par(mfrow = c(1,1))

```

escopage

K-tables of wine-tasting

Description

This data set describes 27 characteristics of 21 wines distributed in four fields : rest, visual, olfactory and global.

Usage

```
data(escopage)
```

Format

escopage is a list of 3 components.

tab is a data frame with 21 observations (wines) and 27 variables.

tab.names is the vector of the names of sub-tables : "rest" "visual" "olfactory" "global".

blo is a vector of the numbers of variables for each sub-table.

Source

Escofier, B. and Pagès, J. (1990) *Analyses factorielles simples et multiples : objectifs, méthodes et interprétation* Dunod, Paris. 1–267.

Escofier, B. and Pagès, J. (1994) Multiple factor analysis (AFMULT package). *Computational Statistics and Data Analysis*, **18**, 121–140.

Examples

```
data(escopage)
w <- data.frame(scale(escopage$tab))
w <- ktab.data.frame(w, escopage$blo)
names(w)[1:4] <- escopage$tab.names
plot(mfa(w, scan = FALSE))
```

euro123

*Triangular Data***Description**

This data set gives the proportions of employment in the primary, secondary and tertiary sectors for 12 European countries in 1978, 1986 and 1997.

Usage

```
data(euro123)
```

Format

euro123 is a list of 4 components.

in78 is a data frame with 12 rows and 3 variables.

in86 : idem in 1986

in97 : idem in 1997

plan is a data frame with two factors to both organize the 3 tables.

Source

Encyclopaedia Universalis, Symposium, Les chiffres du Monde. Encyclopaedia Universalis, Paris. 519.

Université de Barcelone : <http://www.ub.es/medame/nutstat1.html>

Examples

```
data(euro123)
par(mfrow = c(2,2))
triangle.plot(euro123$in78, addaxes = TRUE)
triangle.plot(euro123$in86, addaxes = TRUE)
triangle.plot(euro123$in97, addaxes = TRUE)
triangle.biplot(euro123$in78, euro123$in97)
par(mfrow = c(1,1))
```

fission

Fission pattern and heritable morphological traits

Description

This data set contains the mean values of five highly heritable linear combinations of cranial metric (GM1-GM3) and non metric (GN1-GN2) for 8 social groups of Rhesus Macaques on Cayo Santiago. It also describes the fission tree depicting the historical phyletic relationships.

Usage

```
data(fission)
```

Format

fission is a list containing the 2 following objects :

tre is a character string giving the fission tree in Newick format.

tab is a data frame with 8 social groups and five traits : cranial metrics (GM1, GM2, GM3) and cranial non metrics (GN1, GN2)

References

Cheverud, J. and Dow, M.M. (1985) An autocorrelation analysis of genetic variation due to lineal fission in social groups of rhesus macaques. *American Journal of Physical Anthropology*, **67**, 113–122.

Examples

```
data(fission)
fis.phy <- newick2phylog(fission$tre)
table.phylog(fission$tab[names(fis.phy$leaves),], fis.phy, csi = 2)
gearymoran(fis.phy$Amat, fission$tab)
```

foucart

K-tables Correspondence Analysis with the same rows and the same columns

Description

K tables have the same rows and the same columns.

Each table is transformed by $P = X/\text{sum}(X)$. The average of P is computing.

A correspondence analysis is realized on this average.

The initial rows and the initial columns are projected in supplementary elements.

Usage

```
foucart(X, scannf = TRUE, nf = 2)
## S3 method for class 'foucart'
plot(x, xax = 1, yax = 2, clab = 1, csub = 2,
     possub = "bottomright", ...)
## S3 method for class 'foucart'
print(x, ...)
```

Arguments

X	a list of data frame where the row names and the column names are the same for each table
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
x	an object of class 'foucart'
xax	the column number of the x-axis
yax	the column number of the y-axis
clab	if not NULL, a character size for the labels, used with <code>par("cex")*clab</code>
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
...	further arguments passed to or from other methods

Value

foucart returns a list of the classes 'dudi', 'coa' and 'foucart'

call	origine
nf	axes-components saved
rank	rank
blo	useful vector
cw	vector: column weights
lw	vector: row weights
eig	vector: eigen values
tab	data.frame: modified array
li	data.frame: row coordinates
l1	data.frame: row normed scores
co	data.frame: column coordinates
c1	data.frame: column normed scores
Tli	data.frame: row coordinates (each table)
Tco	data.frame: col coordinates (each table)
TL	data.frame: factors for Tli
TC	data.frame: factors for Tco

Author(s)

P. Bady <pierre.bady@univ-lyon1.fr>
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Foucart, T. (1984) *Analyse factorielle de tableaux multiples*, Masson, Paris.

Examples

```
data(bf88)
fou1 <- foucart(bf88, scann = FALSE, nf = 3)
fou1
plot(fou1)

data(meaudret)
l1 <- split(meaudret$spe, meaudret$design$season)
l1 <- lapply(l1, function(x)
  {row.names(x) <- paste("Sit", 1:5, sep="");x})
fou2 <- foucart(l1, scan = FALSE)
kplot(fou2, clab.r = 2)
```

fourthcorner

Functions to compute the fourth-corner statistic

Description

These functions allow to compute the fourth-corner statistic for abundance or presence-absence data. The fourth-corner statistic has been developed by Legendre et al (1997) and extended in Dray and Legendre (2008). The statistic measures the link between three tables: a table L (n x p) containing the abundances of p species at n sites, a second table R (n x m) with the measurements of m environmental variables for the n sites, and a third table Q (p x s) describing s species traits for the p species.

Usage

```
fourthcorner(tabR, tabL, tabQ, modeltype = 6, nrepet = 999, tr01 =
FALSE, p.adjust.method.G = p.adjust.methods, p.adjust.method.D =
p.adjust.methods, p.adjust.D = c("global", "levels"))
fourthcorner2(tabR, tabL, tabQ, modeltype = 6, nrepet = 999, tr01 =
FALSE, p.adjust.method.G = p.adjust.methods)
## S3 method for class '4thcorner'
print(x, varQ = 1:length(x$varnames.Q), varR =
1:length(x$varnames.R), stat = c("D", "D2"), ...)
## S3 method for class '4thcorner'
summary(object,...)
## S3 method for class '4thcorner'
plot(x, stat = c("D", "D2", "G"), type = c("table",
```

```
"biplot"), xax = 1, yax = 2, x.rlq = NULL, alpha = 0.05, col =
c("lightgrey", "red", "deepskyblue", "purple"), ...)
fourthcorner.rlq(xtest, nrepet = 999, modeltype = 6, typetest =
c("axes", "Q.axes", "R.axes"), p.adjust.method.G = p.adjust.methods,
p.adjust.method.D = p.adjust.methods, p.adjust.D = c("global",
"levels"), ...)
```

Arguments

tabR	a dataframe with the measurements of m environmental variables (columns) for the n sites (rows).
tabL	a dataframe containing the abundances of p species (columns) at n sites (rows).
tabQ	a dataframe describing s species traits (columns) for the p species (rows).
modeltype	an integer (1-6) indicating the permutation model used in the testing procedure (see details).
nrepet	the number of permutations
tr01	a logical indicating if data in tabL must be transformed to presence-absence data (FALSE by default)
object	an object of the class 4thcorner
x	an object of the class 4thcorner
varR	a vector with indices for variables in tabR
varQ	a vector with indices for variables in tabQ
type	results are represented by a table or on a biplot (see x.rlq)
alpha	a value of significance level
p.adjust.method.G	a string indicating a method for multiple adjustment used for output tabG, see p.adjust.methods for possible choices
p.adjust.method.D	a string indicating a method for multiple adjustment used for output tabD/tabD2, see p.adjust.methods for possible choices
p.adjust.D	a string indicating if multiple adjustment for tabD/tabD2 should be done globally or only between levels of a factor ("levels", as in the original paper of Legendre et al. 1997)
stat	a character to specify if results should be plotted for cells (D and D2) or variables (G)
xax	an integer indicating which rlq axis should be plotted on the x-axis
yax	an integer indicating which rlq axis should be plotted on the y-axis
x.rlq	an object created by the rlq function. Used to represent results on a biplot (type should be "biplot" and object created by the fourthcorner functions)
col	a vector of length 4 containing four colors used for the graphical representations. The first is used to represent non-significant associations, the second positive significant, the third negative significant. For the 'biplot' method and objects created by the fourthcorner.rlq function, the second corresponds to variables significantly linked to the x-axis, the third for the y-axis and the fourth for both axes

<code>xtest</code>	an object created by the <code>r1q</code> function
<code>typetest</code>	a string indicating which tests should be performed
<code>...</code>	further arguments passed to or from other methods

Details

For the `fourthcorner` function, the link is measured by a Pearson correlation coefficient for two quantitative variables (trait and environmental variable), by a Pearson Chi2 and G statistic for two qualitative variables and by a Pseudo-F and Pearson r for one quantitative variable and one qualitative variable. The `fourthcorner2` function offers a multivariate statistic (equal to the sum of eigenvalues of RLQ analysis) and measures the link between two variables by a square correlation coefficient (quant/quant), a Chi2/sum(L) (qual/qual) and a correlation ratio (quant/qual). The significance is tested by a permutation procedure. Different models are available:

- model 1 (`modeltype=1`): Permute values for each species independently (i.e., permute within each column of table L)
- model 2 (`modeltype=2`): Permute values of sites (i.e., permute entire rows of table L)
- model 3 (`modeltype=3`): Permute values for each site independently (i.e., permute within each row of table L)
- model 4 (`modeltype=4`): Permute values of species (i.e., permute entire columns of table L)
- model 5 (`modeltype=5`): Permute values of species and after (or before) permute values of sites (i.e., permute entire columns and after (or before) entire rows of table L)
- model 6 (`modeltype=6`): combination of the outputs of models 2 and 4. Dray and Legendre (2008) and ter Braak et al. (20012) showed that all models (except model 6) have inflated type I error.

Note that the model 5 is strictly equivalent to permuting simultaneously the rows of tables R and Q, as proposed by Doledec et al. (1996).

The function `summary` returns results for variables (G). The function `print` returns results for cells (D and D2). In the case of qualitative variables, Holm's corrected pvalues are also provided.

The function `plot` produces a graphical representation of the results (white for non significant, light grey for negative significant and dark grey for positive significant relationships). Results can be plotted for variables (G) or for cells (D and D2). In the case of qualitative / quantitative association, homogeneity (D) or correlation (D2) are plotted.

Value

The `fourthcorner` function returns a a list where:

`tabD` is a `krandtest` object giving the results of tests for cells of the fourth-corner (homogeneity for quant./qual.). `tabD2` is a `krandtest` object giving the results of tests for cells of the fourth-corner (Pearson r for quant./qual.). `tabG` is a `krandtest` object giving the results of tests for variables (Pearson's Chi2 for qual./qual.).

The `fourthcorner2` function returns a list where:

`tabG` is a `krandtest` object giving the results of tests for variables. `trRLQ` is a `krandtest` object giving the results of tests for the multivariate statistic (i.e. equivalent to `randtest.r1q` function).

Author(s)

Stephane Dray <stephane.drays@univ-lyon1.fr>

References

- Doledec, S., Chessel, D., ter Braak, C.J.F. and Champely, S. (1996) Matching species traits to environmental variables: a new three-table ordination method. *Environmental and Ecological Statistics*, **3**, 143–166.
- Legendre, P., R. Galzin, and M. L. Harmelin-Vivien. (1997) Relating behavior to habitat: solutions to the fourth-corner problem. *Ecology*, **78**, 547–562.
- Dray, S. and Legendre, P. (2008) Testing the species traits-environment relationships: the fourth-corner problem revisited. *Ecology*, **89**, 3400–3412.
- ter Braak, C., Cormont, A., and Dray, S. (2012) Improved testing of species traits-environment relationships in the fourth corner problem. *Ecology*, **93**, 1525–1526.
- Dray, S., Choler, P., Doledec, S., Peres-Neto, P.R., Thuiller, W., Pavoine, S. and ter Braak, C.J.F. (2013) Combining the fourth-corner and the RLQ methods for assessing trait responses to environmental variation. *Ecology*, in press.

See Also

[rlq, combine.4thcorner, p.adjust.methods](#)

Examples

```
data(aviurba)

## Version using the sequential test (ter Braak et al 2012)
## as recommended in Dray et al (2013),
## using Holm correction of P-values (only 99 permutations here)
four.comb.default <- fourthcorner(aviurba$mil,aviurba$fau,aviurba$traits,nrepet=99)
summary(four.comb.default)
plot(four.comb.default, stat = "G")

## using fdr correction of P-values
four.comb.fdr <- fourthcorner(aviurba$mil, aviurba$fau, aviurba$traits,
nrepet = 99, p.adjust.method.G = 'fdr', p.adjust.method.D = 'fdr')
summary(four.comb.fdr)
plot(four.comb.fdr, stat = "G")

## Explicit procedure to combine the results of two models
## proposed in Dray and Legendre (2008);the above does this implicitly
four2 <- fourthcorner(aviurba$mil,aviurba$fau,aviurba$traits,nrepet=99,modeltype=2)
four4 <- fourthcorner(aviurba$mil,aviurba$fau,aviurba$traits,nrepet=99,modeltype=4)
four.comb <- combine.4thcorner(four2, four4)
summary(four.comb)
plot(four.comb, stat = "G")
```

friday87	<i>Faunistic K-tables</i>
----------	---------------------------

Description

This data set gives informations about sites, species and environmental variables.

Usage

```
data(frday87)
```

Format

friday87 is a list of 4 components.

fau is a data frame containing a faunistic table with 16 sites and 91 species.

mil is a data frame with 16 sites and 11 environmental variables.

fau.blo is a vector of the number of species per group.

tab.names is the name of each group of species.

Source

Friday, L.E. (1987) The diversity of macroinvertebrate and macrophyte communities in ponds, *Freshwater Biology*, **18**, 87–104.

Examples

```
data(frday87)
wfri <- data.frame(scale(frday87$fau, scal = FALSE))
wfri <- ktab.data.frame(wfri, frday87$fau.blo,
  tabnames = frday87$tab.names)
kplot(sepan(wfri), clab.r = 2, clab.c = 1)
```

fruits	<i>Pair of Tables</i>
--------	-----------------------

Description

28 batches of fruits -two types- are judged by two different ways.

They are classified in order of preference, without ex aequo, by 16 individuals.

15 quantitative variables described the batches of fruits.

Usage

```
data(fruits)
```

Format

fruits is a list of 3 components:

typ is a vector returning the type of the 28 batches of fruits (peaches or nectarines).

jug is a data frame of 28 rows and 16 columns (judges).

var is a data frame of 28 rows and 16 measures (average of 2 judgements).

Details

fruits\$var is a data frame of 15 variables:

1. taches: quantity of cork blemishes (0=absent - maximum 5)
2. stries: quantity of stria (1/none - maximum 4)
3. abmucr: abundance of mucron (1/absent - 4)
4. irform: shape irregularity (0/none - 3)
5. allong: length of the fruit (1/round fruit - 4)
6. suroug: percentage of the red surface (minimum 40% - maximum 90%)
7. homlot: homogeneity of the intra-batch coloring (1/strong - 4)
8. homfru: homogeneity of the intra-fruit coloring (1/strong - 4)
9. pubesc: pubescence (0/none - 4)
10. verrou: intensity of green in red area (1/none - 4)
11. foncee: intensity of dark area (0/pink - 4)
12. comucr: intensity of the mucron color (1=no contrast - 4/dark)
13. impres: kind of impression (1/watched - 4/pointillé)
14. coldom: intensity of the predominating color (0/clear - 4)
15. calibr: grade (1/<90g - 5/>200g)

Source

Kervella, J. (1991) Analyse de l'attrait d'un produit : exemple d'une comparaison de lots de pêches. Agro-Industrie et méthodes statistiques. Compte-rendu des secondes journées européennes. Nantes 13-14 juin 1991. Association pour la Statistique et ses Utilisations, Paris, 313–325.

Examples

```
data(fruits)
par(mfrow = c(2,2))
pcajug <- dudi.pca(fruits$jug, scann = FALSE)
s.corcircle(pcajug$co)
s.class(pcajug$li, fac = fruits$type)

pcavar <- dudi.pca(fruits$var, scann = FALSE)
s.corcircle(pcavar$co)
s.class(pcavar$li, fac = fruits$type)

par(mfrow = c(1,1))
plot(coinertia(pcajug, pcavar, scan = FALSE))
```


fuzzygenet

*Reading a table of genetic data (diploid individuals)***Description**

Reads data like char2genet without a priori population

Usage

```
fuzzygenet(X)
```

Arguments

X a data frame of strings of characters (individuals in row, locus in variables), the value coded '000000' or two alleles of 6 characters

Details

In entry, a row is an individual, a variable is a locus and a value is a string of characters, for example, 012028 for a heterozygote carrying alleles 012 and 028; 020020 for a homozygote carrying two alleles 020 and 000000 for a not classified locus (missing data).

In exit, a fuzzy array with the following encoding for a locus:

0 0 1 ... 0 for a homozygote

0 0.5 0.5 ... 0 for a heterozygote

p1 p2 p3 ... pm for an unknown where (p1 p2 p3 ... pm) is the observed allelic frequencies for all the available data.

Value

returns a data frame with the 6 following attributes:

col.blocks	a vector containing the number of alleles by locus
all.names	a vector containing the names of alleles
loc.names	a vector containing the names of locus
row.w	a vector containing the uniform weighting of rows
col.freq	a vector containing the global allelic frequencies
col.num	a factor ranking the alleles by locus

Note

In the exit data frame, the alleles are numbered 1, 2, 3, ... by locus and the loci are called L01, L02, L03, ... for the simplification of listing. The original names are kept.

Author(s)

Daniel Chessel

References

~put references to the literature/web site here ~

See Also

[char2genet](#) if you have the a priori definition of the groups of individuals (populations). It may be used on the created object `dudi.fca`

Examples

```
data(casitas)
casitas[1:5, ]
casitas <- fuzzygenet(casitas)
attributes(casitas)
rm(casitas)
```

gearymoran	<i>Moran's I and Geary's c randomization tests for spatial and phylogenetic autocorrelation</i>
------------	---

Description

This function performs Moran's I test using phylogenetic and spatial link matrix (binary or general). It uses neighbouring weights so Moran's I and Geary's c randomization tests are equivalent.

Usage

```
gearymoran(bilis, X, nrepet = 999, alter=c("greater", "less", "two-sided"))
```

Arguments

`bilis` : a n by n link matrix where n is the row number of `X`
`X` : a data frame with continuous variables
`nrepet` : number of random vectors for the randomization test
`alter` : a character string specifying the alternative hypothesis, must be one of "greater" (default), "less" or "two-sided"

Details

`bilis` is a squared symmetric matrix which terms are all positive or null.

`bilis` is firstly transformed in frequency matrix `A` by dividing it by the total sum of data matrix :

$$a_{ij} = \frac{bilis_{ij}}{\sum_{i=1}^n \sum_{j=1}^n bilis_{ij}}$$

The neighbouring weights is defined by the matrix $D = \text{diag}(d_1, d_2, \dots)$ where $d_i = \sum_{j=1}^n bilis_{ij}$. For each vector x of the data frame `X`, the test is based on the Moran statistic $x^t A x$ where x is D-centred.

Value

Returns an object of class `krandtest` (randomization tests).

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>
Daniel Chessel

References

Cliff, A. D. and Ord, J. K. (1973) *Spatial autocorrelation*, Pion, London.

Thioulouse, J., Chessel, D. and Champely, S. (1995) Multivariate analysis of spatial patterns: a unified approach to local and global structures. *Environmental and Ecological Statistics*, **2**, 1–14.

See Also

[moran.test](#) and [geary.test](#) for classical versions of Moran's test and Geary's one

Examples

```
# a spatial example
data(mafragh)
tab0 <- (as.data.frame(scalewt(mafragh$mil)))
bilis0 <- neig2mat(mafragh$neig)
gm0 <- gearymoran(bilis0, tab0, 999)
gm0
plot(gm0, nclass = 20)

## Not run:
# a phylogenetic example
data(mjrochet)
mjr.phy <- newick2phylog(mjrochet$tre)
mjr.tab <- log(mjrochet$tab)
gearymoran(mjr.phy$Amat, mjr.tab)
gearymoran(mjr.phy$Wmat, mjr.tab)
par(mfrow = c(1,2))
table.value(mjr.phy$Wmat, csi = 0.25, clabel.r = 0)
table.value(mjr.phy$Amat, csi = 0.35, clabel.r = 0)
par(mfrow = c(1,1))

## End(Not run)
```

Description

There are multiple formats of genetic data. The functions of *ade4* associated genetic data use the class *genet*. An object of the class *genet* is a list containing at least one data frame whose lines are groups of individuals (populations) and columns alleles forming blocks associated with the locus. They contain allelic frequencies expressed as a percentage.

The function *char2genet* ensures the reading of tables crossing diploid individuals arranged by groups (populations) and polymorphic loci. Data frames containing only strings of characters are transformed in tables of allelic frequencies of the class *genet*. In entry a row is an individual, a variable is a locus and a value is a string of characters, for example '012028' for a heterozygote carrying alleles 012 and 028, '020020' for a homozygote carrying two alleles 020 and '000000' for a not classified locus (missing data).

The function *count2genet* reads data frames containing allelic countings by populations and allelic forms classified by locus.

The function *freq2genet* reads data frames containing allelic frequencies by populations and allelic forms classified by locus.

In these two cases, use as names of variables of strings of characters *xx.yyy* where *xx* are the names of locus and *yyy* a name of allelic forms in this locus. The analyses on this kind of data having to use compact labels, these functions classify the names of the populations, the names of the loci and the names of the allelic forms in vectors and re-code in a simple way starting with P for population, L for locus and 1, . . . , m for the alleles.

Usage

```
char2genet(X, pop, complete)
count2genet(PopAllCount)
freq2genet(PopAllFreq)
```

Arguments

<i>X</i>	a data frame of strings of characters (individuals in row, locus in variables), the value coded '000000' or two alleles of 6 characters
<i>pop</i>	a factor with the same number of rows than <i>df</i> classifying the individuals by population
<i>complete</i>	a logical value indicating a complete issue or not, by default FALSE
<i>PopAllCount</i>	a data frame containing integers: the occurrences of each allelic form (column) in each population (row)
<i>PopAllFreq</i>	a data frame containing values between 0 and 1: the frequencies of each allelic form (column) in each population (row)

Details

As a lot of formats for genetic data are published in literature, a list of class *genet* contains at least a table of allelic frequencies and an attribut *loc.blocks*. The populations (row) and the variables (column) are classified by alphabetic order. In the component *comp*, each individual per locus of *m* alleles is re-coded by a vector of length *m*: for hererozygicy 0, . . . , 1, . . . , 1, . . . , 0 and homozygocy 0, . . . , 2, 0.

Value

char2genet returns a list of class genet with :

\$tab	a frequencies table of poplations (row) and alleles (column)
\$center	the global frequency of each allelic form calculated on the overall individuals classified on each locus
\$pop.names	a vector containing the names of populations present in the data re-coded P01, P02, ...
\$all.names	a vector containing the names of the alleles present in the data re-coded L01.1, L01.2, ...
\$loc.blocks	a vector containing the number of alleles by loci
\$loc.fac	a factor sharing the alleles by loci
\$loc.names	a vector containing the names of loci present in the data re-coded L01, ..., L99
\$pop.loc	a data frame containing the number of genus allowing the calculation of frequencies
\$comp	the complete individual typing with the code 02000 or 01001 if the option complete is TRUE
\$comp.pop	a factor indicating the population if the option complete is TRUE

count2genet and freq2genet return a list of class genet which don't contain the components pop.loc and complete.

Author(s)

Daniel Chessel

Examples

```
data(casitas)
casitas[24,]
casitas.pop <- as.factor(rep(c("dome", "cast", "musc", "casi"), c(24,11,9,30)))
casi.genet <- char2genet(casitas, casitas.pop, complete=TRUE)
names(casi.genet$tab)
casi.genet$tab[,1:8]
casi.genet$pop.names
casi.genet$loc.names
casi.genet$all.names
casi.genet$loc.blocks # number of allelic forms by loci
casi.genet$loc.fac # factor classifying the allelic forms by locus
casi.genet$pop.loc # table populations loci
names(casi.genet$comp)
casi.genet$comp[1:4,]
casi.genet$comp.pop
casi.genet$center
apply(casi.genet$tab,2,mean)
casi.genet$pop.loc[,"L15"]
casi.genet$tab[, c("L15.1","L15.2")]
class(casi.genet)
```

```
casitas.coa <- dudi.coa(casi.genet$comp, scannf = FALSE)
s.class(casitas.coa$li, casi.genet$comp.pop)
```

ggtortoises

Microsatellites of Galapagos tortoises populations

Description

This data set gives genetic relationships between Galapagos tortoises populations with 10 microsatellites.

Usage

```
data(ggtortoises)
```

Format

ggtortoises is a list of 6 components.

area is a data frame designed to be used in area.plot function.

ico is a list of three pixmap icons representing the tortoises morphotypes.

pop is a data frame containing meta informations about populations.

misc is a data frame containing the coordinates of the island labels.

loc is a numeric vector giving the number of alleles by marker.

tab is a data frame containing the number of alleles by populations for 10 microsatellites.

Source

M.C. Ciofi, C. Milinkovitch, J.P. Gibbs, A. Caccone, and J.R. Powell (2002) Microsatellite analysis of genetic divergence among populations of giant galapagos tortoises. *Molecular Ecology* **11**: 2265-2283.

References

M.C. Ciofi, C. Milinkovitch, J.P. Gibbs, A. Caccone, and J.R. Powell (2002) Microsatellite analysis of genetic divergence among populations of giant galapagos tortoises. *Molecular Ecology* **11**: 2265-2283.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps069.pdf> (in French).

Examples

```

if(require(pixmap, quiet=TRUE)){
  data(ggtortoises)
  a1 <- ggtortoises$area
  area.plot(a1)
  rect(min(a1$x), min(a1$y), max(a1$x), max(a1$y), col = "lightblue")
  invisible(lapply(split(a1, a1$id), function(x) polygon(x[, -1], col = "white")))
  s.label(ggtortoises$misc, grid=FALSE, include.ori=FALSE, addaxes=FALSE, add.p=TRUE)
  listico <- ggtortoises$ico[as.character(ggtortoises$pop$carap)]
  s.logo(ggtortoises$pop, listico, add.p=TRUE)
}

```

granulo

*Granulometric Curves***Description**

This data set gives the repartition in diameter classes of deposit samples.

Usage

```
data(granulo)
```

Format

granulo is a list of 2 components.

tab contains the 49 deposit samples, 9 diameter classes, weight of grains by size class

born contains the boundaries of the diameter classes

Source

Gaschignard-Fossati, O. (1986) *Répartition spatiale des macroinvertébrés benthiques d'un bras vif du Rhône. Rôle des crues et dynamique saisonnière*. Thèse de doctorat, Université Lyon 1.

Examples

```

data(granulo)
w <- t(apply(granulo$tab, 1, function(x) x / sum(x)))
w <- data.frame(w)
wtr <- data.frame(t(w))
wmoy <- data.frame(matrix(apply(wtr, 1, mean), 1))
d1 <- dudi.pca(w, scal = FALSE, scan = FALSE)
wmoy <- suprow(d1, wmoy)$lisup
s.arrow(d1$c1, clab = 1.5)
s.distri(d1$c1, wtr, cstar = 0.33, cell = 0,
  axesell = FALSE, add.p = TRUE, clab = 0.75)
s.label(wmoy, cpoi = 5, clab = 0, add.p = TRUE)

```

`gridrowcol`*Complete regular grid analysis*

Description

This function defines objects to analyse data sets associated with complete regular grid.

Usage

```
gridrowcol(nrow, ncol, cell.names = NULL)
```

Arguments

<code>nrow</code>	size of the grid (number of rows)
<code>ncol</code>	size of the grid (number of columns)
<code>cell.names</code>	grid cell labels

Value

Returns a list containing the following items :

<code>xy</code>	: a data frame with grid cell coordinates
<code>area</code>	: a data frame with three variables to display grid cells as areas
<code>neig</code>	: an object of class 'neig' corresponding to a neighbouring graph of the grid (rook case)
<code>orthobasis</code>	: an object of class 'orthobasis' corresponding to the analytical solution for the neighbouring graph

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>
Daniel Chessel

References

Méot, A., Chessel, D. and Sabatier, D. (1993) Opérateurs de voisinage et analyse des données spatio-temporelles. *in* J.D. Lebreton and B. Asselain, editors. *Biométrie et environnement*. Masson, 45-72.

Cornillon, P.A. (1998) *Prise en compte de proximités en analyse factorielle et comparative*. Thèse, Ecole Nationale Supérieure Agronomique, Montpellier.

See Also

[orthobasis](#), [orthogram](#), [mld](#)

Examples

```
w <- gridrowcol(8,5)
par(mfrow = c(1,2))
area.plot(w$area,center = w$xy, graph = w$neig, clab = 0.75)
area.plot(w$area,center = w$xy, graph = w$neig, clab = 0.75,
  label = as.character(1:40))
par(mfrow = c(1,1))

par(mfrow = c(5,8))
for(k in 1:39)
  s.value(w$xy, w$orthobasis[,k], csi = 3, cleg = 0, csub = 2,
    sub = as.character(signif(attr(w$orthobasis, "values")[k],3)),
    incl = FALSE, addax = FALSE, cgr = 0, ylim = c(0,10))
par(mfrow = c(1,1))
```

hdpj

*Genetic Variation In Human Populations***Description**

This data set gives genotypes variation of 1066 individuals belonging to 52 predefined populations, for 404 microsatellite markers.

Usage

```
data(hdpj)
```

Format

hdpj is a list of 3 components.

tab is a data frame with the genotypes of 1066 individuals encoded with 6 characters (individuals in row, locus in column), for example '123098' for a heterozygote carrying alleles '123' and '098', '123123' for a homozygote carrying two alleles '123' and, '000000' for a not classified locus (missing data).

ind is a data frame with 4 columns containing information about the 1066 individuals: hdpj\$ind\$id containing the Diversity Panel identification number of each individual, and three factors hdpj\$ind\$sex, hdpj\$ind\$population and hdpj\$ind\$region containing the names of the 52 populations belonging to 7 major geographic regions (see details).

locus is a dataframe containing four columns: hdpj\$locus\$marknames a vector of names of the microsatellite markers, hdpj\$locus\$allbyloc a vector containing the number of alleles by loci, hdpj\$locus\$chromosome a factor defining a number for one chromosome and, hdpj\$locus\$maposition indicating the position of the locus in the chromosome.

Details

The rows of `hdp$pop` are the names of the 52 populations belonging to the geographic regions contained in the rows of `hdp$region`. The chosen regions are: America, Asia, Europe, Middle East North Africa, Oceania, Subsaharan AFRICA.

The 52 populations are: Adygei, Balochi, Bantu, Basque, Bedouin, Bergamo, Biaka Pygmies, Brahui, Burusho, Cambodian, Columbian, Dai, Daur, Druze, French, Han, Hazara, Hezhen, Japanese, Kalash, Karitiana, Lahu, Makrani, Mandenka, Maya, Mbuti Pygmies, Melanesian, Miaozi, Mongola, Mozabite, Naxi, NewGuinea, Nilote, Orcadian, Oroqen, Palestinian, Pathan, Pima, Russian, San, Sardinian, She, Sindhi, Surui, Tu, Tujia, Tuscan, Uygur, Xibo, Yakut, Yizu, Yoruba.

`hdp$freq` is a data frame with 52 rows, corresponding to the 52 populations described above, and 4992 microsatellite markers.

Source

Extract of data prepared by the Human Diversity Panel Genotypes <http://research.marshfieldclinic.org/genetics/Freq/FreqInfo.htm>

prepared by Hinda Haned, from data used in: Noah A. Rosenberg, Jonatahan K. Pritchard, James L. Weber, Howard M. Cabb, Kenneth K. Kidds, Lev A. Zhivotovsky, Marcus W. Feldman (2002) Genetic Structure of human Populations *Science*, **298**, 2381–2385.

Lev A. Zhivotovsky, Noah Rosenberg, and Marcus W. Feldman (2003). Features of Evolution and Expansion of Modern Humans, Inferred from Genomewide Microsatellite Markers *Am. J. Hum. Genet.*, **72**, 1171–1186.

Examples

```
## Not run:
library(ade4)
data(hdp)
freq <- char2genet(hdp$stab, hdp$ind$population)
vec <- apply(freq$stab, 2, function(c) mean(c, na.rm = TRUE))
for (j in 1:4492){
  freq$stab[is.na(freq$stab[,j]),j] = vec[j]}
pcatot <- dudi.pca(freq$stab, center = TRUE, scale = FALSE, scannf = FALSE, nf = 4)
s.label(pcatot$li, xax = 1, yax = 2, sub = "1-2", lab = freq$pop.names)

## End(Not run)
```

housetasks

Contingency Table

Description

The `housetasks` data frame gives 13 `housetasks` and their repartition in the couple.

Usage

```
data(housetasks)
```

Format

This data frame contains four columns : wife, alternating, husband and jointly. Each column is a numeric vector.

Source

Kroonenberg, P. M. and Lombardo, R. (1999) Nonsymmetric correspondence analysis: a tool for analysing contingency tables with a dependence structure. *Multivariate Behavioral Research*, **34**, 367–396

Examples

```
data(housetasks)
nsc1 <- dudi.nsc(housetasks, scan = FALSE)
s.label(nsc1$c1, clab = 1.25)
s.arrow(nsc1$i1, add.pl = TRUE, clab = 0.75)
```

humDNAm

human mitochondrial DNA restriction data

Description

This data set gives the frequencies of haplotypes of mitochondrial DNA restriction data in ten populations all over the world.

It gives also distances among the haplotypes.

Usage

```
data(humDNAm)
```

Format

humDNAm is a list of 3 components.

distances is an object of class `dist` with 56 haplotypes. These distances are computed by counting the number of differences in restriction sites between two haplotypes.

samples is a data frame with 56 haplotypes, 10 abundance variables (populations). These variables give the haplotype abundance in a given population.

structures is a data frame with 10 populations, 1 variable (classification). This variable gives the name of the continent in which a given population is located.

Source

Excoffier, L., Smouse, P.E. and Quattro, J.M. (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics*, **131**, 479–491.

Examples

```
data(humDNAm)
dpcoa(humDNAm) <- dpcoa(humDNAm$samples,
  sqrt(humDNAm$distances), scan = FALSE, nf = 2)
plot(dpcoa(humDNAm), csize = 1.5)
```

 ichtyo

Point sampling of fish community

Description

This data set gives informations between a faunistic array, the total number of sampling points made at each sampling occasion and the year of the sampling occasion.

Usage

```
data(ichtyo)
```

Format

`ichtyo` is a list of 3 components.

tab is a faunistic array with 9 columns and 32 rows.

eff is a vector of the 32 sampling effort.

dat is a factor where the levels are the 10 years of the sampling occasion.

Details

The value $n(i,j)$ at the i th row and the j th column in `tab` corresponds to the number of sampling points of the i th sampling occasion (in `eff`) that contains the j th species.

Source

Dolédec, S., Chessel, D. and Olivier, J. M. (1995) L'analyse des correspondances décentrée: application aux peuplements ichtyologiques du haut-Rhône. *Bulletin Français de la Pêche et de la Pisciculture*, **336**, 29–40.

Examples

```
data(ichtyo)
dudi1 <- dudi.dec(ichtyo$tab, ichtyo$eff, scan = FALSE)
s.class(dudi1$li, ichtyo$dat, wt = ichtyo$eff / sum(ichtyo$eff))
```

inertia.dudi	<i>Statistics of inertia in a one-table analysis</i>
--------------	--

Description

Prints of the statistics of inertia in a one-table analysis

Usage

```
inertia.dudi(dudi, row.inertia = FALSE, col.inertia = FALSE)
```

Arguments

dudi	a duality diagram, object of class dudi
row.inertia	if TRUE, returns the statistics of the decomposition of inertia for the rows
col.inertia	if TRUE, returns the statistics of the decomposition of inertia for the columns

Details

Contributions are printed in 1/10000 and the sign is the sign of the coordinate

Value

a list containing :

TOT	repartition of the total inertia between axes
row.abs	absolute contributions of the decomposition of inertia for the rows
row.rel	relative contributions of the decomposition of inertia for the rows
row.cum	cumulative relative contributions of the decomposition of inertia for the rows
col.abs	absolute contributions of the decomposition of inertia for the columns
col.rel	relative contributions of the decomposition of inertia for the columns
col.cum	cumulative relative contributions of the decomposition of inertia for the columns

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

- Lebart, L., Morineau, A. and Tabart, N. (1977) *Techniques de la description statistique, méthodes et logiciels pour la description des grands tableaux*, Dunod, Paris, 61–62.
- Volle, M. (1981) *Analyse des données*, Economica, Paris, 89–90 and 118
- Lebart, L., Morineau, L. and Warwick, K.M. (1984) *Multivariate descriptive analysis: correspondence and related techniques for large matrices*, John Wiley and Sons, New York.
- Greenacre, M. (1984) *Theory and applications of correspondence analysis*, Academic Press, London, 66.
- Rouanet, H. and Le Roux, B. (1993) *Analyse des données multidimensionnelles*, Dunod, Paris, 143–144.
- Tenenhaus, M. (1994) *Méthodes statistiques en gestion*, Dunod, Paris, p. 160, 161, 166, 204.
- Lebart, L., Morineau, A. and Piron, M. (1995) *Statistique exploratoire multidimensionnelle*, Dunod, Paris, p. 56,95-96.

Examples

```
data(housetasks)
coa1 <- dudi.coa(housetasks, scann = FALSE)
inertia.dudi(coa1, col = TRUE, row = FALSE)$col.rel
```

irishdata

Geary's Irish Data

Description

This data set contains geographical informations about 25 counties of Ireland.

Usage

```
data(irishdata)
```

Format

`irishdata` is a list of 11 objects.

area is a data frame with polygons for each of the 25 contiguous counties.

county.names is a vector with the names of the 25 counties.

xy is a data frame with the coordinates centers of the 25 counties.

tab is a data frame with 25 rows (counties) and 12 variables.

contour is a data frame with the global polygon of all the 25 counties.

link is a matrix containing the common length between two counties from `area`.

area.utm is a data frame with polygons for each of the 25 contiguous counties expressed in Universal Transverse Mercator (UTM) coordinates.

xy.utm is a data frame with the UTM coordinates centers of the 25 counties.

link.utm is a matrix containing the common length between two counties from `area.utm`.

tab.utm is a data frame with the 25 counties (explicitly named) and 12 variables.

contour.utm is a data frame with the global polygon of all the 25 counties expressed in UTM coordinates.

Source

Geary, R.C. (1954) The contiguity ratio and statistical mapping. *The incorporated Statistician*, **5**, 3, 115–145.

Cliff, A.D. and Ord, J.K. (1973) *Spatial autocorrelation*, Pion, London. 1–178.

Examples

```
data(irishdata)
par(mfrow = c(2,2))
area.plot(irishdata$area, lab = irishdata$county.names, clab = 0.75)
area.plot(irishdata$area)
apply(irishdata$contour, 1, function(x) segments(x[1],x[2],x[3],x[4],
  lwd = 3))
s.corcircle(dudi.pca(irishdata$tab, scan = FALSE)$co)
score <- dudi.pca(irishdata$tab, scan = FALSE, nf = 1)$li$Axis1
names(score) <- row.names(irishdata$tab)
area.plot(irishdata$area, score)
par(mfrow = c(1,1))
```

is.euclid

Is a Distance Matrix Euclidean ?

Description

Confirmation of the Euclidean nature of a distance matrix by the Gower's theorem.

`is.euclid` is used in `summary.dist`.

Usage

```
is.euclid(distmat, plot = FALSE, print = FALSE, tol = 1e-07)
## S3 method for class 'dist'
summary(object, ...)
```

Arguments

distmat	an object of class 'dist'
plot	a logical value indicating whether the eigenvalues bar plot of the matrix of the term $-\frac{1}{2}d_{ij}^2$ centred by rows and columns should be displayed
print	a logical value indicating whether the eigenvalues of the matrix of the term $-\frac{1}{2}d_{ij}^2$ centred by rows and columns should be printed
tol	a tolerance threshold : an eigenvalue is considered positive if it is larger than $-\text{tol} \times \text{lambda1}$ where lambda1 is the largest eigenvalue.
object	an object of class 'dist'
...	further arguments passed to or from other methods

Value

returns a logical value indicating if all the eigenvalues are positive or equal to zero

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

Gower, J.C. and Legendre, P. (1986) Metric and Euclidean properties of dissimilarity coefficients. *Journal of Classification*, **3**, 5–48.

Examples

```
w <- matrix(runif(10000), 100, 100)
w <- dist(w)
summary(w)
is.euclid(w) # TRUE
w <- quasieuclid(w) # no correction need in: quasieuclid(w)
w <- lingoew(w) # no correction need in: lingoew(w)
w <- cailliez(w) # no correction need in: cailliez(w)
rm(w)
```

julliot

Seed dispersal

Description

This data set gives the spatial distribution of seeds (quadrats counts) of seven species in the understory of tropical rainforest.

Usage

```
data(julliot)
```


Format

julliot is a list containing the 3 following objects :

tab is a data frame with 160 rows (quadrats) and 7 variables (species).

xy is a data frame with the coordinates of the 160 quadrats (positioned by their centers).

area is a data frame with 3 variables returning the boundary lines of each quadrat. The first variable is a factor. The levels of this one are the row.names of tab. The second and third variables return the coordinates (x,y) of the points of the boundary line.

Species names of julliot\$tab are *Pouteria torta*, *Minquartia guianensis*, *Quiina obovata*, *Chrysophyllum lucentifolium*, *Parahancornia fasciculata*, *Virola michelii*, *Pourouma spp.*

References

Julliot, C. (1992) Utilisation des ressources alimentaires par le singe hurleur roux, *Alouatta seniculus* (Atelidae, Primates), en Guyane : impact de la dissémination des graines sur la régénération forestière. Thèse de troisième cycle, Université de Tours.

Julliot, C. (1997) Impact of seed dispersal by red howler monkeys *Alouatta seniculus* on the seedling population in the understorey of tropical rain forest. *Journal of Ecology*, **85**, 431–440.

Examples

```
data(julliot)
par(mfrow = c(3,3))
## Not run:
for(k in 1:7)
  area.plot(julliot$area, val = log(julliot$tab[,k]+1),
            sub = names(julliot$tab)[k], csub = 2.5)

## End(Not run)

if (require(splancs, quiet = TRUE)){
  par(mfrow = c(3,3))
  for(k in 1:7)
    s.image(julliot$xy, log(julliot$tab[,k]+1), kgrid = 3, span = 0.25,
            sub = names(julliot$tab)[k], csub = 2.5)
}

## Not run:
par(mfrow = c(3,3))
for(k in 1:7) {
  area.plot(julliot$area)
  s.value(julliot$xy, scalewt(log(julliot$tab[,k]+1)),
          sub = names(julliot$tab)[k], csub = 2.5, add.p = TRUE)
}
## End(Not run)
par(mfrow = c(3,3))
for(k in 1:7)
  s.value(julliot$xy, log(julliot$tab[,k]+1),
          sub = names(julliot$tab)[k], csub = 2.5)
```

```
## Not run:
if (require(spdep, quiet = TRUE)){
  par(mfrow = c(1,1))
  neig0 <- nb2neig(dnearneigh(as.matrix(julliot$xy), 1, 1.8))
  s.label(julliot$xy, neig = neig0, clab = 0.75, incl = FALSE,
    addax = FALSE, grid = FALSE)

  gearymoran(neig.util.LtoG(neig0), log(julliot$tab+1))
  orthogram(log(julliot$tab[,3]+1), ortho = scores.neig(neig0),
    nrepet = 9999)}

## End(Not run)
```

jv73

K-tables Multi-Regions

Description

This data set gives physical and physico-chemical variables, fish species, spatial coordinates about 92 sites.

Usage

```
data(jv73)
```

Format

`jv73` is a list of 6 components.

morpho is a data frame with 92 sites and 6 physical variables.

phychi is a data frame with 92 sites and 12 physico-chemical variables.

poi is a data frame with 92 sites and 19 fish species.

xy is a data frame with 92 sites and 2 spatial coordinates.

contour is a data frame for mapping.

fac.riv is a factor distributing the 92 sites on 12 rivers.

Source

Verneaux, J. (1973) Cours d'eau de Franche-Comté (Massif du Jura). Recherches écologiques sur le réseau hydrographique du Doubs. Essai de biotypologie. Thèse d'Etat, Besançon.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps047.pdf> (in French).

Examples

```

data(jv73)
s.label(jv73$xy, contour = jv73$contour, incl = FALSE,
        clab = 0.75)
s.class(jv73$xy, jv73$fac.riv, add.p = TRUE, cell = 0,
        axese = FALSE, csta = 0, cpoi = 0, clab = 1.25)

w <- split(jv73$morpho, jv73$fac.riv)
w <- lapply(w, function(x) t(dudi.pca(x, scann = FALSE)))
w <- ktab.list.dudi(w)
kplot(sepan(w), perm = TRUE, clab.r = 0, clab.c = 2, show = FALSE)

```

kcponds

Ponds in a nature reserve

Description

This data set contains informations about 33 ponds in De Maten reserve (Genk, Belgium).

Usage

```
data(kcponds)
```

Format

tab : a data frame with 15 environmental variables(columns) on 33 ponds(rows)

area : an object of class `area`

xy : a data frame with the coordinates of ponds

neig : an object of class `neig`

Details

Variables of `kcponds$tab` are the following ones : `depth`, `area`, `O2` (oxygen concentration), `cond` (conductivity), `pH`, `Fe` (Fe concentration), `secchi` (Secchi disk depth), `N` (NNO concentration), `TP` (total phosphorus concentration), `chl.a` (chlorophyll-a concentration), `EM` (emergent macrophyte cover), `FM` (floating macrophyte cover), `SM` (submerged macrophyte cover), `denMI` (total density of macroinvertebrates), `divMI` (diversity macroinvertebrates)

Source

Cottenie, K. (2002) Local and regional processes in a zooplankton metacommunity. PhD, Katholieke Universiteit Leuven, Leuven, Belgium.

<http://www.kuleuven.ac.be/bio/eco/phdkarlcottenie.pdf>

Examples

```

data(kcponds)

par(mfrow=c(3,1))
area.plot(kcponds$area)
s.label(kcponds$xy,add.p = TRUE, cpoi = 2, clab = 0)
s.label(kcponds$xy,add.p = TRUE, cpoi = 3, clab = 0)
s.label(kcponds$xy,add.p = TRUE, cpoi = 0, clab = 0,
        neig = kcponds$neig, cneig = 1)
area.plot(kcponds$area)
s.label(kcponds$xy, add.p = TRUE, clab = 1.5)
w <- as.numeric(scalewt(kcponds$tab$N))
s.value(kcponds$xy, w, cleg = 2, sub = "Nitrogen concentration",
        csub = 4, possub = "topright", include = FALSE)
par(mfrow = c(1,1))
## Not run:
par(mfrow=c(3,1))
pca1 <- dudi.pca(kcponds$tab, scan = FALSE, nf = 4)
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
multi1 <- multispati(pca1, nb2listw(neig2nb(kcponds$neig)),
                    scan = FALSE, nfposi = 2, nfnega = 1)
summary(multi1)}
par(mfrow = c(1,1))

## End(Not run)

```

kdist

the class of objects 'kdist' (K distance matrices)

Description

An object of class `kdist` is a list of distance matrices observed on the same individuals

Usage

```
kdist(..., epsi = 1e-07, upper = FALSE)
```

Arguments

<code>...</code>	a sequence of objects of the class <code>kdist</code> .
<code>epsi</code>	a tolerance threshold to test if distances are Euclidean (Gower's theorem) using $\frac{\lambda_m}{\lambda_1}$ is larger than <code>-epsi</code> .
<code>upper</code>	a logical value indicating whether the upper of a distance matrix is used (TRUE) or not (FALSE).

Details

The attributes of a 'kdist' object are:
 names: the names of the distances
 size: the number of points between distances are known
 labels: the labels of points
 euclid: a logical vector indicating whether each distance of the list is Euclidean or not.
 call: a call order
 class: object 'kdist'

Value

returns an object of class 'kdist' containing a list of semidefinite matrices.

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Gower, J. C. (1966) Some distance properties of latent root and vector methods used in multivariate analysis. *Biometrika*, **53**, 325–338.

Examples

```
# starting from a list of matrices
data(yanomama)
lapply(yanomama,class)
kd1 = kdist(yanomama)
print(kd1)

# giving the correlations of Mantel's test
cor(as.data.frame(kd1))
pairs(as.data.frame(kd1))

# starting from a list of objects 'dist'
data(friday87)
fri.w <- ktab.data.frame(friday87$fau, friday87$fau.blo,
  tabnames = friday87$tab.names)
fri.kd = lapply(1:10, function(x) dist.binary(fri.w[[x]],2))
names(fri.kd) = friday87$tab.names
unlist(lapply(fri.kd,class)) # a list of distances
fri.kd = kdist(fri.kd)
fri.kd
s.corcircle(dudi.pca(as.data.frame(fri.kd), scan = FALSE)$co)

# starting from several distances
data(ecomor)
d1 <- dist.binary(ecomor$habitat, 1)
d2 <- dist.prop(ecomor$forsub, 5)
```

```

d3 <- dist.prop(ecomor$diet, 5)
d4 <- dist.quant(ecomor$morpho, 3)
d5 <- dist.taxo(ecomor$taxo)
ecomor.kd <- kdist(d1, d2, d3, d4, d5)
names(ecomor.kd) = c("habitat", "forsub", "diet", "morpho", "taxo")
class(ecomor.kd)
s.corcircle(dudi.pca(as.data.frame(ecomor.kd), scan = FALSE)$co)

data(bsetal97)
X <- prep.fuzzy.var(bsetal97$biol, bsetal97$biol.blo)
w1 <- attr(X, "col.num")
w2 <- levels(w1)
w3 <- lapply(w2, function(x) dist.quant(X[,w1==x], method = 1))
names(w3) <- names(attr(X, "col.blocks"))
w3 <- kdist(list = w3)
s.corcircle(dudi.pca(as.data.frame(w3), scan = FALSE)$co)

data(rpjd1)
w1 = lapply(1:10, function(x) dist.binary(rpjd1$fau, method = x))
w2 = c("JACCARD", "SOCKAL_MICHENER", "SOCKAL_SNEATH_S4", "ROGERS_TANIMOTO")
w2 = c(w2, "CZEKANOWSKI", "S9_GOWER_LEGENDRE", "OCHIAI", "SOKAL_SNEATH_S13")
w2 <- c(w2, "Phi_PEARSON", "S2_GOWER_LEGENDRE")
names(w1) <- w2
w3 = kdist(list = w1)
w4 <- dudi.pca(as.data.frame(w3), scan = FALSE)$co
w4

```

kdist2ktab

Transformation of K distance matrices (object 'kdist') into K Euclidean representations (object 'ktab')

Description

The function creates a ktab object with the Euclidean representations from a kdist object. Notice that the euclid attribute must be TRUE for all elements.

Usage

```
kdist2ktab(kd, scale = TRUE, tol = 1e-07)
```

Arguments

kd	an object of class kdist
scale	a logical value indicating whether the inertia of Euclidean representations are equal to 1 (TRUE) or not (FALSE).
tol	a tolerance threshold, an eigenvalue is considered equal to zero if eig\$values > (eig\$values[1 * tol)

Value

returns a list of class ktab containing for each distance of kd the data frame of its Euclidean representation

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```
data(friday87)
fri.w <- ktab.data.frame(friday87$fau, friday87$fau.blo,
  tabnames = friday87$tab.names)
fri.kd <- lapply(1:10, function(x) dist.binary(fri.w[[x]], 10))
names(fri.kd) <- substr(friday87$tab.names, 1, 4)
fri.kd <- kdist(fri.kd)
fri.ktab = kdist2ktab(kd = fri.kd)
fri.sepan = sepan(fri.ktab)
plot(fri.sepan, csub = 3)

tapply(fri.sepan$Eig, fri.sepan$TC[,1], sum)
# the sum of the eigenvalues is constant and equal to 1, for each K tables

fri.statis <- stasis(fri.ktab, scan = FALSE, nf = 2)
round(fri.statis$RV, dig = 2)

fri.mfa <- mfa(fri.ktab, scan = FALSE, nf = 2)
fri.mcoa <- mcoa(fri.ktab, scan = FALSE, nf = 2)

apply(fri.statis$RV, 1, mean)
fri.statis$RV.tabw
plot(apply(fri.statis$RV, 1, mean), fri.statis$RV.tabw)
plot(fri.statis$RV.tabw, fri.statis$RV.tabw)
```

kdisteuclid

a way to obtain Euclidean distance matrices

Description

a way to obtain Euclidean distance matrices

Usage

```
kdisteuclid(obj, method = c("lingoes", "cailliez", "quasi"))
```

Arguments

obj an object of class kdist
method a method to convert a distance matrix in a Euclidean one

Value

returns an object of class `kdist` with all distances Euclidean.

Note

according to the program `DistPCoa` of P. Legendre and M.J. Anderson
<http://www.fas.umontreal.ca/BIOLOGIE/Casgrain/en/labo/distpcoa.html>

Author(s)

Daniel Chessel
 Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

- Gower, J.C. and Legendre, P. (1986) Metric and Euclidean properties of dissimilarity coefficients. *Journal of Classification*, **3**, 5–48.
- Cailliez, F. (1983) The analytical solution of the additive constant problem. *Psychometrika*, **48**, 305–310.
- Lingoes, J.C. (1971) Somme boundary conditions for a monotone analysis of symmetric matrices. *Psychometrika*, **36**, 195–203.
- Legendre, P. and Anderson, M.J. (1999) Distance-based redundancy analysis: testing multispecies responses in multifactorial ecological experiments. *Ecological Monographs*, **69**, 1–24.
- Legendre, P., and L. Legendre. (1998) Numerical ecology, 2nd English edition edition. Elsevier Science BV, Amsterdam.

Examples

```
w <- c(0.8,0.8,0.377350269,0.8,0.377350269,0.377350269) # see ref.
w <- kdist(w)
w1 <- c(kdisteuclid(kdist(w), "lingoes"), kdisteuclid(kdist(w), "cailliez"),
       kdisteuclid(kdist(w), "quasi"))
print(w, print = TRUE)
print(w1, print = TRUE)

data(eurodist)
par(mfrow = c(1, 3))
eu1 <- kdist(eurodist) # an object of class 'dist'
plot(data.frame(unclass(c(eu1, kdisteuclid(eu1, "quasi")))), asp = 1)
title(main = "Quasi") ; abline(0,1)
plot(data.frame(unclass(c(eu1, kdisteuclid(eu1, "lingoes")))), asp = 1)
title(main = "Lingoes") ; abline(0,1)
plot(data.frame(unclass(c(eu1, kdisteuclid(eu1, "cailliez")))), asp = 1)
title(main = "Cailliez") ; abline(0,1)
```

kplot

Generic Function for Multiple Graphs in a K-tables Analysis

Description

Methods for `foucart`, `mcoa`, `mfa`, `pta`, `sepan`, `sepan.coa` and `statis`

Usage

```
kplot(object, ...)
```

Arguments

<code>object</code>	an object used to select a method
<code>...</code>	further arguments passed to or from other methods

Examples

```
methods(plot)
methods(scatter)
methods(kplot)
```

kplot.foucart

Multiple Graphs for the Foucart's Correspondence Analysis

Description

performs high level plots of a Foucart's Correspondence Analysis, using an object of class `foucart`.

Usage

```
## S3 method for class 'foucart'
kplot(object, xax = 1, yax = 2, mfrow = NULL,
       which.tab = 1:length(object$blo), clab.r = 1, clab.c = 1.25,
       csub = 2, possub = "bottomright", ...)
```

Arguments

<code>object</code>	an object of class <code>foucart</code>
<code>xax</code> , <code>yax</code>	the numbers of the x-axis and the y-axis
<code>mfrow</code>	a vector of the form <code>'c(nr,nc)'</code> , otherwise computed by as special own function <code>n2mfrow</code>
<code>which.tab</code>	vector of table numbers for analyzing
<code>clab.r</code>	a character size for the row labels

clab.c	a character size for the column labels
csub	a character size for the sub-titles used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(bf88)
fou1 <- foucart(bf88, scann = FALSE, nf = 3)
kplot(fou1, clab.r = 0, clab.c = 2, csub = 3)
```

kplot.mcoa

Multiple Graphs for a Multiple Co-inertia Analysis

Description

performs high level plots of a Multiple Co-inertia Analysis, using an object of class mcoa.

Usage

```
## S3 method for class 'mcoa'
kplot(object, xax = 1, yax = 2, which.tab = 1:nrow(object$cov2),
      mfrow = NULL, option = c("points", "axis", "columns"),
      clab = 1, cpoint = 2, csub = 2, possub = "bottomright",...)
```

Arguments

object	an object of class mcoa
xax, yax	the numbers of the x-axis and the y-axis
which.tab	a numeric vector containing the numbers of the tables to analyse
mfrow	a vector of the form 'c(nr,nc)', otherwise computed by as special own function n2mfrow
option	a string of characters for the drawing option "points" plot of the projected scattergram onto the co-inertia axes "axis" projections of inertia axes onto the co-inertia axes. "columns" projections of variables onto the synthetic variables planes.
clab	a character size for the labels
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn.

csub	a character size for the sub-titles, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(friday87)
w1 <- data.frame(scale(friday87$fau, scal = FALSE))
w2 <- ktab.data.frame(w1, friday87$fau.blo,
  tabnames = friday87$tab.names)
mcoa1 <- mcoa(w2, "lambda1", scan = FALSE)
kplot(mcoa1, clab = 2, csub = 2, cpoi = 3, opt = "axis")

kplot(mcoa1, mfrow = c(3,4), clab = 2, csub = 3, cpoi = 3)

kplot(mcoa1, clab = 2, csub = 3, cpoi = 3, mfrow = c(3,4),
  opt = "columns")
```

kplot.mfa

*Multiple Graphs for a Multiple Factorial Analysis***Description**

performs high level plots of a Multiple Factorial Analysis, using an object of class `mfa`.

Usage

```
## S3 method for class 'mfa'
kplot(object, xax = 1, yax = 2, mfrow = NULL,
  which.tab = 1:length(object$blo), row.names = FALSE, col.names = TRUE,
  traject = FALSE, permute.row.col = FALSE,
  clab = 1, csub = 2, possub = "bottomright", ...)
```

Arguments

object	an object of class <code>mfa</code>
xax, yax	the numbers of the x-axis and the y-axis
mfrow	a vector of the form 'c(nr,nc)', otherwise computed by a special own function <code>n2mfrow</code>
which.tab	vector of the numbers of tables used for the analysis
row.names	a logical value indicating whether the row labels should be inserted

col.names	a logical value indicating whether the column labels should be inserted
traject	a logical value indicating whether the trajectories of the rows should be drawn in a natural order
permute.row.col	if TRUE, the rows are represented by vectors and columns by points, otherwise it is the opposite
clab	a character size for the labels
csub	a character size for the sub-titles, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(friday87)
w1 <- data.frame(scale(friday87$fau, scal = FALSE))
w2 <- ktab.data.frame(w1, friday87$fau.blo,
  tabnames = friday87$tab.names)
mfa1 <- mfa(w2, scann = FALSE)
kplot(mfa1)
```

kplot.pta

*Multiple Graphs for a Partial Triadic Analysis***Description**

performs high level plots of a Partial Triadic Analysis, using an object of class pta.

Usage

```
## S3 method for class 'pta'
kplot(object, xax = 1, yax = 2, which.tab = 1:nrow(object$RV),
  mfrow = NULL, which.graph = 1:4, clab = 1, cpoint = 2, csub = 2,
  possub = "bottomright", ask = par("ask"), ...)
```

Arguments

object	an object of class pta
xax, yax	the numbers of the x-axis and the y-axis
which.tab	a numeric vector containing the numbers of the tables to analyse
mfrow	parameter of the array of figures to be drawn, otherwise the graphs associated to a table are drawn on the same row

which.graph	an option for drawing, an integer between 1 and 4. For each table of which.tab, are drawn : <ol style="list-style-type: none"> 1 the projections of the principal axes 2 the projections of the rows 3 the projections of the columns 4 the projections of the principal components onto the planes of the compromise
clab	a character size for the labels
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn.
csub	a character size for the sub-titles, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
ask	a logical value indicating if the graphs requires several arrays of figures
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(meaudret)
wit1 <- wca(dudi.pca(meaudret$spe, scan = FALSE, scal = FALSE),
  meaudret$design$season, scan = FALSE)
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5"), 4))
kta2 <- t(kta1) ; pta1 <- pta(kta2, scann = FALSE)
kplot(pta1, clab = 1.5, csub = 3)
kplot(pta1, clab = 1.5, csub = 3, which.graph = 3, mfrow = c(2,2))
```

kplot.sepan

*Multiple Graphs for Separated Analyses in a K-tables***Description**

performs high level plots for Separated Analyses in a K-tables, using an object of class sepan.

Usage

```
## S3 method for class 'sepan'
kplot(object, xax = 1, yax = 2, which.tab = 1:length(object$blo),
  mfrow = NULL, permute.row.col = FALSE, clab.row = 1,
  clab.col = 1.25, traject.row = FALSE, csub = 2,
  possub = "bottomright", show.eigen.value = TRUE,...)

## S3 method for class 'sepan.coa'
```

```
kplot(object, xax = 1, yax = 2, which.tab = 1:length(object$blo),
      mfrow = NULL, permute.row.col = FALSE, clab.row = 1,
      clab.col = 1.25, csub = 2, possub = "bottomright",
      show.eigen.value = TRUE, poseig = c("bottom", "top"), ...)
```

Arguments

<code>object</code>	an object of class <code>sepan</code>
<code>xax</code> , <code>yax</code>	the numbers of the x-axis and the y-axis
<code>which.tab</code>	a numeric vector containing the numbers of the tables to analyse
<code>mfrow</code>	parameter for the array of figures to be drawn, otherwise use <code>n2mfrow</code>
<code>permute.row.col</code>	if TRUE the rows are represented by arrows and the columns by points, if FALSE it is the opposite
<code>clab.row</code>	a character size for the row labels
<code>clab.col</code>	a character size for the column labels
<code>traject.row</code>	a logical value indicating whether the trajectories between rows should be drawn in a natural order
<code>csub</code>	a character size for the sub-titles, used with <code>par("cex")*csub</code>
<code>possub</code>	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
<code>show.eigen.value</code>	a logical value indicating whether the eigenvalues bar plot should be drawn
<code>poseig</code>	if "top" the eigenvalues bar plot is upside, if "bottom", it is downside
<code>...</code>	further arguments passed to or from other methods

Details

`kplot.sepan` superimposes the points for the rows and the arrows for the columns using an adapted rescaling such as the `scatter.dudi`.

`kplot.sepan.coa` superimposes the row coordinates and the column coordinates with the same scale.

Author(s)

Daniel Chessel

Examples

```
data(escopage)
w <- data.frame(scale(escopage$tab))
w <- ktab.data.frame(w, escopage$blo, tabnames = escopage$tab.names)
sep1 <- sepan(w)
kplot(sep1, show = FALSE)

data(friday87)
```

```

w <- data.frame(scale(friday87$fau, scal = FALSE))
w <- ktab.data.frame(w, friday87$fau.blo, tabnames = friday87$tab.names)
kplot(sepan(w), clab.r = 1.25, clab.c = 0, csub = 3)

data(microsatt)
w <- dudi.coa(data.frame(t(microsatt$tab)), scann = FALSE)
loci.fac <- factor(rep(microsatt$loci.names, microsatt$loci.eff))
wit <- wca(w, loci.fac, scann = FALSE)
microsatt.ktab <- ktab.within(wit)
kplot.sepan.coa(sepan(microsatt.ktab), show = FALSE, clab.c = 0,
mfrow = c(3,3), clab.r = 1.5)

```

kplot.statis

Multiple Graphs of a STATIS Analysis

Description

performs high level plots for a STATIS analysis, using an object of class `statis`.

Usage

```

## S3 method for class 'statis'
kplot(object, xax = 1, yax = 2, mfrow = NULL,
      which.tab = 1:length(object$tab.names), clab = 1.5, cpoi = 2,
      traject = FALSE, arrow = TRUE, class = NULL,
      unique.scale = FALSE, csub = 2, possub = "bottomright",...)

```

Arguments

<code>object</code>	an object of class <code>statis</code>
<code>xax, yax</code>	the numbers of the x-axis and the y-axis
<code>mfrow</code>	parameter for the array of figures to be drawn
<code>which.tab</code>	a numeric vector containing the numbers of the tables to analyse
<code>clab</code>	a character size for the labels
<code>cpoi</code>	the size of points
<code>traject</code>	a logical value indicating whether the trajectories should be drawn in a natural order
<code>arrow</code>	a logical value indicating whether the column factorial diagrams should be plotted
<code>class</code>	if not <code>NULL</code> , a factor of length equal to the number of the total columns of the K-tables
<code>unique.scale</code>	if <code>TRUE</code> , all the arrays of figures have the same scale
<code>csub</code>	a character size for the labels of the arrays of figures used with <code>par("cex")*csub</code>
<code>possub</code>	a string of characters indicating the sub-title position (" <code>opleft</code> ", " <code>oprigh</code> ", " <code>ottomleft</code> ", " <code>ottomright</code> ")
<code>...</code>	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```

data(jv73)
dudi1 <- dudi.pca(jv73$poi, scann = FALSE, scal = FALSE)
wit1 <- wca(dudi1, jv73$fac.riv, scann = FALSE)
kta3 <- ktab.within(wit1)
data(jv73)
statis3 <- statis(kta3, scann = FALSE)
kplot(statis3, traj = TRUE, arrow = FALSE, unique = TRUE,
      clab = 0, csub = 3, cpoi = 3)

```

krandtest*Class of the Permutation Tests (in C).*

Description

Plot and print many permutation tests. Objects of class 'krandtest' are lists.

Usage

```

## S3 method for class 'krandtest'
plot(x, mfrow = NULL, nclass = NULL, main.title = x$names, ...)
## S3 method for class 'krandtest'
print(x, ...)
as.krandtest(sim, obs, alter = "greater", call =
match.call(), names = colnames(sim), p.adjust.method = "none")

```

Arguments

x	: an object of class 'krandtest'
mfrow	: a vector of the form 'c(nr,nc)', otherwise computed by as special own function n2mfrow
nclass	: a number of intervals for the histogram
main.title	: a string of character for the main title
...	: further arguments passed to or from other methods
sim	a matrix or data.frame of simulated values (repetitions as rows, number of tests as columns)
obs	a numeric vector of observed values for each test
alter	a vector of character specifying the alternative hypothesis for each test. Each element must be one of "greater" (default), "less" or "two-sided". The length must be equal to the length of the vector obs, values are recycled if shorter.
call	a call order

names a vector of names for tests
 p.adjust.method a string indicating a method for multiple adjustment, see p.adjust.methods for possible choices.

Value

plot.krandtest draws the p simulated values histograms and the position of the observed value.

Author(s)

Daniel Chessel and Stephane Dray <dray@biomserv.univ-lyon1.fr>

See Also

[randtest](#)

Examples

```
wkrandtest <- as.krandtest(obs=c(0,1.2,2.4,3.4,5.4,20.4),sim=matrix(rnorm(6*200),200,6))
wkrandtest
plot(wkrandtest)
```

ktab *the class of objects 'ktab' (K-tables)*

Description

an object of class ktab is a list of data frames with the same row.names in common.
 a list of class 'ktab' contains moreover :

blo : the vector of the numbers of columns for each table

lw : the vector of the row weightings in common for all tables

cw : the vector of the column weightings

TL : a data frame of two components to manage the parameter positions associated with the rows of tables

TC : a data frame of two components to manage the parameter positions associated with the columns of tables

T4 : a data frame of two components to manage the parameter positions of 4 components associated to an array

Usage

```
## S3 method for class 'ktab'
c(...)
## S3 method for class 'ktab'
x[selection]
is.ktab(x)
## S3 method for class 'ktab'
t(x)
## S3 method for class 'ktab'
row.names(x)
## S3 method for class 'ktab'
col.names(x)
tab.names(x)
col.names(x)
ktab.util.names(x)
```

Arguments

x	an object of the class ktab
...	a sequence of objects of the class ktab
selection	an integer vector

Details

A 'ktab' object can be created with :

- a list of data frame : [ktab.list.df](#)
- a list of dudi objects : [ktab.list.dudi](#)
- a data.frame : [ktab.data.frame](#)
- an object within : [ktab.within](#)
- a couple of ktabs : [ktab.match2ktabs](#)

Value

`c.ktab` returns an object ktab. It concatenates K-tables with the same rows in common.

`t.ktab` returns an object ktab. It permutes each data frame into a K-tables. All tables have the same column names and the same column weightings (a data cube).

`"["` returns an object ktab. It allows to select some arrays in a K-tables.

`is.ktab` returns TRUE if x is a K-tables.

`row.names` returns the vector of the row names common with all the tables of a K-tables and allows to modify them.

`col.names` returns the vector of the column names of a K-tables and allows to modify them.

`tab.names` returns the vector of the array names of a K-tables and allows to modify them.

`ktab.util.names` is a useful function.

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```

data(friday87)
wfri <- data.frame(scale(friday87$fau, scal = FALSE))
wfri <- ktab.data.frame(wfri, friday87$fau.blo)
wfri[2:4]
c(wfri[2:4], wfri[5])

data(meaudret)
wit1 <- withinpca(meaudret$env, meaudret$design$season, scan = FALSE,
  scal = "partial")
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5"), 4))
kta2 <- t(kta1)
kplot(sepan(kta2), clab.r = 1.5, clab.c = 0.75)

```

ktab.data.frame	<i>Creation of K-tables from a data frame</i>
-----------------	---

Description

creates K tables from a data frame.

Usage

```

ktab.data.frame(df, blocks, rownames = NULL, colnames = NULL,
  tabnames = NULL, w.row = rep(1, nrow(df)) / nrow(df),
  w.col = rep(1, ncol(df)))

```

Arguments

df	a data frame
blocks	an integer vector for which the sum must be the number of variables of df. Its length is the number of arrays of the K-tables
rownames	the row names of the K-tables (otherwise the row names of df)
colnames	the column names of the K-tables (otherwise the column names of df)
tabnames	the names of the arrays of the K-tables (otherwise "Ana1", "Ana2", ...)
w.row	a vector of the row weightings
w.col	a vector of the column weightings

Value

returns a list of class ktab. See [ktab](#).

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```

data(escopage)
wescopage <- data.frame(scalewt(escopage$tab))
wescopage <- ktab.data.frame(wescopage, escopage$blo,
                             tabnames = escopage$tab.names)
plot(sepan(wescopage))
data(friday87)
w <- data.frame(scale(friday87$fau, scal = FALSE))
w <- ktab.data.frame(w, friday87$fau.blo, tabnames = friday87$tab.names)
kplot(sepan(w))

```

ktab.list.df

Creating a K-tables from a list of data frames.

Description

creates a list of class ktab from a list of data frames

Usage

```

ktab.list.df(obj, rownames = NULL, colnames = NULL, tabnames = NULL,
             w.row = rep(1, nrow(obj[[1]])), w.col = lapply(obj, function(x)
             rep(1 / ncol(x), ncol(x))))

```

Arguments

obj	a list of data frame
rownames	the names of the K-tables rows (otherwise, the row names of the arrays)
colnames	the names of the K-tables columns (otherwise, the column names of the arrays)
tabnames	the names of the arrays of the K-tables (otherwise, the names of the obj if they exist, or else "Ana1", "Ana2", ...)
w.row	a vector of the row weightings in common with all the arrays
w.col	a list of the vector of the column weightings for each array

Details

Each element of the initial list have to possess the same names and row numbers

Value

returns a list of class ktab. See [ktab](#)

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```

data(jv73)
l0 <- split(jv73$morpho, jv73$fac.riv)
l0 <- lapply(l0, function(x) data.frame(t(scalewt(x))))
kta <- ktab.list.df(l0)
kplot(sepan(kta[c(2,5,7,10)]), perm = TRUE, clab.r = 1, clab.c = 1.5)

```

ktab.list.dudi	<i>Creation of a K-tables from a list of duality diagrams</i>
----------------	---

Description

creates a list of class ktab from a list of duality diagrams.

Usage

```
ktab.list.dudi(obj, rownames = NULL, colnames = NULL, tabnames = NULL)
```

Arguments

obj	a list of objects of class 'dudi'. Each element of the list must have the same row names for \$tab and even for \$lw
rownames	the row names of the K-tables (otherwise the row names of the \$tab)
colnames	the column names of the K-tables (otherwise the column names of the \$tab)
tabnames	the names of the arrays of the K-tables (otherwise the names of the obj if they exist, or else "Ana1", "Ana2", ...)

Value

returns a list of class ktab. See [ktab](#)

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```

data(euro123)
pca1 <- dudi.pca(euro123$in78, scale = FALSE, scann = FALSE)
pca2 <- dudi.pca(euro123$in86, scale = FALSE, scann = FALSE)
pca3 <- dudi.pca(euro123$in97, scale = FALSE, scann = FALSE)
ktabeuro <- ktab.list.dudi(list(pca1, pca2, pca3),
  tabnames = c("1978", "1986", "1997"))
kplot(sepan(ktabeuro), mfr = c(2,2), clab.c = 1.5)

data(meaudret)
w1 <- split(meaudret$env, meaudret$design$season)

```

```

ll <- lapply(w1, dudi.pca, scann = FALSE)
kta <- ktab.list.dudi(ll, rownames <- paste("Site", 1:5, sep=""))
kplot(sepan(kta), clab.r = 1.5, clab.c = 0.75)

data(jv73)
w <- split(jv73$poi, jv73$fac.riv)
wlv73poi <- lapply(w, dudi.pca, scal = FALSE, scan = FALSE)
wlv73poi <- lapply(wlv73poi, t)
wlv73poi <- ktab.list.dudi(wlv73poi)
kplot(sepan(wlv73poi), permut = TRUE, traj = TRUE)

```

ktab.match2ktabs	<i>STATIS and Co-Inertia : Analysis of a series of paired ecological tables</i>
------------------	---

Description

Prepares the analysis of a series of paired ecological tables. Partial Triadic Analysis (see [pta](#)) can be used thereafter to perform the analysis of this k-table.

Usage

```
ktab.match2ktabs(KTX, KTY)
```

Arguments

KTX	an objet of class ktab
KTY	an objet of class ktab

Value

a list of class ktab, subclass kcoinertia. See [ktab](#)

WARNING

IMPORTANT : KTX and KTY must have the same k-tables structure, the same number of columns, and the same column weights.

Author(s)

Jean Thioulouse <Jean.Thioulouse@univ-lyon1.fr>

References

Thioulouse J., Simier M. and Chessel D. (2004). Simultaneous analysis of a sequence of paired ecological tables. *Ecology* **85**, 272-283..

Simier, M., Blanc L., Pellegrin F., and Nandris D. (1999). Approche simultanée de K couples de tableaux : Application a l'étude des relations pathologie végétale - environnement. *Revue de Statistique Appliquée*, **47**, 31-46.

Examples

```

data(meau)
wit1 <- withinpca(meau$env, meau$design$season, scan = FALSE, scal = "total")
pcaspe <- dudi.pca(meau$spe, scale = FALSE, scan = FALSE, nf = 2)
wit2 <- wca(pcaspe, meau$design$season, scan = FALSE, nf = 2)
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5", "S6"), 4))
kta2 <- ktab.within(wit2, colnames = rep(c("S1", "S2", "S3", "S4", "S5", "S6"), 4))
kcoi <- ktab.match2ktabs(kta1, kta2)
ptacoi <- pta(kcoi, scan = FALSE, nf = 2)
plot(ptacoi)
kplot(ptacoi)

```

ktab.within

Process to go from a Within Analysis to a K-tables

Description

performs the process to go from a Within Analysis to a K-tables.

Usage

```

ktab.within(dudiwit, rownames = NULL, colnames = NULL, tabnames = NULL)

```

Arguments

dudiwit	an objet of class within
rownames	the row names of the K-tables (otherwise the row names of dudiwit\$tab)
colnames	the column names of the K-tables (otherwise the column names of dudiwit\$tab)
tabnames	the names of the arrays of the K-tables (otherwise the levels of the factor which defines the within-classes)

Value

a list of class ktab. See [ktab](#)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```

data(bacteria)
w1 <- data.frame(t(bacteria$espcodon))
dudi1 <- dudi.coa(w1, scann = FALSE, nf = 4)
wit1 <- wca(dudi1, bacteria$code, scannf = FALSE)
kta1 <- ktab.within(wit1)
plot(statis(kta1, scann = FALSE))

kta2 <- kta1[kta1$blo>3]
kplot(mfa(kta2, scann = FALSE))

```

 lascaux

Genetic/Environment and types of variables

Description

This data set gives meristic, genetic and morphological data frame for 306 trouts.

Usage

```
data(lascaux)
```

Format

lascaux is a list of 9 components.

riv is a factor returning the river where 306 trouts are captured

code vector of characters : code of the 306 trouts

sex factor sex of the 306 trouts

meris data frame 306 trouts - 5 meristic variables

tap data frame of the total number of red and black points

gen factor of the genetic code of the 306 trouts

morpho data frame 306 trouts 37 morphological variables

colo data frame 306 trouts 15 variables of coloring

ornem data frame 306 trouts 15 factors (ornementation)

Source

Lascaux, J.M. (1996) *Analyse de la variabilité morphologique de la truite commune (Salmo trutta L.) dans les cours d'eau du bassin pyrénéen méditerranéen*. Thèse de doctorat en sciences agronomiques, INP Toulouse.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps022.pdf> (in French).

Examples

```
data(lascaux)
par(mfrow = c(2,2))
barplot(dudi.pca(lascaux$meris, scan = FALSE)$eig)
title(main = "Meristic")
barplot(dudi.pca(lascaux$colo, scan = FALSE)$eig)
title(main = "Coloration")
barplot(dudi.pca(na.omit(lascaux$morpho), scan = FALSE)$eig)
title(main = "Morphometric")
barplot(dudi.acm(na.omit(lascaux$orne), scan = FALSE)$eig)
title(main = "Ornemental")
par(mfrow = c(1,1))
```

lingoes

Transformation of a Distance Matrix for becoming Euclidean

Description

transforms a distance matrix in a Euclidean one.

Usage

```
lingoes(distmat, print = FALSE, tol = 1e-07, cor.zero = TRUE)
```

Arguments

distmat	an object of class dist
print	if TRUE, prints the eigenvalues of the matrix
tol	a tolerance threshold for zero
cor.zero	if TRUE, zero distances are not modified

Details

The function uses the smaller positive constant k which transforms the matrix of $\sqrt{d_{ij}^2 + 2 * k}$ in an Euclidean one

Value

returns an object of class dist with a Euclidean distance

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

Lingoes, J.C. (1971) Some boundary conditions for a monotone analysis of symmetric matrices. *Psychometrika*, **36**, 195–203.

Examples

```
data(capitales)
d0 <- capitales$dist
is.euclid(d0) # FALSE
d1 <- lingoes(d0, TRUE)
# Lingoes constant = 2120982
is.euclid(d1) # TRUE
plot(d0, d1)
x0 <- sort(unclass(d0))
lines(x0, sqrt(x0^2 + 2 * 2120982), lwd = 3)

is.euclid(sqrt(d0^2 + 2 * 2120981), tol = 1e-10) # FALSE
is.euclid(sqrt(d0^2 + 2 * 2120982), tol = 1e-10) # FALSE
is.euclid(sqrt(d0^2 + 2 * 2120983), tol = 1e-10)
# TRUE the smaller constant
```

lizards

Phylogeny and quantitative traits of lizards

Description

This data set describes the phylogeny of 18 lizards as reported by Bauwens and D'iaz-Uriarte (1997). It also gives life-history traits corresponding to these 18 species.

Usage

```
data(lizards)
```

Format

lizards is a list containing the 3 following objects :

traits is a data frame with 18 species and 8 traits.

hprA is a character string giving the phylogenetic tree (hypothesized phylogenetic relationships based on immunological distances) in Newick format.

hprB is a character string giving the phylogenetic tree (hypothesized phylogenetic relationships based on morphological characteristics) in Newick format.

Details

Variables of `lizards$traits` are the following ones : mean.L (mean length (mm)), matur.L (length at maturity (mm)), max.L (maximum length (mm)), hatch.L (hatchling length (mm)), hatch.m (hatchling mass (g)), clutch.S (Clutch size), age.mat (age at maturity (number of months of activity)), clutch.F (clutch frequency).

References

Bauwens, D., and D'iaz-Uriarte, R. (1997) Covariation of life-history traits in lacertid lizards: a comparative study. *American Naturalist*, **149**, 91–111.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps063.pdf> (in French).

Examples

```
data(lizards)
w <- data.frame(scalewt(log(lizards$traits)))
par(mfrow = c(1,2))
wphy <- newick2phylog(lizards$hprA)
table.phylog(w, wphy, csi = 3)
wphy <- newick2phylog(lizards$hprB)
table.phylog(w, wphy, csi = 3)
par(mfrow = c(1,1))
```

macaca

Landmarks

Description

This data set gives the landmarks of a macaca at the ages of 0.9 and 5.77 years.

Usage

```
data(macaca)
```

Format

macaca is a list of 2 components.

xy1 is a data frame with 72 points and 2 coordinates.

xy2 is a data frame with 72 points and 2 coordinates.

Source

Olshan, A.F., Siegel, A.F. and Swindler, D.R. (1982) Robust and least-squares orthogonal mapping: Methods for the study of cephalofacial form and growth. *American Journal of Physical Anthropology*, **59**, 131–137.

Examples

```
data(macaca)
par(mfrow = c(2,2))
s.match(macaca$xy1, macaca$xy2, clab = 0)
pro1 <- procuste(macaca$xy1, macaca$xy2, scal = FALSE)
s.match(pro1$stab1, pro1$rot2, clab = 0.7)
s.match(pro1$stab2, pro1$rot1, clab = 0.7)
```

```
pro2 <- procuste(macaca$xy1, macaca$xy2)
s.match(pro2$tab2, pro2$rot1, clab = 0.7)
par(mfrow = c(1,1))
```

macon

Wine Tasting

Description

The macon data frame has 8 rows-wines and 25 columns-tasters. Each column is a classification of 8 wines (Beaujolais, France).

Usage

```
data(macon)
```

Source

Foire Nationale des Vins de France, Mâcon, 1985

Examples

```
data(macon)
s.corcircle(dudi.pca(macon, scan = FALSE)$co)
```

macroloire

Assemblages of Macroinvertebrates in the Loire River (France)

Description

A total of 38 sites were surveyed along 800 km of the Loire River yielding 40 species of Trichoptera and Coleoptera sampled from riffle habitats. The river was divided into three regions according to geology: granitic highlands (Region#1), limestone lowlands (Region#2) and granitic lowlands (Region#3). This data set has been collected for analyzing changes in macroinvertebrate assemblages along the course of a large river. Four criterias are given here: variation in 1/ species composition and relative abundance, 2/ taxonomic composition, 3/ Body Sizes, 4/ Feeding habits.

Usage

```
data(macroloire)
```

Format

`macroloire` is a list of 5 components.

fau is a data frame containing the abundance of each species in each station.

traits is a data frame describes two traits : the maximal sizes and feeding habits for each species. Each trait is divided into categories. The maximal size achieved by the species is divided into four length categories: ≤ 5 mm ; $>5-10$ mm ; $>10-20$ mm ; $>20-40$ mm. Feeding habits comprise seven categories: engulfers, shredders, scrapers, deposit-feeders, active filter-feeders, passive filter-feeders and piercers, in this order. The affinity of each species to each trait category is quantified using a fuzzy coding approach. A score is assigned to each species for describing its affinity for a given trait category from "0" which indicates no affinity to "3" which indicates high affinity. These affinities are further transformed into percentage per trait per species.

taxo is a data frame with species and 3 factors: Genus, Family and Order. It is a data frame of class "taxo": the variables are factors giving nested classifications.

envir is a data frame giving for each station, its name (variable "SamplingSite"), its distance from the source (km, variable "Distance"), its altitude (m, variable "Altitude"), its position regarding the dams [1: before the first dam; 2: after the first dam; 3: after the second dam] (variable "Dam"), its position in one of the three regions defined according to geology: granitic highlands, limestone lowlands and granitic lowlands (variable "Morphoregion"), presence of confluence (variable "Confluence")

labels is a data frame containing the latin names of the species.

Source

Ivol, J.M., Guinand, B., Richoux, P. and Tachet, H. (1997) Longitudinal changes in Trichoptera and Coleoptera assemblages and environmental conditions in the Loire River (France). *Archiv für Hydrobiologie*, **138**, 525–557.

Pavoine S. and Dolédec S. (2005) The apportionment of quadratic entropy: a useful alternative for partitioning diversity in ecological data. *Environmental and Ecological Statistics*, **12**, 125–138.

Examples

```
data(macroloire)
apqe.Equi <- apqe(macroloire$fau, , macroloire$morphoregions)
apqe.Equi
#test.Equi <- randtest.apqe(apqe.Equi, method = "aggregated", 99)
#plot(test.Equi)

## Not run:

m.phy <- taxo2phylog(macroloire$taxo)
apqe.Tax <- apqe(macroloire$fau, m.phy$Wdist, macroloire$morphoregions)
apqe.Tax
#test.Tax <- randtest.apqe(apqe.Tax, method = "aggregated", 99)
#plot(test.Tax)
```

```

dSize <- sqrt(dist.prop(macroloire$traits[ ,1:4], method = 2))
apqe.Size <- apqe(macroloire$fau, dSize, macroloire$morphoregions)
apqe.Size
#test.Size <- randtest.apqe(apqe.Size, method = "aggregated", 99)
#plot(test.Size)

dFeed <- sqrt(dist.prop(macroloire$traits[ ,-(1:4)], method = 2))
apqe.Feed <- apqe(macroloire$fau, dFeed, macroloire$morphoregions)
apqe.Feed
#test.Feed <- randtest.apqe(apqe.Feed, method = "aggregated", 99)
#plot(test.Size)

## End(Not run)

```

mafragh

Phyto-Ecological Survey

Description

This data set gives environmental and spatial informations about species and sites.

Usage

```
data(mafragh)
```

Format

mafragh is a list of 6 components.

xy are the coordinates of 97 sites.

flo is a data frame with 97 sites and 56 species.

espnames is a vector of the names of species.

neig is the neighbourhood graph of the 97 sites (an object of class 'neig').

mil is a data frame with 97 sites and 11 environmental variables.

partition is a factor classifying the 97 sites in 5 classes.

area is a data frame of class area

Source

Belair, G.d. and Bencheikh-Lehocine, M. (1987) Composition et déterminisme de la végétation d'une plaine côtière marécageuse : La Mafragh (Annaba, Algérie). *Bulletin d'Ecologie*, **18**, 393–407.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps053.pdf> (in French).

Examples

```

data(mafragh)
par(mfrow = c(3,2))
s.label(mafragh$xy, inc = FALSE, neig = mafragh$neig,
        sub = "Samples & Neighbourhood graph")
coa1 <- dudi.coa(mafragh$flo, scan = FALSE)
s.value(mafragh$xy, coa1$li[,1], sub = "Axis 1 - COA")
pca1 <- dudi.pca(mafragh$xy, scan = FALSE)
s.value(mafragh$xy, pca1$li[,1], sub = "Axis 1 - PCA")
s.class(pca1$li, mafragh$partition, sub = "Plane 1-2 - PCA")
s.class(coa1$li, mafragh$partition, sub = "Plane 1-2 - COA")
s.chull(mafragh$xy, mafragh$partition, optchull = 1)
par(mfrow=c(1,1))

## Not run:
link1 <- area2link(mafragh$area)
neig1 <- neig(mat01 = 1*(link1>0))
nb1 <- neig2nb(neig1)
par(mfrow = c(2,1))
area.plot(mafragh$area,center = mafragh$xy,clab=0.75)
area.plot(mafragh$area,center = mafragh$xy,graph=neig1)
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  lw1 <- apply(link1,1,function(x) x[x>0])
  listw1 <- nb2listw(nb1,lw1)
  coa1 <- dudi.coa(mafragh$flo, scan = FALSE, nf = 4)
  ms1 <- multispati(coa1, listw1, scan = FALSE, nfp = 2, nfn = 0)
  summary(ms1)
  par(mfrow = c(2,2))
  barplot(coa1$eig)
  barplot(ms1$eig)
  s.corcircle(ms1$as)
  plot(coa1$li[,1], ms1$li[,1])
}
par(mfrow = c(1,1))

## End(Not run)

```

mantel.randtest

Mantel test (correlation between two distance matrices (in C).)

Description

Performs a Mantel test between two distance matrices.

Usage

```
mantel.randtest(m1, m2, nrepet = 999)
```

Arguments

m1 an object of class dist
m2 an object of class dist
nrepet the number of permutations

Value

an object of class randtest (randomization tests)

Author(s)

Jean Thioulouse <ade4-jt@biomserv.univ-lyon1.fr>

References

Mantel, N. (1967) The detection of disease clustering and a generalized regression approach. *Cancer Research*, **27**, 209–220.

Examples

```
data(yanomama)
gen <- quasieucld(as.dist(yanomama$gen))
geo <- quasieucld(as.dist(yanomama$geo))
plot(r1 <- mantel.randtest(geo,gen), main = "Mantel's test")
r1
```

`mantel.rtest`

Mantel test (correlation between two distance matrices (in R).)

Description

Performs a Mantel test between two distance matrices.

Usage

```
mantel.rtest(m1, m2, nrepet = 99)
```

Arguments

m1 an object of class dist
m2 an object of class dist
nrepet the number of permutations

Value

an object of class rtest (randomization tests)

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

References

Mantel, N. (1967) The detection of disease clustering and a generalized regression approach. *Cancer Research*, **27**, 209–220.

Examples

```
data(yanomama)
gen <- quasieulid(as.dist(yanomama$gen))
geo <- quasieulid(as.dist(yanomama$geo))
plot(r1 <- mantel.rtest(geo,gen), main = "Mantel's test")
r1
```

maples

Phylogeny and quantitative traits of flowers

Description

This data set describes the phylogeny of 17 flowers as reported by Ackerly and Donoghue (1998). It also gives 31 traits corresponding to these 17 species.

Usage

```
data(maples)
```

Format

`ti` `thonia` is a list containing the 2 following objects :

tre is a character string giving the phylogenetic tree in Newick format.

tab is a data frame with 17 species and 31 traits

Source

Data were obtained from the URL <http://www.stanford.edu/~dackerly/acerdata.html>.

References

Ackerly, D. D. and Donoghue, M.J. (1998) Leaf size, sapling allometry, and Corner's rules: phylogeny and correlated evolution in Maples (Acer). *American Naturalist*, **152**, 767–791.

Examples

```

data(maples)
phy <- newick2phylog(maples$tre)
dom <- maples$tab$Dom
bif <- maples$tab$Bif
orthogram(dom, phylog = phy)
orthogram(bif, phylog = phy)
par(mfrow = c(1,2))
dotchart.phylog(phy, dom)
dotchart.phylog(phy, bif, clabel.nodes = 0.7)
par(mfrow = c(1,1))
plot(bif,dom,pch = 20)
abline(lm(dom~bif))
summary(lm(dom~bif))
if (require(ape, quiet = TRUE)){
cor.test(bif,dom)
phylo <- read.tree(text = maples$tre)
pic.bif <- pic(bif, phylo)
pic.dom <- pic(dom, phylo)
cor.test(pic.bif, pic.dom)}

```

mariages

Correspondence Analysis Table

Description

This array contains the socio-professionnal repartitions of 5850 couples.

Usage

```
data(mariages)
```

Format

The `mariages` data frame has 9 rows and 9 columns. The rows represent the wife's socio-professionnal category and the columns the husband's socio-professionnal category (1982).

Codes for rows and columns are identical : `agri` (Farmers), `ouva` (Farm workers), `pat` (Company directors (commerce and industry)), `sup` (Liberal profession, executives and higher intellectual professions), `moy` (Intermediate professions), `emp` (Other white-collar workers), `ouv` (Manual workers), `serv` (Domestic staff), `aut` (other workers).

Source

Vallet, L.A. (1986) Activité professionnelle de la femme mariée et détermination de la position sociale de la famille. Un test empirique : la France entre 1962 et 1982. *Revue Française de Sociologie*, **27**, 656–696.

Examples

```

data(mariages)
par(mfrow = c(2,2))
w <- dudi.coa(mariages, scan = FALSE, nf = 3)
scatter(w, met = 1, posi = "bottom")
scatter(w, met = 2, posi = "bottom")
scatter(w, met = 3, posi = "bottom")
score(w, 3)
par(mfrow = c(1,1))

```

mcoa

Multiple CO-inertia Analysis

Description

performs a multiple CO-inertia analysis, using an object of class `ktab`.

Usage

```

mcoa(X, option = c("inertia", "lambda1", "uniform", "internal"),
     scannf = TRUE, nf = 3, tol = 1e-07)
## S3 method for class 'mcoa'
print(x, ...)
## S3 method for class 'mcoa'
summary(object, ...)
## S3 method for class 'mcoa'
plot(x, xax = 1, yax = 2, eig.bottom = TRUE, ...)

```

Arguments

<code>X</code>	an object of class <code>ktab</code>
<code>option</code>	a string of characters for the weightings of the arrays options : "inertia" weighting of group <code>k</code> by the inverse of the total inertia of the array <code>k</code> "lambda1" weighting of group <code>k</code> by the inverse of the first eigenvalue of the <code>k</code> analysis "uniform" uniform weighting of groups "internal" weighting included in <code>X\$tabw</code>
<code>scannf</code>	a logical value indicating whether the eigenvalues bar plot should be displayed
<code>nf</code>	if <code>scannf</code> <code>FALSE</code> , an integer indicating the number of kept axes
<code>tol</code>	a tolerance threshold, an eigenvalue is considered positive if it is larger than $-tol \times \text{lambda1}$ where <code>lambda1</code> is the largest eigenvalue.
<code>x, object</code>	an object of class <code>'mcoa'</code>
<code>...</code>	further arguments passed to or from other methods
<code>xax, yax</code>	the numbers of the x-axis and the y-axis
<code>eig.bottom</code>	a logical value indicating whether the eigenvalues bar plot should be added

Value

mcoa returns a list of class 'mcoa' containing :

pseudoeig	a numeric vector with the all pseudo eigenvalues
call	the call-up order
nf	a numeric value indicating the number of kept axes
SynVar	a data frame with the synthetic scores
axis	a data frame with the co-inertia axes
Tli	a data frame with the co-inertia coordinates
Tl1	a data frame with the co-inertia normed scores
Tax	a data frame with the inertia axes onto co-inertia axis
Tco	a data frame with the column coordinates onto synthetic scores
TL	a data frame with the factors for Tli Tl1
TC	a data frame with the factors for Tco
T4	a data frame with the factors for Tax
lambda	a data frame with the all eigenvalues (computed on the separate analyses)
cov2	a numeric vector with the all pseudo eigenvalues (synthetic analysis)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Chessel, D. and Hanafi, M. (1996) Analyses de la co-inertie de K nuages de points, *Revue de Statistique Appliquée*, **44**, 35–60.

Examples

```
data(friday87)
w1 <- data.frame(scale(friday87$fau, scal = FALSE))
w2 <- ktab.data.frame(w1, friday87$fau.blo, tabnames = friday87$tab.names)
mcoa1 <- mcoa(w2, "lambda1", scan = FALSE)
mcoa1
summary(mcoa1)
plot(mcoa1)
```

Description

The DPCoA analysis (see [dpcoa](#)) has been developed by Pavoine et al. (2004). It has been used in genetics for describing inter-population nucleotide diversity. However, this procedure can only be used with one locus. In order to measure and describe nucleotide diversity with more than one locus, we developed three versions of multiple DPCoA by using three ordination methods: multiple co-inertia analysis, STATIS, and multiple factorial analysis. The multiple DPCoA allows the impact of various loci in the measurement and description of diversity to be quantified and described. This method is general enough to handle a large variety of data sets. It complements existing methods such as the analysis of molecular variance or other analyses based on linkage disequilibrium measures, and is very useful to study the impact of various loci on the measurement of diversity.

Usage

```
mdpcoa(msamples, mdistances = NULL, method =
  c("mcoa", "statis", "mfa"),
  option = c("inertia", "lambda1", "uniform", "internal"),
  scannf = TRUE, nf = 3, full = TRUE,
  nfsep = NULL, tol = 1e-07)
kplotX.mdpcoa(object, xax = 1, yax = 2, mfrow = NULL,
  which.tab = 1:length(object$X), includepop = FALSE,
  clab = 0.7, cpoi = 0.7, unique.scale = FALSE,
  csub = 2, possub = "bottomright")
prep.mdpcoa(dnaobj, pop, model, ...)
```

Arguments

<code>msamples</code>	A list of data frames with the populations as columns, alleles as rows and abundances as entries. All the tables should have equal numbers of columns (populations). Each table corresponds to a locus;
<code>mdistances</code>	A list of objects of class 'dist', corresponding to the distances among alleles. The order of the loci should be the same in <code>msamples</code> as in <code>mdistances</code> ;
<code>method</code>	One of the three possibilities: "mcoa", "statis", or "mfa". If a vector is given, only its first value is considered;
<code>option</code>	One of the four possibilities for normalizing the population coordinates over the loci: "inertia", "lambda1", "uniform", or "internal". These options are used with MCoA and MFA only;
<code>scannf</code>	a logical value indicating whether the eigenvalues bar plots should be displayed;
<code>nf</code>	if <code>scannf</code> is FALSE, an integer indicating the number of kept axes for the multiple analysis;
<code>full</code>	a logical value indicating whether all the axes should be kept in the separated analyses (one analysis, DPCoA, per locus);

nfsep	if full is FALSE, a vector indicating the number of kept axes for each of the separated analyses;
tol	a tolerance threshold for null eigenvalues (a value less than tol times the first one is considered as null);
object	an object of class 'mdpcoa';
xax	the number of the x-axis;
yax	the number of the y-axis;
mfrow	a vector of the form 'c(nr,nc)', otherwise computed by as special own function 'n2mfrow';
which.tab	a numeric vector containing the numbers of the loci to analyse;
includepop	a logical indicating if the populations must be displayed. In that case, the alleles are displayed by points and the populations by labels;
clab	a character size for the labels;
cpoi	a character size for plotting the points, used with 'par("cex")*cpoi'. If zero, no points are drawn;
unique.scale	if TRUE, all the arrays of figures have the same scale;
csub	a character size for the labels of the arrays of figures used with 'par("cex")*csub';
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright");
dnaobj	a list of dna sequences that can be obtained with the function read.dna of the ape package;
pop	a factor that gives the name of the population to which each sequence belongs;
model	a vector giving the model to be applied for the calculations of the distances for each locus. One model should be attributed to each locus, given that the loci are in alphabetical order. The models can take the following values: "raw", "JC69", "K80" (the default), "F81", "K81", "F84", "BH87", "T92", "TN93", "GG95", "logdet", or "paralin". See the help documentation for the function "dist.dna" of ape for a description of the models.
...	... further arguments passed to or from other methods

Details

An object obtained by the function `mdpcoa` has two classes. The first one is "mdpcoa" and the second is either "mcoa", or "statis", or "mfa", depending on the method chosen. Consequently, other functions already available in `ade4` for displaying graphical results can be used: With `MCoA`, - `plot.mcoa`: this function displays (1) the differences among the populations according to each locus and the compromise, (2) the projection of the principal axes of the individual analyses onto the synthetic variables, (3) the projection of the principal axes of the individual analyses onto the co-inertia axes, (4) the squared vectorial covariance among the co-inertia scores and the synthetic variables; - `kplot.mcoa`: this function divides previous displays (figures 1, 2, or 3 described in `plot.mcoa`) by giving one plot per locus.

With `STATIS`, - `plot.statis`: this function displays (1) the scores of each locus according to the two first eigenvectors of the matrix Rv , (2) the scatter diagram of the differences among populations

according to the compromise, (3) the weight attributed to each locus in abscissa and the vectorial covariance among each individual analysis with the notations in the main text of the paper) and the compromise analysis in ordinates, (4) the covariance between the principal component inertia axes of each locus and the axes of the compromise space; - `kplot.statis`: this function displays for each locus the projection of the principal axes onto the compromise space.

With MFA, - `plot.mfa`: this function displays (1) the differences among the populations according to each locus and the compromise, (2) the projection of the principal axes of the individual analyses onto the compromise, (3) the covariance between the principal component inertia axes of each locus and the axes of the compromise space, (4) for each axis of the compromise, the amount of inertia conserved by the projection of the individual analyses onto the common space. - `kplot.mfa`: this function displays for each locus the projection of the principal axes and populations onto the compromise space.

Value

The functions provide the following results:

`dist.ktab` returns an object of class `dist`;

Author(s)

Sandrine Pavoine <pavoine@mnhn.fr>

References

Pavoine, S. and Bailly, X. (2007) New analysis for consistency among markers in the study of genetic diversity: development and application to the description of bacterial diversity. *BMC Evolutionary Biology*, **7**, e156.

Pavoine, S., Dufour, A.B. and Chessel, D. (2004) From dissimilarities among species to dissimilarities among communities: a double principal coordinate analysis. *Journal of Theoretical Biology*, **228**, 523–537.

See Also

[dpcoa](#)

Examples

```
# The functions used below require the package ape
data(rhizobium)
if (require(ape, quiet = TRUE)) {
  dat <- prep.mdpcoa(rhizobium[[1]], rhizobium[[2]],
    model = c("F84", "F84", "F84", "F81"),
    pairwise.deletion = TRUE)
  sam <- dat$sam
  dis <- dat$dis
  # The distances should be Euclidean.
  # Several transformations exist to render a distance object Euclidean
  # (see functions cailliez, lingoes and quasieuclid in the ade4 package).
```

```

# Here we use the quasieulid function.
dis <- lapply(dis, quasieulid)
mdpcoa1 <- mdpcoa(sam, dis, scannf = FALSE, nf = 2)

# Reference analysis
plot(mdpcoa1)

# Differences between the loci
kplot(mdpcoa1)

# Alleles projected on the population maps.
kplotX.mdpcoa(mdpcoa1)
}

```

meau

Ecological Data : sites-variables, sites-species, where and when

Description

This data set contains information about sites, environmental variables and Ephemeroptera Species.

Usage

```
data(meau)
```

Format

meau is a list of 3 components.

env is a data frame with 24 sites and 10 physicochemical variables.

fau is a data frame with 24 sites and 13 Ephemeroptera Species.

design is a data frame with 24 sites and 2 factors.

- season: is a factor with 4 levels = seasons.
- site: is a factor with 6 levels = sites.

Details

Data set equivalent to [meaudret](#), except that one site (6) along the Bourne (a Meaudret affluent) and one physico chemical variable - the oxygen concentration were added.

Source

Pegaz-Maucet, D. (1980) *Impact d'une perturbation d'origine organique sur la dérive des macro-invertébrés benthiques d'un cours d'eau. Comparaison avec le benthos*. Thèse de troisième cycle, Université Lyon 1, 130 p.

Thioulouse, J., Simier, M. and Chessel, D. (2004) Simultaneous analysis of a sequence of paired ecological tables. *Ecology*, **85**, 1, 272–283.

Examples

```

data(meau)
par(mfrow = c(2,2))
pca1 <- dudi.pca(meau$env, scan = FALSE, nf = 4)
s.class(pca1$li, meau$design$season,
        sub = "Principal Component Analysis")
pca2 <- bca(pca1, meau$design$season, scan = FALSE, nf = 2)
s.class(pca2$ls, meau$design$season, sub = "Between seasons Principal Component Analysis")
s.corcircle(pca1$co)
s.corcircle(pca2$as)

```

meaudret

*Ecological Data : sites-variables, sites-species, where and when***Description**

This data set contains information about sites, environmental variables and Ephemeroptera Species.

Usage

```
data(meaudret)
```

Format

meaudret is a list of 4 components.

env is a data frame with 20 sites and 9 variables.

fau is a data frame with 20 sites and 13 Ephemeroptera Species.

design is a data frame with 20 sites and 2 factors.

- season is a factor with 4 levels = seasons.
- site is a factor with 5 levels = sites along the Meaudret river.

spe.names is a character vector containing the names of the 13 species.

Details

Data set equivalents to [meau](#): site (6) on the Bourne (a Meaudret affluent) and oxygen concentration were removed.

Source

Pegaz-Maucet, D. (1980) *Impact d'une perturbation d'origine organique sur la dérive des macro-invertébrés benthiques d'un cours d'eau. Comparaison avec le benthos*. Thèse de troisième cycle, Université Lyon 1, 130 p.

Thioulouse, J., Simier, M. and Chessel, D. (2004) Simultaneous analysis of a sequence of paired ecological tables. *Ecology*, **85**, 1, 272–283.

Examples

```

data(meaudret)
par(mfrow = c(2,2))
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)
s.class(pca1$li, meaudret$design$season,
        sub = "Principal Component Analysis")
pca2 <- bca(pca1, meaudret$design$season, scan = FALSE, nf = 2)
s.class(pca2$ls, meaudret$design$season, sub = "Between dates Principal Component Analysis")
s.corcircle(pca1$co)
s.corcircle(pca2$as)

```

mfa

*Multiple Factorial Analysis***Description**

performs a multiple factorial analysis, using an object of class ktab.

Usage

```

mfa(X, option = c("lambda1", "inertia", "uniform", "internal"),
    scannf = TRUE, nf = 3)
## S3 method for class 'mfa'
plot(x, xax = 1, yax = 2, option.plot = 1:4, ...)
## S3 method for class 'mfa'
print(x, ...)
## S3 method for class 'mfa'
summary(object, ...)

```

Arguments

X	K-tables, an object of class ktab
option	a string of characters for the weighting of arrays options : lambda1 weighting of group k by the inverse of the first eigenvalue of the k analysis inertia weighting of group k by the inverse of the total inertia of the array k uniform uniform weighting of groups internal weighting included in X\$tabw
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
x, object	an object of class 'mfa'
xax, yax	the numbers of the x-axis and the y-axis
option.plot	an integer between 1 and 4, otherwise the 4 components of the plot are displayed
...	further arguments passed to or from other methods

Value

Returns a list including :

tab	a data frame with the modified array
rank	a vector of ranks for the analyses
eig	a numeric vector with the all eigenvalues
li	a data frame with the coordinates of rows
TL	a data frame with the factors associated to the rows (indicators of table)
co	a data frame with the coordinates of columns
TC	a data frame with the factors associated to the columns (indicators of table)
blo	a vector indicating the number of variables for each table
lisup	a data frame with the projections of normalized scores of rows for each table
link	a data frame containing the projected inertia and the links between the arrays and the reference array

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Escofier, B. and Pagès, J. (1994) Multiple factor analysis (AFMULT package), *Computational Statistics and Data Analysis*, **18**, 121–140.

Examples

```
data(friday87)
w1 <- data.frame(scale(friday87$fau, scal = FALSE))
w2 <- ktab.data.frame(w1, friday87$fau.blo,
  tabnames = friday87$tab.names)
mfa1 <- mfa(w2, scann = FALSE)
mfa1
plot(mfa1)

data(escopage)
w <- data.frame(scale(escopage$tab))
w <- ktab.data.frame(w, escopage$blo, tabnames = escopage$tab.names)
plot(mfa(w, scann = FALSE))
```

 microsatt

Genetic Relationships between cattle breeds with microsatellites

Description

This data set gives genetic relationships between cattle breeds with microsatellites.

Usage

```
data(microsatt)
```

Format

microsatt is a list of 4 components.

tab contains the allelic frequencies for 18 cattle breeds (Taurine or Zebu, French or African) and 9 microsatellites.

loci.names is a vector of the names of loci.

loci.eff is a vector of the number of alleles per locus.

alleles.names is a vector of the names of alleles.

Source

Extract of data prepared by D. Lalo\`e <ugend1a@dga2.jouy.inra.fr> from data used in:

Moazami-Goudarzi, K., D. Lalo\`e, J. P. Furet, and F. Grosclaude (1997) Analysis of genetic relationships between 10 cattle breeds with 17 microsatellites. *Animal Genetics*, **28**, 338–345.

Souvenir Zafindrajaona, P., Zeuh V., Moazami-Goudarzi K., Lalo\`e D., Bourzat D., Idriss A., and Grosclaude F. (1999) Etude du statut phylogénétique du bovin Kouri du lac Tchad à l'aide de marqueurs moléculaires. *Revue d'Élevage et de Médecine Vétérinaire des pays Tropicaux*, **55**, 155–162.

Moazami-Goudarzi, K., Belemsaga D. M. A., Ceriotti G., Lalo\`e D., Fagbohoun F., Kouagou N. T., Sidibé I., Codjia V., Crimella M. C., Grosclaude F. and Touré S. M. (2001) Caractérisation de la race bovine Somba à l'aide de marqueurs moléculaires. *Revue d'Élevage et de Médecine Vétérinaire des pays Tropicaux*, **54**, 1–10.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps055.pdf> (in French).

Examples

```
## Not run:
data(microsatt)
fac <- factor(rep(microsatt$loci.names, microsatt$loci.eff))
w <- dudi.coa(data.frame(t(microsatt$tab)), scann = FALSE)
wit <- within(w, fac, scann = FALSE)
microsatt.ktab <- ktab.within(wit)
```

```
plot(sepan(microsatt.ktab)) # 9 separated correspondence analyses
plot(mcoa(microsatt.ktab, scan = FALSE))
plot(mfa(microsatt.ktab, scan = FALSE))
plot(statis(microsatt.ktab, scan = FALSE))

## End(Not run)
```

mjrochet

Phylogeny and quantitative traits of teleos fishes

Description

This data set describes the phylogeny of 49 teleos fishes as reported by Rochet et al. (2000). It also gives life-history traits corresponding to these 49 species.

Usage

```
data(mjrochet)
```

Format

mjrochet is a list containing the 2 following objects :

tre is a character string giving the phylogenetic tree in Newick format.

tab is a data frame with 49 rows and 7 traits.

Details

Variables of `mjrochet$tab` are the following ones : `tm` (age at maturity (years)), `lm` (length at maturity (cm)), `l05` (length at 5 per cent survival (cm)), `t05` (time to 5 per cent survival (years)), `fb` (slope of the log-log fecundity-length relationship), `fm` (fecundity the year of maturity), `egg` (volume of eggs (mm^3)).

Source

Data taken from:

Summary of data - Clupeiformes : <http://www.ifremer.fr/maerha/clupe.html>

Summary of data - Argentiniformes : <http://www.ifremer.fr/maerha/argentin.html>

Summary of data - Salmoniformes : <http://www.ifremer.fr/maerha/salmon.html>

Summary of data - Gadiformes : <http://www.ifremer.fr/maerha/gadi.html>

Summary of data - Lophiiformes : <http://www.ifremer.fr/maerha/loph.html>

Summary of data - Atheriniformes : <http://www.ifremer.fr/maerha/ather.html>

Summary of data - Perciformes : <http://www.ifremer.fr/maerha/perci.html>

Summary of data - Pleuronectiformes : <http://www.ifremer.fr/maerha/pleuro.html>

Summary of data - Scorpaeniformes : <http://www.ifremer.fr/maerha/scorpa.html>

Phylogenetic tree : http://www.ifremer.fr/maerha/life_history.html

References

Rochet, M. J., Cornillon, P-A., Sabatier, R. and Pontier, D. (2000) Comparative analysis of phylogenetic and fishing effects in life history patterns of teleost fishes. *Oikos*, **91**, 255–270.

Examples

```
data(mjrochet)
mjrochet.phy <- newick2phylog(mjrochet$tre)
tab <- log((mjrochet$tab))
tab0 <- data.frame(scalewt(tab))
table.phylog(tab0, mjrochet.phy, csi = 2, clabel.r = 0.75)
orthogram(tab0[,1], ortho = mjrochet.phy$Bscores)
```

mld

Multi Level Decomposition of unidimensional data

Description

The function `mld` performs an additive decomposition of the input vector `x` onto sub-spaces associated to an orthonormal orthobasis. The sub-spaces are defined by levels of the input factor `level`. The function `haar2level` builds the factor `level` such that the multi level decomposition corresponds exactly to a multiresolution analysis performed with the haar basis.

Usage

```
mld(x, orthobas, level, na.action = c("fail", "mean"),
    plot = TRUE, dfxy = NULL, phylog = NULL, ...)
haar2level(x)
```

Arguments

<code>x</code>	is a vector or a time serie containing the data to be decomposed. This must be a dyadic length vector (power of 2) for the function <code>haar2level</code> .
<code>orthobas</code>	is a data frame containing the vectors of the orthonormal basis.
<code>level</code>	is a factor which levels define the sub-spaces on which the function <code>mld</code> performs the additive decomposition.
<code>na.action</code>	if 'fail' stops the execution of the current expression when <code>x</code> contains any missing value. If 'mean' replaces any missing values by <code>mean(x)</code> .
<code>plot</code>	if TRUE plot <code>x</code> and the components resulting from the decomposition.
<code>dfxy</code>	is a data frame with two coordinates.
<code>phylog</code>	is an object of class <code>phylog</code> .
<code>...</code>	further arguments passed to or from other methods.

Value

A data frame with the components resulting from the decomposition.

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

References

Mallat, S. G. (1989) A theory for multiresolution signal decomposition: the wavelet representation. *IEEE Transactions on Pattern Analysis and Machine Intelligence*, **11**, 7, 674–693.

Percival, D. B. and Walden, A. T. (2000) *Wavelet Methods for Time Series Analysis*, Cambridge University Press.

See Also

[gridrowcol](#), [orthobasis](#), [orthogram](#), [mra](#) for multiresolution analysis with various families of wavelets

Examples

```
## Not run:
# decomposition of a time serie
data(co2)
x <- log(co2)
orthobas <- orthobasis.line(length(x))
level<-rep("D", 467)
level[1:3]<-rep("A", 3)
level[c(77,78,79,81)]<-rep("B", 4)
level[156]<-"C"
level<-as.factor(level)
res <- mld(x, orthobas, level)
sum(scale(x, scale = FALSE) - apply(res, 1, sum))

## End(Not run)
# decomposition of a biological trait on a phylogeny
data(palm)
vfruit<-palm$traits$vfruit
vfruit<-scalewt(vfruit)
palm.phy<-newick2phylog(palm$tre)
level <- rep("F", 65)
level[c(4, 21, 3, 6, 13)] <- LETTERS[1:5]
level <- as.factor(level)
res <- mld(as.vector(vfruit), palm.phy$Bscores, level,
  phylog = palm.phy, clabel.nod = 0.7, f.phylog=0.8,
  csize = 2, clabel.row = 0.7, clabel.col = 0.7)
```

Description

This data set gives the abundance of 32 mollusk species in 163 samples. For each sample, 4 informations are known : the sampling sites, the seasons, the sampler types and the time of exposure.

Usage

```
data(mollusc)
```

Format

mollusc is a list of 2 objects.

fau is a data frame with 163 samples and 32 mollusk species (abundance).

plan contains the 163 samples and 4 variables.

Source

Richardot-Coulet, M., Chessel D. and Bournaud M. (1986) Typological value of the benthos of old beds of a large river. Methodological approach. *Archiv für Hydrobiologie*, **107**, 363–383.

Examples

```
data(mollusc)
coa1 <- dudi.coa(log(mollusc$fau + 1), scannf = FALSE, nf = 3)
par(mfrow = c(2,2))
s.chull(coa1$li, mollusc$plan$site, 2, 3, opt = 1, cpoi = 1)
s.chull(coa1$li, mollusc$plan$season, 2, 3, opt = 1, cpoi = 1)
s.chull(coa1$li, mollusc$plan$method, 2, 3, opt = 1, cpoi = 1)
s.chull(coa1$li, mollusc$plan$duration, 2, 3, opt = 1, cpoi = 1)
par(mfrow=c(1,1))
```

monde84

Global State of the World in 1984

Description

The monde84 data frame gives five demographic variables for 48 countries in the world.

Usage

```
data(monde84)
```


Format

This data frame contains the following columns:

1. `pib`: Gross Domestic Product
2. `croipop`: Growth of the population
3. `morta`: Infant Mortality
4. `anal`: Literacy Rate
5. `scol`: Percentage of children in full-time education

Source

Geze, F. and Coll., eds. (1984) *L'état du Monde 1984 : annuaire économique et géopolitique mondial*. La Découverte, Paris.

Examples

```
data(monde84)
X <- cbind.data.frame(lpib = log(monde84$pib), monde84$croipop)
Y <- cbind.data.frame(lmorta = log(monde84$morta),
  lanal = log(monde84$anal + 1), rscol = sqrt(100 - monde84$scol))
pcaY <- dudi.pca(Y, scan = FALSE)
pcaiv1 <- pcaiv(pcaY, X0 <- scale(X), scan = FALSE)
sum(cor(pcaiv1$11[,1], Y0 <- scale(Y))^2)
pcaiv1$eig[1] #the same
```

morphosport

Athletes' Morphology

Description

This data set gives a morphological description of 153 athletes split in five different sports.

Usage

```
data(morphosport)
```

Format

`morphosport` is a list of 2 objects.

tab is a data frame with 153 athletes and 5 variables.

sport is a factor with 6 items

Details

Variables of `morphosport$tab` are the following ones: `dbi` (biacromial diameter (cm)), `tde` (height (cm)), `tas` (distance from the buttocks to the top of the head (cm)), `lms` (length of the upper limbs (cm)), `poids` (weight (kg)).

The levels of `morphosport$sport` are: `athl` (athletics), `foot` (football), `hand` (handball), `judo`, `nata` (swimming), `voll` (volleyball).

Source

Mimouni , N. (1996) *Contribution de méthodes biométriques à l'analyse de la morphotypologie des sportifs*. Thèse de doctorat. Université Lyon 1.

Examples

```
data(morphosport)
plot(discrimin(dudi.pca(morphosport$tab, scan = FALSE),
  morphosport$sport, scan = FALSE))
```

mstree

Minimal Spanning Tree

Description

Minimal Spanning Tree

Usage

```
mstree(xdist, ngmax = 1)
```

Arguments

<code>xdist</code>	an object of class <code>dist</code> containing an observed dissimilarity
<code>ngmax</code>	a component number (default=1). Select 1 for getting classical MST. To add <code>n</code> supplementary edges <code>k</code> times: select <code>k+1</code> .

Value

returns an object of class `neig`

Author(s)

Daniel Chessel

Examples

```

data(mafragh)
maf.coa = dudi.coa(mafragh$flo, scan = FALSE)
maf.mst = mstree(dist.dudi(maf.coa), 1)
s.label(maf.coa$li, clab = 0, cpoi = 2, neig = maf.mst, cnei = 1)

xy = data.frame(x = runif(20), y = runif(20))
par(mfrow = c(2,2))
for (k in 1:4) {
  neig = mstree (dist.quant(xy,1), k)
  s.label(xy, xlim = c(0,1), ylim = c(0,1), addax = FALSE, neig = neig)
}

```

multispati

Multivariate spatial analysis

Description

This function ensures a multivariate extension of the univariate method of spatial autocorrelation analysis. By accounting for the spatial dependence of data observations and their multivariate covariance simultaneously, complex interactions among many variables are analysed. Using a methodological scheme borrowed from duality diagram analysis, a strategy for the exploratory analysis of spatial pattern in the multivariate is developed.

Usage

```

multispati(dudi, listw, scannf = TRUE, nfposi = 2, nfnega = 0)
## S3 method for class 'multispati'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'multispati'
summary(object, ...)
## S3 method for class 'multispati'
print(x, ...)

```

Arguments

dudi	an object of class dudi for the duality diagram analysis
listw	an object of class listw for the spatial dependence of data observations
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nfposi	an integer indicating the number of kept positive axes
nfnega	an integer indicating the number of kept negative axes
x, object	an object of class multispati
xax, yax	the numbers of the x-axis and the y-axis
...	further arguments passed to or from other methods

Details

This analysis generalizes the Wartenberg's multivariate spatial correlation analysis to various duality diagrams created by the functions (`dudi.pca`, `dudi.coa`, `dudi.acm`, `dudi.mix...`) If `dudi` is a duality diagram created by the function `dudi.pca` and `listw` gives spatial weights created by a row normalized coding scheme, the analysis is equivalent to Wartenberg's analysis.

We note X the data frame with the variables, Q the column weights matrix and D the row weights matrix associated to the duality diagram `dudi`. We note L the neighbouring weights matrix associated to `listw`. Then, the 'multispati' analysis gives principal axes v that maximize the product of spatial autocorrelation and inertia of row scores :

$$I(XQv) * \|XQv\|^2 = v^t Q^t X^t D L X Q v$$

Value

Returns an object of class `multispati`, which contains the following elements :

<code>eig</code>	a numeric vector containing the eigenvalues
<code>nfposi</code>	integer, number of kept axes associated to positive eigenvalues
<code>nfnega</code>	integer, number of kept axes associated to negative eigenvalues
<code>c1</code>	principle axes (v), data frame with p rows and (<code>nfposi</code> + <code>nfnega</code>) columns
<code>li</code>	principal components (XQv), data frame with n rows and (<code>nfposi</code> + <code>nfnega</code>) columns
<code>ls</code>	lag vector onto the principal axes ($LXQv$), data frame with n rows and (<code>nfposi</code> + <code>nfnega</code>) columns
<code>as</code>	principal axes of the <code>dudi</code> analysis (u) onto principal axes of <code>multispati</code> ($t(u)Qv$), data frame with <code>dudi</code> 's n rows and (<code>nfposi</code> + <code>nfnega</code>) columns

Author(s)

Daniel Chessel
 Sebastien Ollier <ollier@biomserv.univ-lyon1.fr>
 Thibaut Jombart <jombart@biomserv.univ-lyon1.fr>

References

- Dray, S., Said, S. and Debias, F. (2008) Spatial ordination of vegetation data using a generalization of Wartenberg's multivariate spatial correlation. *Journal of vegetation science*, **19**, 45–56.
- Grunsky, E. C. and Agterberg, F. P. (1988) Spatial and multivariate analysis of geochemical data from metavolcanic rocks in the Ben Nevis area, Ontario. *Mathematical Geology*, **20**, 825–861.
- Switzer, P. and Green, A.A. (1984) Min/max autocorrelation factors for multivariate spatial imagery. Tech. rep. 6, Stanford University.
- Thioulouse, J., Chessel, D. and Champely, S. (1995) Multivariate analysis of spatial patterns: a unified approach to local and global structures. *Environmental and Ecological Statistics*, **2**, 1–14.
- Wartenberg, D. E. (1985) Multivariate spatial correlation: a method for exploratory geographical analysis. *Geographical Analysis*, **17**, 263–283.
- Jombart, T., Devillard, S., Dufour, A.-B. and Pontier, D. A spatially explicit multivariate method to disentangle global and local patterns of genetic variability. Submitted to *Genetics*.

See Also

[dudi,mat2listw](#)

Examples

```
## Not run:
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  data(mafragh)
  maf.xy <- mafragh$xy
  maf.flo <- mafragh$flo
  maf.listw <- nb2listw(neig2nb(mafragh$neig))
  s.label(maf.xy, neig = mafragh$neig, clab = 0.75)
  maf.coa <- dudi.coa(maf.flo,scannf = FALSE)
  maf.coa.ms <- multispati(maf.coa, maf.listw, scannf = FALSE, nfposi = 2,
    nfnega = 2)
  maf.coa.ms

  ### detail eigenvalues components
  fgraph <- function(obj){
    # use multispati summary
    sum.obj <- summary(obj)
    # compute Imin and Imax
    L <- listw2mat(eval(as.list(obj$call)$listw))
    Imin <- min(eigen(0.5*(L+t(L)))$values)
    Imax <- max(eigen(0.5*(L+t(L)))$values)
    I0 <- -1/(nrow(obj$I)-1)
    # create labels
    labels <- lapply(1:length(obj$eig),function(i) bquote(lambda[.(i)]))
    # draw the plot
    xmax <- eval(as.list(obj$call)$dudi)$eig[1]*1.1
    par(las=1)
    var <- sum.obj[,2]
    moran <- sum.obj[,3]
    plot(x=var,y=moran,type='n',xlab='Inertia',ylab="Spatial autocorrelation (I)",
      xlim=c(0,xmax),ylim=c(Imin*1.1,Imax*1.1),yaxt='n')
    text(x=var,y=moran,do.call(expression,labels))
    ytick <- c(I0,round(seq(Imin,Imax,le=5),1))
    ytlab <- as.character(round(seq(Imin,Imax,le=5),1))
    ytlab <- c(as.character(round(I0,1)),as.character(round(Imin,1)),
      ytlab[2:4],as.character(round(Imax,1)))
    axis(side=2,at=ytick,labels=ytlab)
    rect(0,Imin,xmax,Imax,lty=2)
    segments(0,I0,xmax,I0,lty=2)
    abline(v=0)
    title("Spatial and inertia components of the eigenvalues")
  }
  fgraph(maf.coa.ms)
  ## end eigenvalues details

  par(mfrow = c(1,3))
  barplot(maf.coa$eig)
```

```

barplot(maf.coa.ms$eig)
s.corcircle(maf.coa.ms$as)

par(mfrow = c(2,2))
s.value(maf.xy, -maf.coa$li[,1])
s.value(maf.xy, -maf.coa$li[,2])
s.value(maf.xy, maf.coa.ms$li[,1])
s.value(maf.xy, maf.coa.ms$li[,2])
par(mfrow = c(1,1))

par(mfrow = c(1,2))
w1 <- -maf.coa$li[,1:2]
w1m <- apply(w1, 2, lag.listw, x = maf.listw)
s.match(w1, w1m, clab = 0.75)
w1.ms <- maf.coa.ms$li[,1:2]
w1.msm <- apply(w1.ms, 2, lag.listw, x = maf.listw)
s.match(w1.ms, w1.msm, clab = 0.75)
par(mfrow = c(1,1))

maf.pca <- dudi.pca(mafragh$mil, scannf = FALSE)
multispati.randtest(maf.pca, maf.listw)
maf.pca.ms <- multispati(maf.pca, maf.listw, scannf=FALSE)
plot(maf.pca.ms)
}

## End(Not run)

```

multispati.randtest *Multivariate spatial autocorrelation test (in C)*

Description

This function performs a multivariate autocorrelation test.

Usage

```
multispati.randtest(dudi, listw, nrepet = 999)
```

Arguments

dudi	an object of class dudi for the duality diagram analysis
listw	an object of class listw for the spatial dependence of data observations
nrepet	the number of permutations

Details

We note X the data frame with the variables, Q the column weights matrix and D the row weights matrix associated to the duality diagram *dudi*. We note L the neighbouring weights matrix associated to *listw*. This function performs a Monte-Carlo Test on the multivariate spatial autocorrelation index :

$$r = \frac{\text{trace}(X^t DLXQ)}{\text{trace}(X^t DXQ)}$$

Value

Returns an object of class `randtest` (randomization tests).

Author(s)

Daniel Chessel
Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

References

Smouse, P. E. and Peakall, R. (1999) Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. *Heredity*, **82**, 561–573.

See Also

[dudi,mat2listw](#)

Examples

```
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  data(mafragh)
  maf.listw <- nb2listw(neig2nb(mafragh$neig))
  maf.pca <- dudi.pca(mafragh$mil, scannf = FALSE)
  multispati.randtest(maf.pca, maf.listw)
  maf.pca.ms <- multispati(maf.pca, maf.listw, scannf=FALSE)
  plot(maf.pca.ms)
}
```

`multispati.rtest`

Multivariate spatial autocorrelation test

Description

This function performs a multivariate autocorrelation test.

Usage

```
multispati.rtest(dudi, listw, nrepet = 99)
```

Arguments

dudi	an object of class dudi for the duality diagram analysis
listw	an object of class listw for the spatial dependence of data observations
nrepet	the number of permutations

Details

We note X the data frame with the variables, Q the column weight matrix and D the row weight matrix associated to the duality diagram *dudi*. We note L the neighbouring weights matrix associated to *listw*. This function performs a Monte-Carlo Test on the multivariate spatial autocorrelation index :

$$r = \frac{X^t DLXQ}{X^t DXQ}$$

Value

Returns an object of class randtest (randomization tests).

Author(s)

Daniel Chessel
Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

References

Smouse, P. E. and Peakall, R. (1999) Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. *Heredity*, **82**, 561–573.

See Also

[dudi,mat2listw](#)

Examples

```
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  data(mafragh)
  maf.listw <- nb2listw(neig2nb(mafragh$neig))
  maf.pca <- dudi.pca(mafragh$mil, scannf = FALSE)
  multispati.rtest(maf.pca, maf.listw)
  maf.pca.ms <- multispati(maf.pca, maf.listw, scannf=FALSE)
  plot(maf.pca.ms)
}
```


Description

neig creates objects of class neig with :

- a list of edges
- a binary square matrix
- a list of vectors of neighbours
- an integer (linear and circular graphs)
- a data frame of polygons (area)

scores.neig returns the eigenvectors of neighbouring, orthonormalized scores (null average, unit variance $1/n$ and null covariances) of maximal autocorrelation.

nb2neig returns an object of class neig using an object of class nb in the library 'spdep'

neig2nb returns an object of class nb using an object of class neig

neig2mat returns the incidence matrix between edges (1 = neighbour ; 0 = no neighbour)

neig.util.GtoL and neig.util.LtoG are utilities.

Usage

```
neig(list = NULL, mat01 = NULL, edges = NULL,
     n.line = NULL, n.circle = NULL, area = NULL)
```

```
scores.neig (obj)
## S3 method for class 'neig'
print(x, ...)
## S3 method for class 'neig'
summary(object, ...)
nb2neig (nb)
neig2nb (neig)
neig2mat (neig)
```

Arguments

list	a list which each component gives the number of neighbours
mat01	a symmetric square matrix of 0-1 values
edges	a matrix of 2 columns with integer values giving a list of edges
n.line	the number of points for a linear plot
n.circle	the number of points for a circular plot
area	a data frame containing a polygon set (see area.plot)
nb	an object of class 'nb'

neig, x, obj, object
 an object of class 'neig'
 ... further arguments passed to or from other methods

Author(s)

Daniel Chessel

References

Thioulouse, J., D. Chessel, and S. Champely. 1995. Multivariate analysis of spatial patterns: a unified approach to local and global structures. *Environmental and Ecological Statistics*, 2, 1–14.

Examples

```
data(mafragh)
if (require(deldir, quietly=TRUE)) {
  par(mfrow = c(2,1))
  provi <- deldir(mafragh$xy)
  provi.neig <- neig(edges = as.matrix(provi$delsgs[,5:6]))

  s.label(mafragh$xy, neig = provi.neig, inc = FALSE,
          addax = FALSE, clab = 0, cnei = 2)
  dist <- apply(provi.neig, 1, function(x)
    sqrt(sum((mafragh$xy[x[1],] - mafragh$xy[x[2],])^2)))
  #hist(dist, nclass = 50)
  mafragh.neig <- neig(edges = provi.neig[dist<50,])
  s.label(mafragh$xy, neig = mafragh.neig, inc = FALSE,
          addax = FALSE, clab = 0, cnei = 2)
  par(mfrow = c(1,1))

  data(irishdata)
  irish.neig <- neig(area = irishdata$area)
  summary(irish.neig)
  print(irish.neig)
  s.label(irishdata$xy, neig = irish.neig, cneig = 3,
          area = irishdata$area, clab = 0.8, inc = FALSE)

  irish.scores <- scores.neig(irish.neig)
  par(mfrow = c(2,3))
  for (i in 1:6) s.value(irishdata$xy, irish.scores[,i],
    inc = FALSE, grid = FALSE, addax = FALSE,
    neig = irish.neig,
    csi = 2, cleg = 0, sub = paste("Eigenvector ",i), csub = 2)
  par(mfrow = c(1,1))

  a.neig <- neig(n.circle = 16)
  a.scores <- scores.neig(a.neig)
  xy <- cbind.data.frame(cos((1:16) * pi / 8), sin((1:16) * pi / 8))
  par(mfrow = c(4,4))
  for (i in 1:15) s.value(xy, a.scores[,i], neig = a.neig,
    csi = 3, cleg = 0)
```

```

par(mfrow = c(1,1))

a.neig <- neig(n.line = 28)
a.scores <- scores.neig(a.neig)
par(mfrow = c(7,4))
par(mar = c(1.1,2.1,0.1,0.1))
for (i in 1:27) barplot(a.scores[,i], col = grey(0.8))
}
par(mfrow = c(1,1))

if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  data(columbus)
  par(mfrow = c(2,1))
  par(mar = c(0.1,0.1,0.1,0.1))
  plot(col.gal.nb, coords)
  s.label(data.frame(coords), neig = neig(list = col.gal.nb),
    inc = FALSE, clab = 0.6, cneig = 1)
  par(mfrow = c(1,1))

  data(mafragh)
  maf.rel <- relativeneigh(as.matrix(mafragh$xy))
  maf.rel <- graph2nb(maf.rel)
  s.label(mafragh$xy, neig = neig(list = maf.rel), inc = FALSE,
    clab = 0, addax = FALSE, cne = 1, cpo = 2)

  par(mfrow = c(2,2))
  w <- matrix(runif(100), 50, 2)
  x.gab <- gabrielneigh(w)
  x.gab <- graph2nb(x.gab)
  s.label(data.frame(w), neig = neig(list = x.gab), inc = FALSE,
    clab = 0, addax = FALSE, cne = 1, cpo = 2, sub = "relative")
  x.rel <- relativeneigh(w)
  x.rel <- graph2nb(x.rel)
  s.label(data.frame(w), neig = neig(list = x.rel), inc = FALSE,
    clab = 0, addax = FALSE, cne = 1, cpo = 2, sub = "Gabriel")
  k1 <- knn2nb(knearneigh(w))
  s.label(data.frame(w), neig = neig(list = k1), inc = FALSE,
    clab = 0, addax = FALSE, cne = 1, cpo = 2, sub = "k nearest neighbours")

  all.linked <- max(unlist(nbdists(k1, w)))
  z <- dnearneigh(w, 0, all.linked)
  s.label(data.frame(w), neig = neig(list = z), inc = FALSE,
    clab = 0, addax = FALSE, cne = 1, cpo = 2,
    sub = "Neighbourhood contiguity by distance")
}
par(mfrow = c(1,1))

```

Description

This data set contains various examples of phylogenetic trees in Newick format.

Usage

```
data(newick.eg)
```

Format

newick.eg is a list containing 14 character strings in Newick format.

Source

Trees 1 to 7 were obtained from the URL

<http://evolution.genetics.washington.edu/phylip/newicktree.html>.

Trees 8 and 9 were obtained by Clémentine Carpentier-Gimaret <gimaret@biomserv.univ-lyon1.fr>.

Tree 10 was obtained from Treezilla Data Sets starting from

<http://www.cis.upenn.edu/~krice/treezilla/>.

Trees 11 and 12 are taken from Bauwens and D'iaz-Uriarte (1997).

Tree 13 is taken from Cheverud and Dow (1985).

Tree 13 is taken from Martins and Hansen (1997).

References

Bauwens, D. and D'iaz-Uriarte, R. (1997) Covariation of life-history traits in lacertid lizards: a comparative study. *American Naturalist*, **149**, 91–111.

Cheverud, J. and Dow, M.M. (1985) An autocorrelation analysis of genetic variation due to lineal fission in social groups of rhesus macaques. *American Journal of Physical Anthropology*, **67**, 113–122.

Martins, E. P. and Hansen, T.F. (1997) Phylogenies and the comparative method: a general approach to incorporating phylogenetic information into the analysis of interspecific data. *American Naturalist*, **149**, 646–667.

Examples

```
data(newick.eg)
newick2phylog(newick.eg[[11]])
radial.phylog(newick2phylog(newick.eg[[7]]), circ = 1,
  clabel.1 = 0.75)
```

newick2phylog	<i>Create phylogeny</i>
---------------	-------------------------

Description

The first three functions ensure to create object of class phylog from either a character string in Newick format (newick2phylog) or an object of class 'hclust' (hclust2phylog) or a taxonomy (taxo2phylog). The function newick2phylog.addtools is an internal function called by newick2phylog, hclust2phylog and taxo2phylog when newick2phylog.addtools = TRUE. It adds some items in 'phylog' objects.

Usage

```
newick2phylog(x.tre, add.tools = TRUE, call = match.call())
hclust2phylog(hc, add.tools = TRUE)
taxo2phylog(taxo, add.tools = FALSE, root="Root", abbrev=TRUE)
newick2phylog.addtools(res, tol = 1e-07)
```

Arguments

x.tre	a character string corresponding to a phylogenetic tree in Newick format (http://evolution.genetics.washington.edu/phylip/newicktree.html)
add.tools	if TRUE, executes the function newick2phylog.addtools
call	call
hc	an object of class hclust
taxo	an object of class taxo
res	an object of class phylog (an internal argument of the function newick2phylog)
tol	used in case 3 of method as a tolerance threshold for null eigenvalues
root	a character string for the root of the tree
abbrev	logical : if TRUE levels are abbreviated by column and two characters are added before

Value

Return object of class phylog.

Author(s)

Daniel Chessel
Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

See Also

[phylog](#), [plot.phylog](#), [as.taxo](#)

Examples

```

w <- "(((, ), (, ), ), (, ));"
w.phy <- newick2phylog(w)
print(w.phy)
plot(w.phy)

## Not run:
# newick2phylog
data(newick.eg)
radial.phylog(newick2phylog(newick.eg[[8]], FALSE), cnode = 1,
  clabel.l = 0.8)

w <- NULL
w[1] <- "(, (((((((((((((((((((((, (, (, ))), ), (((, (, ), (, ), ), (, (, ), (, ), (((((("
w[2] <- ", (, ), ), ), (, ), (((((, (, ), ((, (, ), ))), (, ), (, (, ), ((, (, ), ), (, "
w[3] <- "(((, (, ), (, (, ))), (, ), ((, (, ), ))), ((, (, (, ), ), (, ), (, (, ), "
w[4] <- ")), (((((((, (, ), ), ), ), ((, (, ), ), (, ), ((, (, ), ), (((((, (, ), ), (, ), "
w[5] <- ")), ((, (, (, (, ))), (, ), ((, (, ), (((((((, (, ), ), ), (, ), (, ), ), ((, "
w[6] <- ", )))), (, (, (, ), (, ), ((, (, ), ))), (((((, (, (, ))), ((, (, ), ), ((, (, ), "
w[7] <- ", ))), ((, (, ), ((, (, ), ))), ((, (, ))), (((((((, (, ), ), (, ), ), (, "
w[8] <- ", (, ))), (((((((, (, ), (, ), ((, (, (, ), (, (, (, ))), ((, (, (, ))), "
w[9] <- ")), ((, (, ))), (((((((, (, ), (, ), ((, (, ))), ), ), ((, (, (, ))), (, (, (, "
w[10] <- ")), (, ), ), (, (, (, ))), ((, (, (, ), (, ), ((, (, (, ), (, ), (, ), (, ))), (, "
w[11] <- "))) (, , , ))), (((, (, ), ), ((, (, (, ))), ((, (, ), (, ))), (, ))), (, ((, (, "
w[12] <- ", ))), ((, (, ), ))), ((, (, ), (, ), (, (, ))), ((, (, ), (, ), (, (, ))), "
w[13] <- "(((, (, ), ), (((((((, , , , ), (, ), (, ), (, (, ), ))), ), (, (((, (, (((("
w[14] <- ", (, ), ), ), ), ), ((, (, ), ((, (, ), (, ))), ))), ((, (, ), ((, (, ), (, )) "
w[15] <- ")), ), ), ), ((, ))), ((, (, ), ((, (, ), ))), ), ((, ))), (, ), (, ))), (( "
w[16] <- ", ), ), (((, (, ), (, (, ), ((, (, ), (, (, (, ), ))), ))), (, ), ), ), ), "
w[17] <- ")), (((, (, ), ), (, (, ), (, (, (, ))), ))), (((, (, (, ), ), (, ), ), (, (, "
w[18] <- ")), (((, (, (, ), ), ((, (, ), ), (, ))), ), ), ), (, (, ), ), ), (((, (, "
w[19] <- "(, (, ), ), ((, (, ), ), ), ), (((((((, (((, (, ), (, (, ))), ((, (, ), ), "
w[20] <- "(, ))), ), ), ), (, ), ), ), ((, ))), ((, ))), (((((((, (, ), (((((((("
w[21] <- " , ), (, (, ), (, ), ), ), (, ), ), (((, (, ), (, (, ), ))), (, ), (, )) "
w[22] <- " , ), ), ), (, ), ), (, (, ), ), ), (, (, (, ))), ), (, ), ), );"
phy1 <- newick2phylog(w, FALSE)
phy1
radial.phylog(phy1, clabel.l = 0, circle = 2.2, clea = 0.5,
  cnod = 0.5)
data(newick.eg)
radial.phylog(newick2phylog(newick.eg[[8]], FALSE), cnode = 1,
  clabel.l = 0.8)

# hclust2phylog
data(USArrests)
hc <- hclust(dist(USArrests), "ave")
par(mfrow = c(1, 2))
plot(hc, hang = -1)
phy <- hclust2phylog(hc)
plot.phylog(phy, clabel.l = 0.75, clabel.n = 0.6, f = 0.75)

```

```

par(mfrow = c(1,1))
row.names(USArrests)
names(phy$leaves) #WARNING not the same for two reasons
row.names(USArrests) <- gsub(" ", "_", row.names(USArrests))
row.names(USArrests)
names(phy$leaves) #WARNING not the same for one reason
USArrests <- USArrests[names(phy$leaves),]
row.names(USArrests)
names(phy$leaves) #the same
table.phylog(data.frame(scalewt(USArrests)), phy, csi = 2.5,
  clabel.r = 0.75, f = 0.7)

#taxo2phylog
data(taxo.eg)
tax <- as.taxo(taxo.eg[[1]])
tax.phy <- taxo2phylog(as.taxo(taxo.eg[[1]]))
par(mfrow = c(1,2))
plot.phylog(tax.phy, clabel.l = 1.25, clabel.n = 1.25, f = 0.75)
plot.phylog(taxo2phylog(as.taxo(taxo.eg[[1]][sample(15),])),
  clabel.l = 1.25, clabel.n = 1.25, f = 0.75)

par(mfrow=c(1,1))
plot.phylog(taxo2phylog(as.taxo(taxo.eg[[2]])), clabel.l = 1,
  clabel.n = 0.75, f = 0.65)

## End(Not run)

```

niche

Method to Analyse a pair of tables : Environmental and Faunistic Data

Description

performs a special multivariate analysis for ecological data.

Usage

```

niche(dudiX, Y, scannf = TRUE, nf = 2)
## S3 method for class 'niche'
print(x, ...)
## S3 method for class 'niche'
plot(x, xax = 1, yax = 2, ...)
niche.param(x)
## S3 method for class 'niche'
rtest(xtest, nrepet=99, ...)

```

Arguments

dudiX a duality diagram providing from a function dudi.coa, dudi.pca, ... using an array sites-variables

Y	a data frame sites-species according to <code>dudiX\$tab</code> with no columns of zero
<code>scannf</code>	a logical value indicating whether the eigenvalues bar plot should be displayed
<code>nf</code>	if <code>scannf</code> FALSE, an integer indicating the number of kept axes
<code>x</code>	an object of class <code>niche</code>
<code>...</code>	further arguments passed to or from other methods
<code>xax, yax</code>	the numbers of the x-axis and the y-axis
<code>xtest</code>	an object of class <code>niche</code>
<code>nrepet</code>	the number of permutations for the testing procedure

Value

Returns a list of the class `niche` (sub-class of `dudi`) containing :

<code>rank</code>	an integer indicating the rank of the studied matrix
<code>nf</code>	an integer indicating the number of kept axes
<code>RV</code>	a numeric value indicating the RV coefficient
<code>eig</code>	a numeric vector with the all eigenvalues
<code>lw</code>	a data frame with the row weights (crossed array)
<code>tab</code>	a data frame with the crossed array (averaging species/sites)
<code>li</code>	a data frame with the species coordinates
<code>l1</code>	a data frame with the species normed scores
<code>co</code>	a data frame with the variable coordinates
<code>c1</code>	a data frame with the variable normed scores
<code>ls</code>	a data frame with the site coordinates
<code>as</code>	a data frame with the axis upon niche axis

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>
 Stephane Dray <dray@biomserv.univ-lyon1.fr>

References

Dolédec, S., Chessel, D. and Gimaret, C. (2000) Niche separation in community analysis: a new method. *Ecology*, **81**, 2914–2927.

Examples

```
data(doubs)
dudi1 <- dudi.pca(doubs$env, scale = TRUE, scan = FALSE, nf = 3)
nic1 <- niche(dudi1, doubs$fish, scann = FALSE)

par(mfrow = c(2,2))
s.traject(dudi1$li, clab = 0)
```



```

s.traject(nic1$ls, clab = 0)
s.corcircle(nic1$as)
s.arrow(nic1$c1)

par(mfrow = c(5,6))
for (i in 1:27) s.distrib(nic1$ls, as.data.frame(doubs$fish[,i]),
  csub = 2, sub = names(doubs$fish)[i])

par(mfrow = c(1,1))
s.arrow(nic1$li, clab = 0.7)

par(mfrow = c(1,1))
data(trichometeo)
pca1 <- dudi.pca(trichometeo$meteo, scan = FALSE)
nic1 <- niche(pca1, log(trichometeo$fau + 1), scan = FALSE)
plot(nic1)
niche.param(nic1)
rtest(nic1,19)

data(rpjdl)
plot(niche(dudi.pca(rpjdl$mil, scan = FALSE), rpjdl$fau, scan = FALSE))

```

nipals

Non-linear Iterative Partial Least Squares (NIPALS) algorithm

Description

This function performs NIPALS algorithm, i.e. a principal component analysis of a data table that can contain missing values.

Usage

```

nipals(df, nf = 2, rec = FALSE, niter = 100, tol = 1e-09)
## S3 method for class 'nipals'
scatter(x, xax = 1, yax = 2, clab.row = 0.75, clab.col
= 1, posieig = "top", sub = NULL, ...)
## S3 method for class 'nipals'
print(x, ...)

```

Arguments

df	a data frame that can contain missing values
nf	an integer, the number of axes to keep
rec	a logical that specify if the functions must perform the reconstitution of the data using the nf axes
niter	an integer, the maximum number of iterations
tol	a real, the tolerance used in the iterative algorithm

x	an object of class nipals
xax	the column number for the x-axis
yax	the column number for the y-axis
clab.row	a character size for the rows
clab.col	a character size for the columns
posieig	if "top" the eigenvalues bar plot is upside, if "bottom" it is downside, if "none" no plot
sub	a string of characters to be inserted as legend
...	further arguments passed to or from other methods

Details

Data are scaled (mean 0 and variance 1) prior to the analysis.

Value

Returns a list of classes nipals:

tab	the scaled data frame
eig	the pseudoeigenvalues
rank	the rank of the analyzed matrice
nf	the number of factors
c1	the column normed scores
co	the column coordinates
li	the row coordinates
call	the call function
nb	the number of iterations for each axis
rec	a data frame obtained by the reconstitution of the scaled data using the nf axes

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>

References

Wold, H. (1966) Estimation of principal components and related models by iterative least squares. In P. Krishnaiah, editors. *Multivariate Analysis*, Academic Press, 391–420.

Wold, S., Esbensen, K. and Geladi, P. (1987) Principal component analysis *Chemometrics and Intelligent Laboratory Systems*, **2**, 37–52.

See Also

[dudi.pca](#)

Examples

```

data(doubs)
## nipals is equivalent to dudi.pca when there are no NA
acp1 <- dudi.pca(doubs$env, scannf = FALSE, nf = 2)
nip1 <- nipals(doubs$env)
par(mfrow=c(2,2))
barplot(acp1$eig, main = "dudi.pca")
barplot(nip1$eig, main = "nipals")
plot(acp1$c1[,1], nip1$c1[,1], main = "col scores", xlab="dudi.pca", ylab="nipals")
plot(acp1$li[,1], nip1$li[,1], main = "row scores", xlab="dudi.pca",
ylab="nipals")

## Not run:
## with NAs:
doubs$env[1,1] <- NA
nip2 <- nipals(doubs$env)
cor(nip1$li, nip2$li)
nip1$eig
nip2$eig

## End(Not run)

```

njplot

Phylogeny and trait of bacteria

Description

This data set describes the phylogeny of 36 bacteria as reported by Perrière and Gouy (1996). It also gives the GC rate corresponding to these 36 species.

Usage

```
data(njplot)
```

Format

njplot is a list containing the 2 following objects:

tre is a character string giving the fission tree in Newick format.

tauxcg is a numeric vector that gives the CG rate of the 36 species.

Source

Data were obtained by Manolo Gouy <mgouy@biomserv.univ-lyon1.fr>

References

Perrière, G. and Gouy, M. (1996) WWW-Query : an on-line retrieval system for biological sequence banks. *Biochimie*, **78**, 364–369.

Examples

```

data(njplot)
njplot.phy <- newick2phylog(njplot$tre)
par(mfrow = c(2,1))
tauxcg0 <- njplot$tauxcg - mean(njplot$tauxcg)
symbols.phylog(njplot.phy, squares = tauxcg0)
symbols.phylog(njplot.phy, circles = tauxcg0)
par(mfrow = c(1,1))

```

olympic

Olympic Decathlon

Description

This data set gives the performances of 33 men's decathlon at the Olympic Games (1988).

Usage

```
data(olympic)
```

Format

olympic is a list of 2 components.

tab is a data frame with 33 rows and 10 columns events of the decathlon: 100 meters (100), long jump (long), shotput (poid), high jump (haut), 400 meters (400), 110-meter hurdles (110), discus throw (disq), pole vault (perc), javelin (jave) and 1500 meters (1500).

score is a vector of the final points scores of the competition.

Source

Example 357 in:

Hand, D.J., Daly, F., Lunn, A.D., McConway, K.J. and Ostrowski, E. (1994) *A handbook of small data sets*, Chapman & Hall, London. 458 p.

Lunn, A. D. and McNeil, D.R. (1991) *Computer-Interactive Data Analysis*, Wiley, New York

Examples

```

data(olympic)
pca1 <- dudi.pca(olympic$tab, scan = FALSE)
par(mfrow = c(2,2))
barplot(pca1$eig)
s.corcircle(pca1$co)
plot(olympic$score, pca1$l1[,1])
abline(lm(pca1$l1[,1]~olympic$score))
s.label(pca1$l1, clab = 0.5)
s.arrow(2 * pca1$co, add.p = TRUE)
par(mfrow = c(1,1))

```

`optimEH`*Nee and May's optimizing process*

Description

performs Nee and May's optimizing scheme. When branch lengths in an ultrametric phylogenetic tree are expressed as divergence times, the total sum of branch lengths in that tree expresses the amount of evolutionary history. Nee and May's algorithm optimizes the amount of evolutionary history preserved if only k species out of n were to be saved. The $k-1$ closest-to-root nodes are selected, which defines k clades; one species from each clade is picked. At this last step, we decide to select the most original species of each from the k clades.

Usage

```
optimEH(phy1, nbofsp, tol = 1e-8, give.list = TRUE)
```

Arguments

<code>phy1</code>	an object of class <code>phylog</code>
<code>nbofsp</code>	an integer indicating the number of species saved (k).
<code>tol</code>	a tolerance threshold for null values (a value less than <code>tol</code> in absolute terms is considered as <code>NULL</code>).
<code>give.list</code>	logical value indicating whether a list of optimizing species should be provided. If <code>give.list = TRUE</code> , <code>optimEH</code> provides the list of the k species which optimize the amount of evolutionary history preserved and are the most original species in their clades. If <code>give.list = FALSE</code> , <code>optimEH</code> returns directly the real value giving the amount of evolutionary history preserved.

Value

Returns a list containing:

<code>value</code>	a real value providing the amount of evolutionary history preserved.
<code>selected.sp</code>	a data frame containing the list of the k species which optimize the amount of evolutionary history preserved and are the most original species in their clades.

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Nee, S. and May, R.M. (1997) Extinction and the loss of evolutionary history. *Science* **278**, 692–694.

Pavoine, S., Ollier, S. and Dufour, A.-B. (2005) Is the originality of a species measurable? *Ecology Letters*, **8**, 579–586.

See Also[randEH](#)**Examples**

```
data(carni70)
carni70.phy <- newick2phylog(carni70$tre)
optimEH(carni70.phy, nbofsp = 7, give.list = TRUE)
```

oribatid

Oribatid mite

Description

This data set contains informations about environmental control and spatial structure in ecological communities of Oribatid mites.

Usage

```
data(oribatid)
```

Format

oribatid is a list containing the following objects :

fau : a data frame with 70 rows (sites) and 35 columns (Oribatid species)

envir : a data frame with 70 rows (sites) and 5 columns (environmental variables)

xy : a data frame that contains spatial coordinates of the 70 sites

Details

Variables of oribatid\$envir are the following ones :

substrate: a factor with seven levels that describes the nature of the substratum

shrubs: a factor with three levels that describes the absence/presence of shrubs

topo: a factor with two levels that describes the microtopography

density: substratum density ($g.L^{-1}$)

water: water content of the substratum ($g.L^{-1}$)

Source

Data prepared by P. Legendre <Pierre.Legendre@umontreal.ca> and

D. Borcard <borcardd@magellan.umontreal.ca> starting from

<http://www.fas.umontreal.ca/biol/casgrain/fr/labo/oribates.html>

References

Borcard, D., and Legendre, P. (1994) Environmental control and spatial structure in ecological communities: an example using Oribatid mites (*Acari Oribatei*). *Environmental and Ecological Statistics*, **1**, 37–61.

Borcard, D., Legendre, P., and Drapeau, P. (1992) Partialling out the spatial component of ecological variation. *Ecology*, **73**, 1045–1055.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps039.pdf> (in French).

Examples

```
data(oribatid)
ori.xy <- oribatid$xy[,c(2,1)]
names(ori.xy) <- c("x","y")
plot(ori.xy,pch = 20, cex = 2, asp = 1)

if (require(deldir, quiet = TRUE)) {
  if (require(spdep, quiet = TRUE)) {
    plot(deldir(ori.xy), add = TRUE)
    s.label(ori.xy, add.p = TRUE,
            neig = nb2neig(knn2nb(knearneigh(as.matrix(ori.xy), 3))),
            clab = 0)
  }
}
```

originality

Originality of a species

Description

computes originality values for species from an ultrametric phylogenetic tree.

Usage

```
originality(phy1, method = 5)
```

Arguments

phy1 an object of class phylog
method a vector containing integers between 1 and 7.

Details

1 = Vane-Wright et al.'s (1991) node-counting index 2 = May's (1990) branch-counting index 3 = Nixon and Wheeler's (1991) unweighted index, based on the sum of units in binary values 4 = Nixon and Wheeler's (1991) weighted index 5 = QE-based index 6 = Isaac et al. (2007) ED index 7 = Redding et al. (2006) Equal-split index

Value

Returns a data frame with species in rows, and the selected indices of originality in columns. Indices are expressed as percentages.

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

- Isaac, N.J.B., Turvey, S.T., Collen, B., Waterman, C. and Baillie, J.E.M. (2007) Mammals on the EDGE: conservation priorities based on threat and phylogeny. *PloS ONE*, **2**, e–296.
- Redding, D. and Mooers, A. (2006) Incorporating evolutionary measures into conservation prioritization. *Conservation Biology*, **20**, 1670–1678.
- Pavoine, S., Ollier, S. and Dufour, A.-B. (2005) Is the originality of a species measurable? *Ecology Letters*, **8**, 579–586.
- Vane-Wright, R.I., Humphries, C.J. and Williams, P.H. (1991). What to protect? Systematics and the agony of choice. *Biological Conservation*, **55**, 235–254.
- May, R.M. (1990). Taxonomy as destiny. *Nature*, **347**, 129–130.
- Nixon, K.C. and Wheeler, Q.D. (1992). Measures of phylogenetic diversity. In: *Extinction and Phylogeny* (eds. Novacek, M.J. and Wheeler, Q.D.), 216–234, Columbia University Press, New York.

Examples

```
data(carni70)
carni70.phy <- newick2phylog(carni70$tre)
ori.tab <- originality(carni70.phy, 1:7)
names(ori.tab)
dotchart.phylog(carni70.phy, ori.tab, scaling = FALSE, yjoining = 0,
  ranging = FALSE, cleaves = 0, ceti = 0.5, csub = 0.7, cdot = 0.5)
```

orisaved	<i>Maximal or minimal amount of originality saved under optimal conditions</i>
----------	--

Description

computes the maximal or minimal amount of originality saved over all combinations of species optimizing the amount of evolutionary history preserved. The originality of a species is measured with the QE-based index.

Usage

```
orisaved(phy1, rate = 0.1, method = 1)
```


Arguments

phy1	an object of class phylog
rate	a real value (between 0 and 1) indicating how many species will be saved for each calculation. For example, if the total number of species is 70 and 'rate = 0.1' then the calculations will be done at a rate of 10 % i.e. for 0 (= 0 %), 7 (= 10 %), 14 (= 20 %), 21 (= 30 %), ..., 63 (= 90 %) and 70(= 100 %) species saved. If 'rate = 0.5' then the calculations will be done for only 0 (= 0 %), 35 (= 50 %) and 70(= 100 %) species saved.
method	an integer either 1 or 2 (see details).

Details

1 = maximum amount of originality saved 2 = minimum amount of originality saved

Value

Returns a numeric vector.

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Pavoine, S., Ollier, S. and Dufour, A.-B. (2005) Is the originality of a species measurable? *Ecology Letters*, **8**, 579–586.

Examples

```
data(carni70)
carni70.phy <- newick2phylog(carni70$tre)
tmax <- orisaved(carni70.phy, rate = 1 / 70, method = 1)
tmin <- orisaved(carni70.phy, rate = 1 / 70, method = 2)
plot(c(0, 1:70), tmax, xlab = "nb of species saved", ylab = "Originality saved", type = "l")
lines(c(0, 1:70), tmin, lty = 2)
```

orthobasis

Orthonormal basis for orthonormal transform

Description

These functions returns object of class 'orthobasis' that contains data frame with n rows and $n-1$ columns. Each data frame defines an orthonormal basis for the uniform weights.

orthobasic.neig returns the eigen vectors of the matrix $N-M$ where M is the symmetric n by n matrix of the between-sites neighbouring graph and N is the diagonal matrix of neighbour numbers.

orthobasis.line returns the analytical solution for the linear neighbouring graph.

orthobasic.circ returns the analytical solution for the circular neighbouring graph.

orthobasis.mat returns the eigen vectors of the general link matrix M .
 orthobasis.listw returns the eigen vectors of the general link matrix M associated to a listw object.
 orthobasis.haar returns wavelet haar basis.

Usage

```
orthobasis.neig(neig)
orthobasis.line(n)
orthobasis.circ(n)
orthobasis.mat(mat, cnw=TRUE)
orthobasis.listw(listw)
orthobasis.haar(n)
## S3 method for class 'orthobasis'
print(x,...)
```

Arguments

neig	is an object of class neig
n	is an integer that defines length of vectors
mat	is a n by n phylogenetic or spatial link matrix
listw	is a 'listw' object
cnw	if TRUE, the matrix of the neighbouring graph is modified to give Constant Neighbouring Weights
x	is an object of class orthobasis
...	: further arguments passed to or from other methods

Value

All the functions excepted print.orthobasis return an object of class orthobasis containing a data frame. This data frame defines an orthonormal basis with $n-1$ vectors of length n . Various attributes are associated to it :

names	: names of the vectors
row.names	: row names of the data frame
class	: class
values	: row weights (uniform weights)
weights	: numeric values to class vectors according to their quadratic forms (Moran ones)
call	: call

Note

the function orthobasis.haar uses function [wavelet.filter](#) from package waveslim.

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>
Daniel Chessel

References

Misiti, M., Misiti, Y., Oppenheim, G. and Poggi, J.M. (1993) Analyse de signaux classiques par décomposition en ondelettes. *Revue de Statistique Appliquée*, **41**, 5–32.

Cornillon, P.A. (1998) *Prise en compte de proximités en analyse factorielle et comparative*. Thèse, Ecole Nationale Supérieure Agronomique, Montpellier.

See Also

[gridrowcol](#) that defines an orthobasis for square grid, [phylog](#) that defines an orthobasis for phylogenetic tree, [orthogram](#) and [mld](#)

Examples

```
# a 2D spatial orthobasis
par(mfrow = c(4,4))
w <- gridrowcol(8,8)
for (k in 1:16)
  s.value(w$xy, w$orthobasis[,k], cleg = 0, csi = 2, incl = FALSE,
    addax = FALSE, sub = k, csub = 4, ylim = c(0,10), cgri = 0)
par(mfrow = c(1,1))
barplot(attr(w$orthobasis, "values"))

# Haar 1D orthobasis
w <- orthobasis.haar(32)
par(mfrow = c(8,4))
par(mar = c(0.1,0.1,0.1,0.1))
for (k in 1:31) {
  plot(w[,k], type="S", xlab = "", ylab = "", xaxt = "n",
    yaxt = "n", xaxs = "i", yaxs = "i", ylim=c(-4.5,4.5))
  points(w[,k], type = "p", pch = 20, cex = 1.5)
}

# a 1D orthobasis
w <- orthobasis.line(n = 33)
par(mfrow = c(8,4))
par(mar = c(0.1,0.1,0.1,0.1))
for (k in 1:32) {
  plot(w[,k], type="l", xlab = "", ylab = "", xaxt = "n",
    yaxt = "n", xaxs = "i", yaxs = "i", ylim=c(-1.5,1.5))
  points(w[,k], type = "p", pch = 20, cex = 1.5)
}

par(mfrow = c(1,1))
barplot(attr(w, "values"))
```

```

w <- orthobasis.circ(n = 26)
#par(mfrow = c(5,5))
#par(mar = c(0.1,0.1,0.1,0.1))
# for (k in 1:25)
#   dotcircle(w[,k], xlim = c(-1.5,1.5), cleg = 0)

par(mfrow = c(1,1))
#barplot(attr(w, "values"))

## Not run:
# a spatial orthobasis
data(mafragh)
w <- orthobasis.neig(mafragh$neig)
par(mfrow = c(4,2))
for (k in 1:8)
  s.value(mafragh$xy, w[,k],cleg = 0, sub = as.character(k),
          csub = 3)

par(mfrow = c(1,1))
barplot(attr(w, "values"))

# a phylogenetic orthobasis
data(njplot)
phy <- newick2phylog(njplot$tre)
wA <- phy$Ascores
wW <- phy$Wscores
table.phylog(phylog = phy, wA, clabel.row = 0, clabel.col = 0.5)
table.phylog(phylog = phy, wW, clabel.row = 0, clabel.col = 0.5)

## End(Not run)

```

orthogram

Orthonormal decomposition of variance

Description

This function is deprecated. See orthogram in adephylo.

This function performs the orthonormal decomposition of variance of a quantitative variable on an orthonormal basis. It also returns the results of five non parametric tests associated to the variance decomposition. It thus provides tools (graphical displays and test) for analysing phylogenetic, spatial and temporal pattern of one quantitative variable.

Usage

```

orthogram(x, orthobas = NULL, neig = NULL, phylog = NULL,
          nrepet = 999, posinega = 0, tol = 1e-07, na.action = c("fail",
          "mean"), cdot = 1.5, cfont.main = 1.5, lwd = 2, nclass,
          high.scores = 0, alter=c("greater", "less", "two-sided"))

```

Arguments

x	a numeric vector corresponding to the quantitative variable
orthobas	an object of class 'orthobasis'
neig	an object of class 'neig'
phylog	an object of class 'phylog'
nrepet	an integer giving the number of permutations
posinega	a parameter for the ratio test. If $\text{posinega} > 0$, the function computes the ratio test.
tol	a tolerance threshold for orthonormality condition
na.action	if 'fail' stops the execution of the current expression when z contains any missing value. If 'mean' replaces any missing values by $\text{mean}(z)$
cdot	a character size for points on the cumulative decomposition display
cfont.main	a character size for titles
lwd	a character size for dash lines
nclass	a single number giving the number of cells for the histogram
high.scores	a single number giving the number of vectors to return. If > 0 , the function returns labels of vectors that explains the larger part of variance.
alter	a character string specifying the alternative hypothesis, must be one of "greater" (default), "less" or "two-sided"

Details

The function computes the variance decomposition of a quantitative vector x on an orthonormal basis B . The variable is normalized given the uniform weight to eliminate problem of scales. It plots the squared correlations R^2 between x and vectors of B (variance decomposition) and the cumulated squared correlations SR^2 (cumulative decomposition). The function also provides five non parametric tests to test the existence of autocorrelation. The tests derive from the five following statistics :

R2Max $= \max(R^2)$. It takes high value when a high part of the variability is explained by one score.

SkR2k $= \sum_{i=1}^{n-1} (iR_i^2)$. It compares the part of variance explained by internal nodes to the one explained by end nodes.

Dmax $= \max_{m=1, \dots, n-1} (\sum_{j=1}^m R_j^2 - \frac{m}{n-1})$. It examines the accumulation of variance for a sequence of scores.

SCE $= \sum_{m=1}^{n-1} (\sum_{j=1}^m R_j^2 - \frac{m}{n-1})^2$. It examines also the accumulation of variance for a sequence of scores.

ratio depends of the parameter posinega . If $\text{posinega} > 0$, the statistic ratio exists and equals $\sum_{i=1}^{\text{posinega}} R_i^2$. It compares the part of variance explained by internal nodes to the one explained by end nodes when we can define how many vectors correspond to internal nodes.

Value

If (`high.scores = 0`), returns an object of class 'krandtest' (randomization tests) corresponding to the five non parametric tests.

If (`high.scores > 0`), returns a list containing :

`w` : an object of class 'krandtest' (randomization tests)
`scores.order` : a vector which terms give labels of vectors that explain the larger part of variance

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>
 Daniel Chessel

References

Ollier, S., Chessel, D. and Couteron, P. (2005) Orthonormal Transform to Decompose the Variance of a Life-History Trait across a Phylogenetic Tree. *Biometrics*, **62**, 471–477.

See Also

[gridrowcol](#), [orthobasis](#), [mld](#)

Examples

```
# a phylogenetic example
data(ungulates)
ung.phy <- newick2phylog(ungulates$tre)
FemBodyMass <- log(ungulates$tab[,1])
NeonatBodyMass <- log((ungulates$tab[,2]+ungulates$tab[,3])/2)
plot(FemBodyMass,NeonatBodyMass, pch = 20, cex = 2)
abline(lm(NeonatBodyMass~FemBodyMass))
z <- residuals(lm(NeonatBodyMass~FemBodyMass))
dotchart.phylog(ung.phy, val = z, clabel.n = 1,
  labels.n = ung.phy$Blabels, cle = 1.5, cdot = 2)
table.phylog(ung.phy$Bscores, ung.phy, clabel.n = 1,
  labels.n = ung.phy$Blabels)
orthogram(z, ung.phy$Bscores)
orthogram(z, phyl=ung.phy) # the same thing

# a spatial example
data(irishdata)
neig1 <- neig(mat01 = 1*(irishdata$link > 0))
sco1 <- scores.neig(neig1)
z <- scalewt(irishdata$tab$cow)
orthogram(z, sco1)

# a temporal example
data(arrival)
w <- orthobasis.circ(24)
```

```

orthogram(arrival$hours, w)
par(mfrow = c(1,2))
dotcircle(arrival$hours)
dotcircle(w[,2])
par(mfrow = c(1,1))

data(lynx)
ortho <- orthobasis.line(114)
orthogram(lynx,ortho)
attributes(lynx)$tsp
par(mfrow = c(2,1))
par(mar = c(4,4,2,2))
plot.ts(lynx)
plot(ts(ortho[,23], start = 1821, end = 1934, freq = 1), ylab = "score 23")
par(mfrow = c(1,1))

```

ours

A table of Qualitative Variables

Description

The ours (bears) data frame has 38 rows, areas of the "Inventaire National Forestier", and 10 columns.

Usage

```
data(ours)
```

Format

This data frame contains the following columns:

1. altit: importance of the altitudinal area inhabited by bears, a factor with levels:
 - 1 less than 50% of the area between 800 and 2000 meters
 - 2 between 50 and 70%
 - 3 more than 70%
2. deniv: importance of the average variation in level by square of 50 km², a factor with levels:
 - 1 less than 700m
 - 2 between 700 and 900 m
 - 3 more than 900 m
3. cloiso: partitioning of the massif, a factor with levels:
 - 1 a great valley or a ridge isolates at least a quarter of the massif
 - 2 less than a quarter of the massif is isolated
 - 3 the massif has no split
4. domain: importance of the national forests on contact with the massif, a factor with levels:
 - 1 less than 400 km²

- 2 between 400 and 1000 km²
 - 3 more than 1000 km²
5. boise: rate of afforestation, a factor with levels:
 - 1 less than 30%
 - 2 between 30 and 50%
 - 3 more than 50%
 6. hetra: importance of plantations and mixed forests, a factor with levels:
 - 1 less than 5%
 - 2 between 5 and 10%
 - 3 more than 10% of the massif
 7. favor: importance of favorable forests, plantations, mixed forests, fir plantations, a factor with levels:
 - 1 less than 5%
 - 2 between 5 and 10%
 - 3 more than 10% of the massif
 8. inexp: importance of unworked forests, a factor with levels:
 - 1 less than 4%
 - 2 between 4 and 8%
 - 3 more than 8% of the total area
 9. citat: presence of the bear before its disappearance, a factor with levels:
 - 1 no quotation since 1840
 - 2 1 to 3 quotations before 1900 and none after
 - 3 4 quotations before 1900 and none after
 - 4 at least 4 quotations before 1900 and at least 1 quotation between 1900 and 1940
 10. depart: district, a factor with levels:
 - AHP Alpes-de-Haute-Provence
 - AM Alpes-Maritimes
 - D Drôme
 - HP Hautes-Alpes
 - HS Haute-Savoie
 - I Isère
 - S Savoie

Source

Erome, G. (1989) *L'ours brun dans les Alpes françaises. Historique de sa disparition*. Centre Ornithologique Rhône-Alpes, Villeurbanne. 120 p.

Examples

```
data(ours)
boxplot(dudi.acm(ours, scan = FALSE))
```


palm

*Phylogenetic and quantitative traits of amazonian palm trees***Description**

This data set describes the phylogeny of 66 amazonian palm trees. It also gives 7 traits corresponding to these 66 species.

Usage

```
data(palm)
```

Format

`palm` is a list containing the 2 following objects:

tre is a character string giving the phylogenetic tree in Newick format.

traits is a data frame with 66 species (rows) and 7 traits (columns).

Details

Variables of `palm$traits` are the following ones:

rord: specific richness with five ordered levels

h: height in meter (squared transform)

dqual: diameter at breast height in centimeter with five levels sout : subterranean, d1(0, 5 cm), d2(5, 15 cm), d3(15, 30 cm) and d4(30, 100 cm)

vfruit: fruit volume in mm^3 (logged transform)

vgrain: seed volume in mm^3 (logged transform)

aire: spatial distribution area (km^2)

alti: maximum altitude in meter (logged transform)

Source

This data set was obtained by Clémentine Gimaret-Carpentier
<gimaret@biomserv.univ-lyon1.fr>.

Examples

```
## Not run:
data(palm)
palm.phy <- newick2phylog(palm$tre)
radial.phylog(palm.phy, clabel.l=1.25)

orthogram(palm$traits[,4], palm.phy$Bscores)
dotchart.phylog(palm.phy, palm$traits[,4], clabel.l = 1,
  labels.n = palm.phy$Blabels, clabel.n = 0.75)
w <- cbind.data.frame(palm.phy$Bscores[,c(3,4,6,13,21)],
```

```

scalewt((palm$traits[,4]))
names(w)[6] <- names(palm$traits[4])
table.phylog(w, palm.phy, clabel.r = 0.75, f = 0.5)

gearymoran(palm.phy$Amat, palm$traits[,-c(1,3)])

## End(Not run)

```

pap

Taxonomy and quantitative traits of carnivora

Description

This data set describes the taxonomy of 39 carnivora. It also gives life-history traits corresponding to these 39 species.

Usage

```
data(pap)
```

Format

pap is a list containing the 2 following objects :

taxo is a data frame with 39 species and 3 columns.

tab is a data frame with 39 species and 4 traits.

Details

Variables of pap\$tab are the following ones : genre (genus with 30 levels), famille (family with 6 levels), superfamille (superfamily with 2 levels).

Variables of pap\$taxo are Group Size, Body Weight, Brain Weight, Litter Size.

Source

Data taken from the phylogenetic autocorrelation package starting from:
<http://www.ent.orst.edu/pa/demopa.asp>.

Examples

```

data(pap)
taxo <- taxo2phylog(as.taxo(pap$taxo))
table.phylog(as.data.frame(scalewt(pap$tab)), taxo, csi = 2, clabel.nod = 0.6,
f.phylog = 0.6)

```

pcaiv

*Principal component analysis with respect to instrumental variables***Description**

performs a principal component analysis with respect to instrumental variables.

Usage

```
pcaiv(dudi, df, scannf = TRUE, nf = 2)
## S3 method for class 'pcaiv'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'pcaiv'
print(x, ...)
## S3 method for class 'pcaiv'
summary(object, ...)
```

Arguments

dudi	a duality diagram, object of class dudi
df	a data frame with the same rows
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
x, object	an object of class pcaiv
xax	the column number for the x-axis
yax	the column number for the y-axis
...	further arguments passed to or from other methods

Value

returns an object of class pcaiv, sub-class of class dudi

tab	a data frame with the modified array (projected variables)
cw	a numeric vector with the column weights (from dudi)
lw	a numeric vector with the row weights (from dudi)
eig	a vector with the all eigenvalues
rank	an integer indicating the rank of the studied matrix
nf	an integer indicating the number of kept axes
c1	a data frame with the Pseudo Principal Axes (PPA)
li	a data frame dudi\$ls with the predicted values by X
co	a data frame with the inner products between the CPC and Y

l1	data frame with the Constraint Principal Components (CPC)
call	the matched call
X	a data frame with the explanatory variables
Y	a data frame with the dependant variables
ls	a data frame with the projections of lines of <code>dudi\$tab</code> on PPA
param	a table containing information about contributions of the analyses : absolute (1) and cumulative (2) contributions of the decomposition of inertia of the <code>dudi</code> object, absolute (3) and cumulative (4) variances of the projections, the ration (5) between the cumulative variances of the projections (4) and the cumulative contributions (2), the square coefficient of correlation (6) and the eigenvalues of the <code>pcaiv</code> (7)
as	a data frame with the Principal axes of <code>dudi\$tab</code> on PPA
fa	a data frame with the loadings (Constraint Principal Components as linear combinations of X)
cor	a data frame with the correlations between the CPC and X

Author(s)

Daniel Chessel
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 Stephane Dray <stephane.dray@univ-lyon1.fr>

References

- Rao, C. R. (1964) The use and interpretation of principal component analysis in applied research. *Sankhya*, **A 26**, 329–359.
- Obadia, J. (1978) L'analyse en composantes explicatives. *Revue de Statistique Appliquee*, **24**, 5–28.
- Lebreton, J. D., Sabatier, R., Banco G. and Bacou A. M. (1991) Principal component and correspondence analyses with respect to instrumental variables : an overview of their role in studies of structure-activity and species- environment relationships. In J. Devillers and W. Karcher, editors. *Applied Multivariate Analysis in SAR and Environmental Studies*, Kluwer Academic Publishers, 85–114.

Examples

```
data(rhone)
pca1 <- dudi.pca(rhone$tab, scan = FALSE, nf = 3)
iv1 <- pcaiv(pca1, rhone$disch, scan = FALSE)
summary(iv1)
plot(iv1)
```

pcaivortho	<i>Principal Component Analysis with respect to orthogonal instrumental variables</i>
------------	---

Description

performs a Principal Component Analysis with respect to orthogonal instrumental variables.

Usage

```
pcaivortho(dudi, df, scannf = TRUE, nf = 2)
## S3 method for class 'pcaivortho'
summary(object, ...)
```

Arguments

dudi	a duality diagram, object of class dudi
df	a data frame with the same rows
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
object	an object of class pcaiv
...	further arguments passed to or from other methods

Value

an object of class 'pcaivortho' sub-class of class dudi

rank	an integer indicating the rank of the studied matrix
nf	an integer indicating the number of kept axes
eig	a vector with the all eigenvalues
lw	a numeric vector with the row weigths (from dudi)
cw	a numeric vector with the column weigths (from dudi)
Y	a data frame with the dependant variables
X	a data frame with the explanatory variables
tab	a data frame with the modified array (projected variables)
c1	a data frame with the Pseudo Principal Axes (PPA)
as	a data frame with the Principal axis of dudi\$tab on PAP
ls	a data frame with the projection of lines of dudi\$tab on PPA
li	a data frame dudi\$ls with the predicted values by X
l1	a data frame with the Constraint Principal Components (CPC)
co	a data frame with the inner product between the CPC and Y
param	a data frame containing a summary

Author(s)

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 Stephane Dray <stephane.dray@univ-lyon1.fr>

References

Rao, C. R. (1964) The use and interpretation of principal component analysis in applied research. *Sankhya*, **A 26**, 329–359.

Sabatier, R., Lebreton J. D. and Chessel D. (1989) Principal component analysis with instrumental variables as a tool for modelling composition data. In R. Coppi and S. Bolasco, editors. *Multiway data analysis*, Elsevier Science Publishers B.V., North-Holland, 341–352

Examples

```
## Not run:
par(mfrow = c(2,2))
data(avimedi)
cla <- avimedi$plan$reg:avimedi$plan$str

# simple ordination
coa1 <- dudi.coa(avimedi$fau, scan = FALSE, nf = 3)
s.class(coa1$li, cla, sub = "Sans contrainte")

# within region
w1 <- wca(coa1, avimedi$plan$reg, scan = FALSE)
s.match(w1$li, w1$ls, clab = 0, sub = "Intra Région")
s.class(w1$li, cla, add.plot = TRUE)

# no region the same result
pcaivnonA <- pcaivortho(coa1, avimedi$plan$reg, scan = FALSE)
summary(pcaivnonA)

s.match(pcaivnonA$li, pcaivnonA$ls, clab = 0,
        sub = "Contrainte Non A")
s.class(pcaivnonA$li, cla, add.plot = TRUE)

# region + strate
interAplusB <- pcaiv(coa1, avimedi$plan, scan = FALSE)
s.match(interAplusB$li, interAplusB$ls, clab = 0,
        sub = "Contrainte A + B")
s.class(interAplusB$li, cla, add.plot = TRUE)

par(mfrow = c(1,1))
## End(Not run)
```

Description

performs a simplified analysis in principal coordinates, using an object of class `dist`.

Usage

```
pcoscaled(distmat, tol = 1e-07)
```

Arguments

<code>distmat</code>	an object of class <code>dist</code>
<code>tol</code>	a tolerance threshold, an eigenvalue is considered as positive if it is larger than $-\text{tol} \times \text{lambda1}$ where <code>lambda1</code> is the largest eigenvalue

Value

returns a data frame containing the Euclidean representation of the distance matrix with a total inertia equal to 1

Author(s)

Daniel Chessel

References

Gower, J. C. (1966) Some distance properties of latent root and vector methods used in multivariate analysis. *Biometrika*, **53**, 325–338.

Examples

```
a <- 1 / sqrt(3) - 0.2
w <- matrix(c(0,0.8,0.8,a,0.8,0,0.8,a,
             0.8,0.8,0,a,a,a,a,0),4,4)
w <- as.dist(w)
w <- cailliez(w)
w
pcoscaled(w)
dist(pcoscaled(w)) # w
dist(pcoscaled(2 * w)) # the same
sum(pcoscaled(w)^2) # unity
```

perthi02

*Contingency Table with a partition in Molecular Biology***Description**

This data set gives the amino acids of 904 proteins distributed in three classes.

Usage

```
data(perthi02)
```

Format

perthi02 is a list of 2 components.

tab is a data frame 904 rows (proteins of 201 species) 20 columns (amino acids).

cla is a factor of 3 classes of protein

The levels of perthi02\$cla are cyto (cytoplasmic proteins) memb (integral membran proteins) peri (periplasmic proteins)

Source

Perriere, G. and Thioulouse, J. (2002) Use of Correspondence Discriminant Analysis to predict the subcellular location of bacterial proteins. *Computer Methods and Programs in Biomedicine*, **70**, 2, 99–105.

Examples

```
data(perthi02)
plot(discrimin.coa(perthi02$tab, perthi02$cla, scan = FALSE))
```

phylog

*Phylogeny***Description**

Create and use objects of class phylog.

phylog.extract returns objects of class phylog. It extracts sub-trees from a tree.

phylog.permut returns objects of class phylog. It creates the different representations compatible with tree topology.

Usage

```
## S3 method for class 'phylog'
print(x, ...)
phylog.extract(phylog, node, distance = TRUE)
phylog.permut(phylog, list.nodes = NULL, distance = TRUE)
```


Arguments

<code>x, phylog</code>	: an object of class <code>phylog</code>
<code>...</code>	: further arguments passed to or from other methods
<code>node</code>	: a string of characters giving a node name. The functions extracts the tree rooted at this node.
<code>distance</code>	: if TRUE, both functions retain branch lengths. If FALSE, they returns tree with arbitrary branch lengths (each branch length equals one)
<code>list.nodes</code>	: a list which elements are vectors of string of character corresponding to direct descendants of nodes. This list defines one representation compatible with tree topology among the set of possibilities.

Value

Returns a list of class `phylog` :

<code>tre</code>	: a character string of the phylogenetic tree in Newick format without branch length values
<code>leaves</code>	: a vector which names corresponds to leaves and values gives the distance between leaves and nodes closest to these leaves
<code>nodes</code>	: a vector which names corresponds to nodes and values gives the distance between nodes and nodes closest to these leaves
<code>parts</code>	: a list which elements gives the direct descendants of each nodes
<code>paths</code>	: a list which elements gives the path leading from the root to taxonomic units (leaves and nodes)
<code>droot</code>	: a vector which names corresponds to taxonomic units and values gives distance between taxonomic units and the root
<code>call</code>	: call
<code>Wmat</code>	: a phylogenetic link matrix, generally called the covariance matrix. Matrix values $Wmat_{ij}$ correspond to path length that lead from root to the first common ancestor of the two leaves <i>i</i> and <i>j</i>
<code>Wdist</code>	: a phylogenetic distance matrix of class 'dist'. Matrix values $Wdist_{ij}$ correspond to $\sqrt{d_{ij}}$ where d_{ij} is the classical distance between two leaves <i>i</i> and <i>j</i>
<code>Wvalues</code>	: a vector with the eigen values of <code>Wmat</code>
<code>Wscores</code>	: a data frame with eigen vectors of <code>Wmat</code> . This data frame defines an orthobasis that could be used to calculate the orthonormal decomposition of a biological trait on a tree.
<code>Amat</code>	: a phylogenetic link matrix stemed from Abouheif's test and defined in Ollier et al. (submitted)
<code>Avalues</code>	: a vector with the eigen values of <code>Amat</code>
<code>Adim</code>	: number of positive eigen values
<code>Ascores</code>	: a data frame with eigen vectors of <code>Amat</code> . This data frame defines an orthobasis that could be used to calculate the orthonormal decomposition of a biological trait on a tree.

Aparam	: a data frame with attributes associated to nodes.
Bindica	: a data frame giving for some taxonomic units the partition of leaves that is associated to its
Bscores	: a data frame giving an orthobasis defined by Ollier et al. (submitted) that could be used to calculate the orthonormal decomposition of a biological trait on a tree.
Bvalues	: a vector giving the degree of phylogenetic autocorrelation for each vectors of Bscores (Moran's form calculated with the matrix Wmat)
Blabels	: a vector giving for each nodes the name of the vector of Bscores that is associated to its

Author(s)

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References

Ollier, S., Couteron, P. and Chessel, D. (2005) Orthonormal transforms to detect and describe phylogenetic autocorrelation. *Biometrics* (in press).

See Also

[newick2phylog](#), [plot.phylog](#)

Examples

```
marthans.tre <- NULL
marthans.tre[1] <- "(((1:4,2:4)a:5,(3:7,4:7)b:2)c:2,5:11)d:2,"
marthans.tre[2] <- "((6:5,7:5)e:4,(8:4,9:4)f:5)g:4);"
marthans.phylog <- newick2phylog(marthans.tre)
marthans.phylog

if (require(ape, quietly=TRUE)) {
  marthans.phylo <- read.tree(text = marthans.tre)
  marthans.phylo

  par(mfrow = c(1,2))
  plot(marthans.phylog, cnode = 3, f = 0.8, cle = 3)
  plot(marthans.phylo)
  par(mfrow = c(1,1))
}
```

PI2newick

Import data files from Phylogenetic Independance Package

Description

This function ensures to transform a data set written for the Phylogenetic Independance package of Abouheif (1999) in a data set formatting for the functions of ade4.

Usage

```
PI2newick(x)
```

Arguments

`x` is a data frame that contains information on phylogeny topology and trait values

Value

Returns a list containing :

`tre` : a character string giving the phylogenetic tree in Newick format

`trait` : a vector containing values of the trait

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>
Daniel Chessel

References

Abouheif, E. (1999) A method for testing the assumption of phylogenetic independence in comparative data. *Evolutionary Ecology Research*, **1**, 895–909.

Examples

```
x <- c(2.0266, 0.5832, 0.2460, 1.2963, 0.2460, 0.1565, -99.0000,
      -99.0000, 10.1000, -99.0000, 20.2000, 28.2000, -99.0000,
      14.1000, 11.2000, -99.0000, 21.3000, 27.5000, 1.0000, 2.0000,
      -1.0000, 4.0000, -1.0000, -1.0000, 3.0000, -1.0000, -1.0000,
      5.0000, -1.0000, -1.0000, 0.0000, 0.0000, 0.0000, 0.0000,
      0.0000, 0.0000)
x <- matrix(x, nrow = 6)
x <- as.data.frame(x)
res <- PI2newick(x)
dotchart.phylog(newick2phylog(res$tre), res$trait)
```

piosphere

Plant traits response to grazing

Description

Plant species cover, traits and environmental parameters recorded around livestock watering points in different habitats of central Namibian farmlands. See the Wesuls et al. (2012) paper for a full description of the data set.

Usage

```
data(piosphere)
```

Format

`piosphere` is a list of 4 components.

`veg` is a data frame containing plant species cover

`traits` is a data frame with plant traits

`env` is a data frame with environmental variables

`habitat` is a factor describing habitat/years for each site

Source

Wesuls, D., Oldeland, J. and Dray, S. (2012) Disentangling plant trait responses to livestock grazing from spatio-temporal variation: the partial RLQ approach. *Journal of Vegetation Science*, **23**, 98–113.

Examples

```
data(piosphere)
names(piosphere)
afcL <- dudi.coa(log(piosphere$veg + 1), scannf = FALSE)
acpR <- dudi.pca(piosphere$env, scannf = FALSE, row.w = afcL$lw)
acpQ <- dudi.hillsmith(piosphere$traits, scannf = FALSE, row.w = afcL$cw)
rlq1 <- rlq(acpR, afcL, acpQ, scannf = FALSE)
plot(rlq1)
```

plot.phylog

*Plot phylogenies***Description**

plot.phylog draws phylogenetic trees as linear dendograms.
 radial.phylog draws phylogenetic trees as circular dendograms.
 enum.phylog enumerate all the possible representations for a phylogeny.

Usage

```
## S3 method for class 'phylog'
plot(x, y = NULL, f.phylog = 0.5, cleaves = 1, cnodes = 0,
     labels.leaves = names(x$leaves), clabel.leaves = 1,
     labels.nodes = names(x$nodes), clabel.nodes = 0, sub = "",
     csub = 1.25, possub = "bottomleft", draw.box = FALSE, ...)
radial.phylog(phylog, circle = 1, cleaves = 1, cnodes = 0,
             labels.leaves = names(phylog$leaves), clabel.leaves = 1,
             labels.nodes = names(phylog$nodes), clabel.nodes = 0,
             draw.box = FALSE)
enum.phylog(phylog, no.over = 1000)
```

Arguments

x, phylog	an object of class phylog
y	a vector which values correspond to leaves positions
f.phylog	a size coefficient for tree size (a parameter to draw the tree in proportion to leaves label)
circle	a size coefficient for the outer circle
cleaves	a character size for plotting the points that represent the leaves, used with par("cex")*cleaves. If zero, no points are drawn
cnodes	a character size for plotting the points that represent the nodes, used with par("cex")*cnodes. If zero, no points are drawn
labels.leaves	a vector of strings of characters for the leaves labels
clabel.leaves	a character size for the leaves labels, used with par("cex")*clabel.leaves. If zero, no leaves labels are drawn
labels.nodes	a vector of strings of characters for the nodes labels
clabel.nodes	a character size for the nodes labels, used with par("cex")*clabel.nodes. If zero, no nodes labels are drawn
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with par("cex")*csub
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")

draw.box	if TRUE draws a box around the current plot with the function box()
...	further arguments passed to or from other methods
no.over	a size coefficient for the number of representations

Details

The vector `y` is an argument of the function `plot.phylog` that ensures to plot one of the possible representations of a phylogeny. The vector `y` is a permutation of the set of leaves $\{1,2,\dots,f\}$ compatible with the phylogeny's topology.

Value

The function `enum.phylog` returns a matrix with as many columns as leaves. Each row gives a permutation of the set of leaves $\{1,2,\dots,f\}$ compatible with the phylogeny's topology.

Author(s)

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See Also

[phylog](#)

Examples

```
data(newick.eg)
par(mfrow = c(3,2))
for(i in 1:6) plot(newick2phylog(newick.eg[[i]], FALSE),
  clea = 2, clabel.l = 3, cnod = 2.5)
par(mfrow = c(1,1))

## Not run:
par(mfrow = c(1,2))
plot(newick2phylog(newick.eg[[11]], FALSE), clea = 1.5,
  clabel.l = 1.5, clabel.nod = 0.75, f = 0.8)
plot(newick2phylog(newick.eg[[10]], FALSE), clabel.l = 0,
  clea = 0, cn = 0, f = 1)
par(mfrow = c(1,1))

## End(Not run)

par(mfrow = c(2,2))
w7 <- newick2phylog("((((((1,2,3)b),(6)c),(4,5)d,7)f);)")
plot(w7,clabel.l = 1.5, clabel.n = 1.5, f = 0.8, cle = 2,
  cnod = 3, sub = "((((((1,2,3)b),(6)c),(4,5)d,7)f);", csub = 2)
w <- NULL
w[1] <- "(((e1:4,e2:4)a:5,(e3:7,e4:7)b:2)c:2,e5:11)d:2,"
w[2] <- "((e6:5,e7:5)e:4,(e8:4,e9:4)f:5)g:4);"
plot(newick2phylog(w), f = 0.8, cnod = 2, cleav = 2, clabel.l = 2)
```

```

data(taxo.eg)
w <- taxo2phylog(as.taxo(taxo.eg[[1]]))
plot(w, clabel.lea = 1.25, clabel.n = 1.25, sub = "Taxonomy",
     csub = 3, f = 0.8, possub = "topleft")

provi.tre <- "(((a,b,c,d,e)A,(f,g,h)B)C)D;"
provi.phy <- newick2phylog(provi.tre)
plot(provi.phy, clabel.l = 2, clabel.n = 2, f = 0.8)
par(mfrow = c(1,1))

## Not run:
par(mfrow = c(3,3))
for (j in 1:6) radial.phylog(newick2phylog(newick.eg[[j]],
    FALSE), clabel.l = 2, cnodes = 2)
radial.phylog(newick2phylog(newick.eg[[7]],FALSE), clabel.l = 2)
radial.phylog(newick2phylog(newick.eg[[8]],FALSE), clabel.l = 0,
    circle = 1.8)
radial.phylog(newick2phylog(newick.eg[[9]],FALSE), clabel.l = 1,
    clabel.n = 1, cle = 0, cnode = 1)
par(mfrow = c(1,1))

data(bsetal97)
bsetal.phy = taxo2phylog(as.taxo(bsetal97$taxo[,1:3]), FALSE)
radial.phylog(bsetal.phy, cnod = 1, clea = 1, clabel.l = 0.75,
    draw.box = TRUE, cir = 1.1)
par(mfrow = c(1,1))

## End(Not run)

## Not run:
# plot all the possible representations of a phylogenetic tree
a <- "((a,b)A,(c,d,(e,f)B)C)D;"
wa <- newick2phylog(a)
wx <- enum.phylog(wa)
dim(wx)

par(mfrow = c(6,8))
fun <- function(x) {
  w <-NULL
  lapply(x, function(y) w<<-paste(w,as.character(y),sep=""))
  plot(wa, x, clabel.n = 1.25, f = 0.75, clabel.l = 2,
    box = FALSE, cle = 1.5, sub = w, csub = 2)
  invisible()}
apply(wx,1,fun)
par(mfrow = c(1,1))

## End(Not run)

```

Description

presid2002 is a list of two data frames `tour1` and `tour2` with 93 rows (93 departments from continental Metropolitan France) and, 4 and 12 variables respectively .

Usage

```
data(presid2002)
```

Format

`tour1` contains the following arguments:

the number of registered voters (`inscrits`); the number of abstentions (`abstentions`); the number of voters (`votes`); the number of expressed votes (`exprimes`) and, the numbers of votes for each candidate: Megret, Lepage, Gluksten, Bayrou, Chirac, Le_Pen, Taubira, Saint.josse, Mamere, Jospin, Boutin, Hue, Chevenement, Madelin, Besancenot.

`tour2` contains the following arguments:

the number of registered voters (`inscrits`); the number of abstentions (`abstentions`); the number of voters (`votes`); the number of expressed votes (`exprimes`) and, the numbers of votes for each candidate: Chirac and Le_Pen.

Source

Site of the ministry of the Interior, of the Internal Security and of the local liberties

<http://www.interieur.gouv.fr/avotreservice/elections/presid2002/>

See Also

This dataset is compatible with `elec88` and `cnc2003`

Examples

```
data(presid2002)
all((presid2002$tour2$Chirac + presid2002$tour2$Le_Pen) == presid2002$tour2$exprimes)
## Not run:
data(elec88)
data(cnc2003)
w1 = area.util.class(elec88$area, cnc2003$reg)

par(mfrow = c(2,2))
par(mar = c(0.1,0.1,0.1,0.1))

area.plot(w1)
w = scale(elec88$tab$Chirac)
s.value(elec88$xy, w, add.plot = TRUE)
scatterutil.sub("Chirac 1988 T1", csub = 2, "topleft")

area.plot(w1)
w = scale(presid2002$tour1$Chirac/ presid2002$tour1$exprimes)
s.value(elec88$xy, w, add.plot = TRUE)
scatterutil.sub("Chirac 2002 T1", csub = 2, "topleft")
```



```
area.plot(w1)
w = scale(elec88$tab$Mitterand)
s.value(elec88$xy, w, add.plot = TRUE)
scatterutil.sub("Mitterand 1988 T1", csub = 2, "topleft")

area.plot(w1)
w = scale(presid2002$tour2$Chirac/ presid2002$tour2$exprimes)
s.value(elec88$xy, w, add.plot = TRUE)
scatterutil.sub("Chirac 2002 T2", csub = 2, "topleft")

## End(Not run)
```

procella

Phylogeny and quantitative traits of birds

Description

This data set describes the phylogeny of 19 birds as reported by Bried et al. (2002). It also gives 6 traits corresponding to these 19 species.

Usage

```
data(procella)
```

Format

procella is a list containing the 2 following objects:

tre is a character string giving the phylogenetic tree in Newick format.

traits is a data frame with 19 species and 6 traits

Details

Variables of procella\$traits are the following ones:

site.fid: a numeric vector that describes the percentage of site fidelity

mate.fid: a numeric vector that describes the percentage of mate fidelity

mass: an integer vector that describes the adult body weight (g)

ALE: a numeric vector that describes the adult life expectancy (years)

BF: a numeric vector that describes the breeding frequencies

col.size: an integer vector that describes the colony size (no nests monitored)

References

Bried, J., Pontier, D. and Jouventin, P. (2002) Mate fidelity in monogamous birds: a re-examination of the Procellariiformes. *Animal Behaviour*, **65**, 235–246.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps037.pdf> (in French).

Examples

```

data(procella)
pro.phy <- newick2phylog(procella$tre)
plot(pro.phy, clabel.n = 1, clabel.l = 1)
wt <- procella$traits
wt$site.fid[is.na(wt$site.fid)] <- mean(wt$site.fid[!is.na(wt$site.fid)])
wt$site.fid <- asin(sqrt(wt$site.fid/100))
wt$ALE[is.na(wt$ALE)] <- mean(wt$ALE[!is.na(wt$ALE)])
wt$ALE <- sqrt(wt$ALE)
wt$BF[is.na(wt$BF)] <- mean(wt$BF[!is.na(wt$BF)])
wt$mass <- log(wt$mass)
wt <- wt[, -6]
table.phylog(scalewt(wt), pro.phy, csi = 2)
gearymoran(pro.phy$Amat, wt, 9999)

```

procuste

Simple Procruste Rotation between two sets of points

Description

performs a simple procruste rotation between two sets of points.

Usage

```

procuste(df1, df2, scale = TRUE, nf = 4, tol = 1e-07)
## S3 method for class 'procuste'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'procuste'
print(x, ...)

```

Arguments

df1, df2	two data frames with the same rows
scale	a logical value indicating whether a transformation by the Gower's scaling (1971) should be applied
nf	an integer indicating the number of kept axes
tol	a tolerance threshold to test whether the distance matrix is Euclidean : an eigenvalue is considered positive if it is larger than $-tol * \lambda_1$ where λ_1 is the largest eigenvalue.
x	an objet of class procuste
xax	the column number for the x-axis
yax	the column number for the y-axis
...	further arguments passed to or from other methods

Value

returns a list of the class `procuste` with 9 components

<code>d</code>	a numeric vector of the singular values
<code>rank</code>	an integer indicating the rank of the crossed matrix
<code>nf</code>	an integer indicating the number of kept axes
<code>tab1</code>	a data frame with the array 1, possibly scaled
<code>tab2</code>	a data frame with the array 2, possibly scaled
<code>rot1</code>	a data frame with the result of the rotation from array 1 to array 2
<code>rot2</code>	a data frame with the result of the rotation from array 2 to array 1
<code>load1</code>	a data frame with the loadings of array 1
<code>load2</code>	a data frame with the loadings of array 2
<code>scor1</code>	a data frame with the scores of array 1
<code>scor2</code>	a data frame with the scores of array 2
<code>call</code>	a call order of the analysis

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

- Digby, P. G. N. and Kempton, R. A. (1987) *Multivariate Analysis of Ecological Communities. Population and Community Biology Series*, Chapman and Hall, London.
- Gower, J.C. (1971) Statistical methods of comparing different multivariate analyses of the same data. In *Mathematics in the archaeological and historical sciences*, Hodson, F.R, Kendall, D.G. & Tautu, P. (Eds.) University Press, Edinburgh, 138–149.
- Schönemann, P.H. (1968) On two-sided Procustes problems. *Psychometrika*, **33**, 19–34.
- Torre, F. and Chessel, D. (1994) Co-structure de deux tableaux totalement appariés. *Revue de Statistique Appliquée*, **43**, 109–121.
- Dray, S., Chessel, D. and Thioulouse, J. (2003) Procustean co-inertia analysis for the linking of multivariate datasets. *Ecoscience*, **10**, 1, 110-119.

Examples

```
data(macaca)
par(mfrow = c(2,2))
pro1 <- procuste(macaca$xy1, macaca$xy2, scal = FALSE)
s.match(pro1$tab1, pro1$rot2, clab = 0.7)
s.match(pro1$tab2, pro1$rot1, clab = 0.7)
pro2 <- procuste(macaca$xy1, macaca$xy2)
```

```
s.match(pro2$stab1, pro2$rot2, clab = 0.7)
s.match(pro2$stab2, pro2$rot1, clab = 0.7)
par(mfrow = c(1,1))

data(doubs)
pca1 <- dudi.pca(doubs$env, scal = TRUE, scann = FALSE)
pca2 <- dudi.pca(doubs$fish, scal = FALSE, scann = FALSE)
pro3 <- procuste(pca1$stab, pca2$stab, nf = 2)
par(mfrow = c(2,2))
s.traject(pro3$scor1, clab = 0)
s.label(pro3$scor1, clab = 0.8, add.p = TRUE)
s.traject(pro3$scor2, clab = 0)
s.label(pro3$scor2, clab = 0.8, add.p = TRUE)
s.arrow(pro3$load1, clab = 0.75)
s.arrow(pro3$load2, clab = 0.75)
plot(pro3)
par(mfrow = c(1,1))

data(fruits)
plot(procuste(scalewt(fruits$jug), scalewt(fruits$var)))
```

procuste.randtest	<i>Monte-Carlo Test on the sum of the singular values of a procustean rotation (in C).</i>
-------------------	--

Description

performs a Monte-Carlo Test on the sum of the singular values of a procustean rotation.

Usage

```
procuste.randtest(df1, df2, nrepet = 999)
```

Arguments

df1	a data frame
df2	a data frame
nrepet	the number of permutations

Value

returns a list of class randtest

Author(s)

Jean Thioulouse <ade4-jt@biomserv.univ-lyon1.fr>

References

Jackson, D.A. (1995) PROTEST: a PROcustean randomization TEST of community environment concordance. *Ecosciences*, **2**, 297–303.

Examples

```
data(doubs)
pca1 <- dudi.pca(doubs$env, scal = TRUE, scann = FALSE)
pca2 <- dudi.pca(doubs$fish, scal = FALSE, scann = FALSE)
protest1 <- procuste.randtest(pca1$tab, pca2$tab, 999)
protest1
plot(protest1, main="PROTEST")
```

procuste.rtest	<i>Monte-Carlo Test on the sum of the singular values of a procustean rotation (in R).</i>
----------------	--

Description

performs a Monte-Carlo Test on the sum of the singular values of a procustean rotation.

Usage

```
procuste.rtest(df1, df2, nrepet = 99)
```

Arguments

df1	a data frame
df2	a data frame
nrepet	the number of permutations

Value

returns a list of class `rtest`

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Jackson, D.A. (1995) PROTEST: a PROcustean randomization TEST of community environment concordance. *Ecosciences*, **2**, 297–303.

Examples

```

data(doubs)
pca1 <- dudi.pca(doubs$env, scal = TRUE, scann = FALSE)
pca2 <- dudi.pca(doubs$fish, scal = FALSE, scann = FALSE)
proc1 <- procuste(pca1$stab, pca2$stab)
protest1 <- procuste.rtest(pca1$stab, pca2$stab, 999)
protest1
plot(protest1)

```

pta

*Partial Triadic Analysis of a K-tables***Description**

performs a partial triadic analysis of a K-tables, using an object of class ktab.

Usage

```

pta(X, scannf = TRUE, nf = 2)
## S3 method for class 'pta'
plot(x, xax = 1, yax = 2, option = 1:4, ...)
## S3 method for class 'pta'
print(x, ...)

```

Arguments

X	an object of class ktab where the arrays have 1) the same dimensions 2) the same names for columns 3) the same column weightings
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
x	an object of class 'pta'
xax, yax	the numbers of the x-axis and the y-axis
option	an integer between 1 and 4, otherwise the 4 components of the plot are displayed
...	further arguments passed to or from other methods

Value

returns a list of class 'pta', sub-class of 'dudi' containing :

RV	a matrix with the all RV coefficients
RV.eig	a numeric vector with the all eigenvalues (interstructure)
RV.coo	a data frame with the scores of the arrays
tab.names	a vector of characters with the array names
nf	an integer indicating the number of kept axes

rank	an integer indicating the rank of the studied matrix
tabw	a numeric vector with the array weights
cw	a numeric vector with the column weights
lw	a numeric vector with the row weights
eig	a numeric vector with the all eigenvalues (compromis)
cos2	a numeric vector with the \cos^2 between compromise and arrays
tab	a data frame with the modified array
li	a data frame with the row coordinates
l1	a data frame with the row normed scores
co	a data frame with the column coordinates
c1	a data frame with the column normed scores
Tli	a data frame with the row coordinates (each table)
Tco	a data frame with the column coordinates (each table)
Tcomp	a data frame with the principal components (each table)
Tax	a data frame with the principal axes (each table)
TL	a data frame with the factors for Tli
TC	a data frame with the factors for Tco
T4	a data frame with the factors for Tax and Tcomp

Author(s)

Pierre Bady <pierre.bady@univ-lyon1.fr>
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Blanc, L., Chessel, D. and Dolédec, S. (1998) Etude de la stabilité temporelle des structures spatiales par Analyse d'une série de tableaux faunistiques totalement appariés. *Bulletin Français de la Pêche et de la Pisciculture*, **348**, 1–21.

Thioulouse, J., and D. Chessel. 1987. Les analyses multi-tableaux en écologie factorielle. I De la typologie d'état à la typologie de fonctionnement par l'analyse triadique. *Acta Oecologica, Oecologia Generalis*, **8**, 463–480.

Examples

```
data(meaudret)
wit1 <- withinpca(meaudret$env, meaudret$design$season, scan = FALSE,
  scal = "partial")
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5"), 4))
kta2 <- t(kta1)
pta1 <- pta(kta2, scann = FALSE)
pta1
plot(pta1)
```

`quasieuclid`*Transformation of a distance matrice to a Euclidean one*

Description

transforms a distance matrix in a Euclidean one.

Usage

```
quasieuclid(distmat)
```

Arguments

`distmat` an object of class `dist`

Details

The function creates a distance matrice with the positive eigenvalues of the Euclidean representation.

Only for Euclidean distances which are not Euclidean for numeric approximations (for examples, in papers as the following example).

Value

object of class `dist` containing a Euclidean distance matrice

Author(s)

Daniel Chessel
Stéphane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
data(yanomama)
geo <- as.dist(yanomama$geo)
is.euclid(geo) # FALSE
geo1 <- quasieuclid(geo)
is.euclid(geo1) # TRUE
par(mfrow = c(2,2))
lapply(yanomama, function(x) plot(as.dist(x), quasieuclid(as.dist(x))))

par(mfrow = c(1,1))
```

randEH	<i>Nee and May's random process</i>
--------	-------------------------------------

Description

When branch lengths in an ultrametric phylogenetic tree are expressed as divergence times, the total sum of branch lengths in that tree expresses the amount of evolutionary history. The function randEH calculates the amount of evolutionary history preserved when k random species out of n original species are saved.

Usage

```
randEH(phy1, nbofsp, nbrep = 10)
```

Arguments

phy1	an object of class phylog
nbofsp	an integer indicating the number of species saved (k).
nbrep	an integer indicating the number of random sampling.

Value

Returns a numeric vector

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Nee, S. and May, R.M. (1997) Extinction and the loss of evolutionary history. *Science* **278**, 692–694.

Pavoine, S., Ollier, S. and Dufour, A.-B. (2005) Is the originality of a species measurable? *Ecology Letters*, **8**, 579–586.

See Also

[optimEH](#)

Examples

```
data(carni70)
carni70.phy <- newick2phylog(carni70$tre)
mean(randEH(carni70.phy, nbofsp = 7, nbrep = 1000))

## Not run:
# the following instructions can last about 2 minutes.
data(carni70)
```

```

carni70.phy <- newick2phylog(carni70$tre)
percent <- c(0,0.04,0.07,seq(0.1,1,by=0.1))
pres <- round(percent*70)
topt <- sapply(pres, function(i) optimEH(carni70.phy, nbofsp = i, give = F))
topt <- topt / EH(carni70.phy)
tsam <- sapply(pres, function(i) mean(randEH(carni70.phy, nbofsp = i, nbrep = 1000)))
tsam <- tsam / EH(carni70.phy)
plot(pres, topt, xlab = "nb of species saved", ylab = "Evolutionary history saved", type = "l")
lines(pres, tsam)

## End(Not run)

```

randtest

Class of the Permutation Tests (in C).

Description

randtest is a generic function. It proposes methods for the following objects between, discrimin, coinertia ...

Usage

```

randtest(xtest, ...)
## S3 method for class 'randtest'
plot(x, nclass = 10, coeff = 1, ...)
as.randtest(sim, obs, alter=c("greater", "less", "two-sided"), call = match.call())
## S3 method for class 'randtest'
print(x, ...)

```

Arguments

xtest	an object used to select a method
x	an object of class randtest
...	... further arguments passed to or from other methods; in plot.randtest to hist
nclass	a number of intervals for the histogram
coeff	to fit the magnitude of the graph
sim	a numeric vector of simulated values
obs	a numeric vector of an observed value
alter	a character string specifying the alternative hypothesis, must be one of "greater" (default), "less" or "two-sided"
call	a call order

Details

If the alternative hypothesis is "greater", a p-value is estimated as: (number of random values equal to or greater than the observed one + 1)/(number of permutations + 1). The null hypothesis is rejected if the p-value is less than the significance level. If the alternative hypothesis is "less", a p-value is estimated as: (number of random values equal to or less than the observed one + 1)/(number of permutations + 1). Again, the null hypothesis is rejected if the p-value is less than the significance level. Lastly, if the alternative hypothesis is "two-sided", the estimation of the p-value is equivalent to the one used for "greater" except that random and observed values are firstly centered (using the average of random values) and secondly transformed to their absolute values. Note that this is only suitable for symmetric random distribution.

Value

`as.randtest` returns a list of class `randtest`
`plot.randtest` draws the simulated values histograms and the position of the observed value

See Also

[mantel.randtest](#), [procuste.randtest](#), [rtest](#)

Examples

```
par(mfrow = c(2,2))
for (x0 in c(2.4,3.4,5.4,20.4)) {
  l0 <- as.randtest(sim = rnorm(200), obs = x0)
  print(l0)
  plot(l0,main=paste("p.value = ", round(l0$pvalue, dig = 5)))
}
par(mfrow = c(1,1))
```

randtest.amova

Permutation tests on an analysis of molecular variance (in C).

Description

Tests the components of covariance with permutation processes described by Excoffier et al. (1992).

Usage

```
## S3 method for class 'amova'
randtest(xtest, nrepet = 99, ...)
```

Arguments

<code>xtest</code>	an object of class <code>amova</code>
<code>nrepet</code>	the number of permutations
<code>...</code>	further arguments passed to or from other methods

Value

returns an object of class krandtest or randtest

Author(s)

Sandrine Pavoine <pavoine@biomserv.univ-lyon1.fr>

References

Excoffier, L., Smouse, P.E. and Quattro, J.M. (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics*, **131**, 479–491.

Examples

```
data(humDNAm)
amovahum <- amova(humDNAm$samples, sqrt(humDNAm$distances), humDNAm$structures)
amovahum
randtesthum <- randtest(amovahum, 49)
plot(randtesthum)
```

randtest.between *Monte-Carlo Test on the between-groups inertia percentage (in C).*

Description

Performs a Monte-Carlo test on the between-groups inertia percentage.

Usage

```
## S3 method for class 'between'
randtest(xtest, nrepet = 999, ...)
```

Arguments

xtest	an object of class between
nrepet	the number of permutations
...	further arguments passed to or from other methods

Value

a list of the class randtest

Author(s)

Jean Thioulouse <ade4-jt@biomserv.univ-lyon1.fr>

References

Romesburg, H. C. (1985) Exploring, confirming and randomization tests. *Computers and Geosciences*, **11**, 19–37.

Examples

```
data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 3)
rand1 <- randtest(bca(pca1, meaudret$design$season, scan = FALSE), 99)
rand1
plot(rand1, main = "Monte-Carlo test")
```

randtest.coinertia *Monte-Carlo test on a Co-inertia analysis (in C).*

Description

Performs a Monte-Carlo test on a Co-inertia analysis.

Usage

```
## S3 method for class 'coinertia'
randtest(xtest, nrepet = 999, fixed=0, ...)
```

Arguments

xtest	an object of class coinertia
nrepet	the number of permutations
fixed	when non uniform row weights are used in the coinertia analysis, this parameter must be the number of the table that should be kept fixed in the permutations
...	further arguments passed to or from other methods

Value

a list of the class randtest

Note

A testing procedure based on the total coinertia of the analysis is available by the function `randtest.coinertia`. The function allows to deal with various analyses for the two tables. The test is based on random permutations of the rows of the two tables. If the row weights are not uniform, mean and variances are recomputed for each permutation (PCA); for MCA, tables are recentred and column weights are recomputed. If weights are computed using the data contained in one table (e.g. COA), you must fix this table and permute only the rows of the other table. The case of decentred PCA (PCA where centers are entered by the user) is not yet implemented. If you want to use the testing procedure for this case, you must firstly center the table and then perform a non-centered PCA on the modified table. The case where one table is treated by hill-smith analysis (mix of quantitative and qualitative variables) will be soon implemented.

Author(s)

Jean Thioulouse <ade4-jt@biomserv.univ-lyon1.fr> modified by Stephane Dray <dray@biomserv.univ-lyon1.fr>

References

Doledec, S. and Chessel, D. (1994) Co-inertia analysis: an alternative method for studying species-environment relationships. *Freshwater Biology*, **31**, 277–294.

Examples

```
data(doubs)
dudi1 <- dudi.pca(doubs$env, scale = TRUE, scan = FALSE, nf = 3)
dudi2 <- dudi.pca(doubs$fish, scale = FALSE, scan = FALSE, nf = 2)
coin1 <- coinertia(dudi1,dudi2, scan = FALSE, nf = 2)
plot(randtest(coin1))
```

randtest.discrimin *Monte-Carlo Test on a Discriminant Analysis (in C).*

Description

Test of the sum of a discriminant analysis eigenvalues (divided by the rank). Non parametric version of the Pillai’s test. It authorizes any weighting.

Usage

```
## S3 method for class 'discrimin'
randtest(xtest, nrepet = 999, ...)
```

Arguments

xtest	an object of class discrimin
nrepet	the number of permutations
...	further arguments passed to or from other methods

Value

returns a list of class randtest

Author(s)

Jean Thioulouse <ade4-jt@biomserv.univ-lyon1.fr>

Examples

```

data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 3)
rand1 <- randtest(discrimin(pca1, meaudret$design$season, scan = FALSE), 99)
rand1
#Monte-Carlo test
#Observation: 0.3035
#Call: as.randtest(sim = sim, obs = obs)
#Based on 999 replicates
#Simulated p-value: 0.001
plot(rand1, main = "Monte-Carlo test")
summary.manova(manova(as.matrix(meaudret$env)~meaudret$design$season), "Pillai")
#           Df Pillai approx F num Df den Df Pr(>F)
# meaudret$design$season 3  2.73  11.30  27  30 1.6e-09 ***
# Residuals           16
# ---
# Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
# 2.731/9 = 0.3034

```

randtest.pcaiv	<i>Monte-Carlo Test on the percentage of explained (i.e. constrained) inertia</i>
----------------	---

Description

Performs a Monte-Carlo test on on the percentage of explained (i.e. constrained) inertia. The statistic is the ratio of the inertia (sum of eigenvalues) of the constrained analysis divided by the inertia of the unconstrained analysis.

Usage

```

## S3 method for class 'pcaiv'
randtest(xtest, nrepet = 99, ...)
## S3 method for class 'cca'
randtest(xtest, nrepet = 99, ...)
## S3 method for class 'pcaivortho'
randtest(xtest, nrepet = 99, ...)

```

Arguments

xtest	an object of class pcaiv, pcaivortho or cca
nrepet	the number of permutations
...	further arguments passed to or from other methods

Value

a list of the class randtest

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>, original code by Raphael Pelissier

Examples

```
data(rpjd1)
millog <- log(rpjd1$mil + 1)
iv1 <- cca(rpjd1$fau, millog, scan = FALSE)
randtest(iv1)
```

rankrock

Ordination Table

Description

This data set gives the classification in order of preference of 10 music groups by 51 students.

Usage

```
data(rankrock)
```

Format

A data frame with 10 rows and 51 columns.

Each column contains the rank (1 for the favorite, ..., 10 for the less appreciated) attributed to the group by a student.

Examples

```
data(rankrock)
dudi1 <- dudi.pca(rankrock, scannf = FALSE, nf = 3)
scatter(dudi1, clab.r = 1.5)
```

reconst

Reconstitution of Data from a Duality Diagram

Description

Generic Function for the reconstitution of data from a principal component analysis or a correspondence analysis

Usage

```
reconst (dudi, ...)
## S3 method for class 'pca'
reconst(dudi, nf = 1, ...)
## S3 method for class 'coa'
reconst(dudi, nf = 1, ...)
```


Arguments

dudi an object of class dudi used to select a method: pca or coa
 nf an integer indicating the number of kept axes for the reconstitution
 ... further arguments passed to or from other methods

Value

returns a data frame containing the reconstituted data

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Gabriel, K.R. (1978) Least-squares approximation of matrices by additive and multiplicative models. *Journal of the Royal Statistical Society, B* , **40**, 186–196.

Examples

```
data(rhone)
dd1 <- dudi.pca(rhone$tab, nf = 2, scann = FALSE)
rh1 <- reconst(dd1, 1)
rh2 <- reconst(dd1, 2)
par(mfrow = c(4,4))
par(mar = c(2.6,2.6,1.1,1.1))
for (i in 1:15) {
  plot(rhone$date, rhone$tab[,i])
  lines(rhone$date, rh1[,i], lty = 2)
  lines(rhone$date, rh2[,i], lty = 1)
  ade4:::scatterutil.sub(names(rhone$tab)[i], 2, "topright")
}

data(chats)
chatsw <- data.frame(t(chats))
chatscoa <- dudi.coa(chatsw, scann = FALSE)
model0 <- reconst(chatscoa, 0)
round(model0,3)
round(chisq.test(chatsw)$expected,3)
chisq.test(chatsw)$statistic
sum(((chatsw-model0)^2)/model0)
effectif <- sum(chatsw)
sum(chatscoa$eig)*effectif
model1 <- reconst(chatscoa, 1)
round(model1, 3)
sum(((chatsw-model1)^2)/model0)
sum(chatscoa$eig[-1])*effectif
```

rhizobium

Genetic structure of two nitrogen fixing bacteria influenced by geographical isolation and host specialization

Description

The data set concerns fixing bacteria belonging to the genus *Sinorhizobium* (Rhizobiaceae) associated with the plant genus *Medicago* (Fabaceae). It is a combination of two data sets fully available online from GenBank and published in two recent papers (see reference below). The complete sampling procedure is described in the Additional file 3 of the reference below. We delineated six populations according to geographical origin (France: F, Tunisia Hadjeb: TH, Tunisia Enfidha: TE), the host plant (*M. truncatula* or similar symbiotic specificity: T, *M. laciniata*: L), and the taxonomical status of bacteria (*S. meliloti*: mlt, *S. medicae*: mdc). Each population will be called hereafter according to the three above criteria, e.g. THLmlt is the population sampled in Tunisia at Hadjeb from *M. laciniata* nodules which include *S. meliloti* isolates. *S. medicae* interacts with *M. truncatula* while *S. meliloti* interacts with both *M. laciniata* (*S. meliloti* bv. *medicaginis*) and *M. truncatula* (*S. meliloti* bv. *meliloti*). The numbers of individuals are respectively 46 for FTmdc, 43 for FTmlt, 20 for TETmdc, 24 for TETmlt, 20 for TELmlt, 42 for THTmlt and 20 for THLmlt.

Four different intergenic spacers (IGS), IGSNOD, IGSEXO, IGSGAB, and IGSRKP, distributed on the different replication units of the model strain 1021 of *S. meliloti* bv. *meliloti* had been sequenced to characterize each bacterial isolate (DNA extraction and sequencing procedures are described in an additional file). It is noteworthy that the IGSNOD marker is located within the nod gene cluster and that specific alleles at these loci determine the ability of *S. meliloti* strains to interact with either *M. laciniata* or *M. truncatula*.

Usage

`data(rhizobium)`

Format

rhizobium is a list of 2 components.

- dnaobj: list of dna lists. Each dna list corresponds to a locus. For a given locus, the dna list provides the dna sequences. The *i*th sequences of all loci corresponds to the *i*th individual of the data set.
- pop: The list of the populations which each individual sequence belongs to.

Source

Pavoine, S. and Bailly, X. (2007) New analysis for consistency among markers in the study of genetic diversity: development and application to the description of bacterial diversity. *BMC Evolutionary Biology*, **7**, e156.

Examples

```
# The functions used below require the package ape
data(rhizobium)
if (require(ape, quiet = TRUE)) {
  dat <- prep.mdpcoa(rhizobium[[1]], rhizobium[[2]],
    model = c("F84", "F84", "F84", "F81"),
    pairwise.deletion = TRUE)
  sam <- dat$sam
  dis <- dat$dis
  # The distances should be Euclidean.
  # Several transformations exist to render a distance object Euclidean
  # (see functions cailliez, lingoies and quasieulid in the ade4 package).
  # Here we use the quasieulid function.
  dis <- lapply(dis, quasieulid)
  mdpcoa1 <- mdpcoa(sam, dis, scann = FALSE, nf = 2)

  # Reference analysis
  plot(mdpcoa1)

  # Differences between the loci
  kplot(mdpcoa1)

  # Alleles projected on the population maps.
  kplotX.mdpcoa(mdpcoa1)
}
```

rhone

Physico-Chemistry Data

Description

This data set gives for 39 water samples a physico-chemical description with the number of sample date and the flows of three tributaries.

Usage

```
data(rhone)
```

Format

rhone is a list of 3 components.

tab is a data frame with 39 water samples and 15 physico-chemical variables.

date is a vector of the sample date (in days).

disch is a data frame with 39 water samples and the flows of the three tributaries.

Source

Carrel, G., Barthelemy, D., Auda, Y. and Chessel, D. (1986) Approche graphique de l'analyse en composantes principales normée : utilisation en hydrobiologie. *Acta Oecologica, Oecologia Generalis*, **7**, 189–203.

Examples

```
data(rhone)
pca1 <- dudi.pca(rhone$stab, nf = 2, scann = FALSE)
rh1 <- reconst(pca1, 1)
rh2 <- reconst(pca1, 2)
par(mfrow = c(4,4))
par(mar = c(2.6,2.6,1.1,1.1))
for (i in 1:15) {
  plot(rhone$date, rhone$stab[,i])
  lines(rhone$date, rh1[,i], lwd = 2)
  lines(rhone$date, rh2[,i])
  ade4:::scatterutil.sub(names(rhone$stab)[i], 2, "topright")
}
par(mfrow = c(1,1))
```

rlq

RLQ analysis

Description

RLQ analysis performs a double inertia analysis of two arrays (R and Q) with a link expressed by a contingency table (L). The rows of L correspond to the rows of R and the columns of L correspond to the rows of Q.

Usage

```
rlq(dudiR, dudiL, dudiQ, scannf = TRUE, nf = 2)
## S3 method for class 'rlq'
print(x, ...)
## S3 method for class 'rlq'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'rlq'
summary(object, ...)
## S3 method for class 'rlq'
randtest(xtest, nrepet = 999, modeltype = 6, ...)
```

Arguments

dudiR a duality diagram providing from one of the functions dudi.hillsmith, dudi.pca,
 ...

dudiL a duality diagram of the function dudi.coa

dudiQ	a duality diagram providing from one of the functions dudi.hillsmith, dudi.pca, ...
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
x	an rlq object
xax	the column number for the x-axis
yax	the column number for the y-axis
object	an rlq object
xtest	an rlq object
nrepet	the number of permutations
modeltype	the model used to permute data(2: permute rows of R, 4: permute rows of Q, 5: permute both, 6: sequential approach, see ter Braak et al. 2012)
...	further arguments passed to or from other methods

Value

Returns a list of class 'dudi', sub-class 'rlq' containing:

call	call
rank	rank
nf	a numeric value indicating the number of kept axes
RV	a numeric value, the RV coefficient
eig	a numeric vector with all the eigenvalues
lw	a numeric vector with the rows weights (crossed array)
cw	a numeric vector with the columns weights (crossed array)
tab	a crossed array (CA)
li	R col = CA row: coordinates
l1	R col = CA row: normed scores
co	Q col = CA column: coordinates
c1	Q col = CA column: normed scores
lR	the row coordinates (R)
mR	the normed row scores (R)
lQ	the row coordinates (Q)
mQ	the normed row scores (Q)
aR	the axis onto co-inertia axis (R)
aQ	the axis onto co-inertia axis (Q)

WARNING

IMPORTANT : row weights for dudiR and dudiQ must be taken from dudiL.

Note

A testing procedure based on the total coinertia of the RLQ analysis is available by the function `randtest.rlq`. The function allows to deal with various analyses for tables R and Q. Means and variances are recomputed for each permutation (PCA); for MCA, tables are recentred and column weights are recomputed. The case of decentred PCA (PCA where centers are entered by the user) for R or Q is not yet implemented. If you want to use the testing procedure for this case, you must firstly center the table and then perform a non-centered PCA on the modified table.

Author(s)

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References

- Doledec, S., Chessel, D., ter Braak, C.J.F. and Champely, S. (1996) Matching species traits to environmental variables: a new three-table ordination method. *Environmental and Ecological Statistics*, **3**, 143–166.
- Dray, S., Pettorelli, N., Chessel, D. (2002) Matching data sets from two different spatial samplings. *Journal of Vegetation Science*, **13**, 867–874.
- Dray, S. and Legendre, P. (2008) Testing the species traits-environment relationships: the fourth-corner problem revisited. *Ecology*, **89**, 3400–3412.
- ter Braak, C., Cormont, A., Dray, S. (2012) Improved testing of species traits-environment relationships in the fourth corner problem. *Ecology*, **93**, 1525–1526.

See Also

[coinertia](#), [fourthcorner](#)

Examples

```
data(aviurba)
coa1 <- dudi.coa(aviurba$fau, scannf = FALSE, nf = 2)
dudimil <- dudi.hillsmith(aviurba$mil, scannf = FALSE, nf = 2, row.w = coa1$lw)
duditrait <- dudi.hillsmith(aviurba$traits, scannf = FALSE, nf = 2, row.w = coa1$cw)
rlq1 <- rlq(dudimil, coa1, duditrait, scannf = FALSE, nf = 2)
plot(rlq1)
summary(rlq1)
randtest(rlq1)
fourthcorner.rlq(rlq1, type="Q.axes")
fourthcorner.rlq(rlq1, type="R.axes")
```

rpjdl

*Avifauna and Vegetation***Description**

This data set gives the abundance of 52 species and 8 environmental variables in 182 sites.

Usage

```
data(rpjdl)
```

Format

rpjdl is a list of 5 components.

fau is the faunistic array of 182 sites (rows) and 52 species (columns).

mil is the array of environmental variables : 182 sites and 8 variables.

frlab is a vector of the names of species in French.

lalab is a vector of the names of species in Latin.

lab is a vector of the simplified labels of species.

Source

Prodon, R. and Lebreton, J.D. (1981) Breeding avifauna of a Mediterranean succession : the holm oak and cork oak series in the eastern Pyrénées. 1 : Analysis and modelling of the structure gradient. *Oikos*, **37**, 21–38.

Lebreton, J. D., Chessel D., Prodon R. and Yoccoz N. (1988) L'analyse des relations espèces-milieu par l'analyse canonique des correspondances. I. Variables de milieu quantitatives. *Acta Oecologica, Oecologia Generalis*, **9**, 53–67.

References

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps048.pdf> (in French).

Examples

```
## Not run:
data(rpjdl)
xy <- dudi.coa(rpjdl$fau, scann = FALSE)$l1
s.distri(xy, rpjdl$fau, 2, 1, cstar = 0.3, cell = 0)

xy1 <- dudi.pca(rpjdl$fau, scal = FALSE, scann = FALSE)$l1
s.distri(xy1, rpjdl$fau, 2, 1, cstar = 0.3, cell = 0)

cca1 <- cca(rpjdl$fau, rpjdl$mil, scan = FALSE)
plot(cca1)

## End(Not run)
```

`rtest`*Class of the Permutation Tests (in R).*

Description

`rtest` is a generic function. It proposes methods for the following objects `between`, `discrimin`, `procuste` ...

Usage

```
rtest(xtest, ...)  
## S3 method for class 'rtest'  
plot(x, nclass = 10, coeff = 1, ...)  
  as.rtest (sim, obs, call = match.call())  
## S3 method for class 'rtest'  
print(x, ...)
```

Arguments

<code>xtest</code>	an object used to select a method
<code>x</code>	an object of class 'rtest'
<code>...</code>	further arguments passed to or from other methods; in <code>plot.randtest</code> to <code>hist</code>
<code>nclass</code>	a number of intervals for the histogram
<code>coeff</code>	to fit the magnitude of the graph
<code>sim</code>	a numeric vector of simulated values
<code>obs</code>	a numeric vector of an observed value
<code>call</code>	a call order

Value

`as.rtest` returns a list of class `rtest`
`plot.rtest` draws the simulated values histograms and the position of the observed value

Author(s)

Daniel Chessel

See Also

[RV.rtest](#), [mantel.rtest](#), [procuste.rtest](#), [randtest](#)

Examples

```

par(mfrow = c(2,2))
for (x0 in c(2.4,3.4,5.4,20.4)) {
  l0 <- as.rtest(sim = rnorm(200), obs = x0)
  print(l0)
  plot(l0,main=paste("p.value = ", round(l0$pvalue, dig = 5)))
}

```

rtest.between	<i>Monte-Carlo Test on the between-groups inertia percentage (in R).</i>
---------------	--

Description

Performs a Monte-Carlo test on the between-groups inertia percentage.

Usage

```

## S3 method for class 'between'
rtest(xtest, nrepet = 99, ...)

```

Arguments

xtest	an object of class between
nrepet	the number of permutations
...	further arguments passed to or from other methods

Value

a list of the class rtest

Author(s)

Daniel Chessel

References

Romesburg, H. C. (1985) Exploring, confirming and randomization tests. *Computers and Geosciences*, **11**, 19–37.

Examples

```

data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 3)
rand1 <- rtest(bca(pca1, meaudret$design$season, scan = FALSE), 99)
rand1
plot(rand1, main = "Monte-Carlo test")

```

rtest.discrimin	<i>Monte-Carlo Test on a Discriminant Analysis (in R).</i>
-----------------	--

Description

Test of the sum of a discriminant analysis eigenvalues (divided by the rank). Non parametric version of the Pillai's test. It authorizes any weighting.

Usage

```
## S3 method for class 'discrimin'
rtest(xtest, nrepet = 99, ...)
```

Arguments

xtest	an object of class discrimin
nrepet	the number of permutations
...	further arguments passed to or from other methods

Value

returns a list of class rtest

Author(s)

Daniel Chessel

Examples

```
data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 3)
rand1 <- rtest(discrimin(pca1, meaudret$design$season, scan = FALSE), 99)
rand1
#Monte-Carlo test
#Observation: 0.3035
#Call: as.rtest(sim = sim, obs = obs)
#Based on 999 replicates
#Simulated p-value: 0.001
plot(rand1, main = "Monte-Carlo test")
summary.manova(manova(as.matrix(meaudret$env)~meaudret$design$season), "Pillai")
#           Df Pillai approx F num Df den Df Pr(>F)
# meaudret$design$season 3  2.73  11.30   27   30 1.6e-09 ***
# Residuals           16
# ---
# Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
# 2.731/9 = 0.3034
```

RV.rtest	<i>Monte-Carlo Test on the sum of eigenvalues of a co-inertia analysis (in R).</i>
----------	--

Description

performs a Monte-Carlo Test on the sum of eigenvalues of a co-inertia analysis.

Usage

```
RV.rtest(df1, df2, nrepet = 99)
```

Arguments

df1, df2	two data frames with the same rows
nrepet	the number of permutations

Value

returns a list of class 'rtest'

Author(s)

Daniel Chessel

References

Heo, M. & Gabriel, K.R. (1997) A permutation test of association between configurations by means of the RV coefficient. *Communications in Statistics - Simulation and Computation*, **27**, 843-856.

Examples

```
data(doubs)
pca1 <- dudi.pca(doubs$env, scal = TRUE, scann = FALSE)
pca2 <- dudi.pca(doubs$fish, scal = FALSE, scann = FALSE)
rv1 <- RV.rtest(pca1$stab, pca2$stab, 99)
rv1
plot(rv1)
```

RVdist.randtest	<i>Tests of randomization on the correlation between two distance matrices (in R).</i>
-----------------	--

Description

performs a RV Test between two distance matrices.

Usage

```
RVdist.randtest(m1, m2, nrepet = 999)
```

Arguments

m1, m2	two Euclidean matrices
nrepet	the number of permutations

Value

returns a list of class 'randtest'

Author(s)

Daniel Chessel

References

Heo, M. & Gabriel, K.R. (1997) A permutation test of association between configurations by means of the RV coefficient. *Communications in Statistics - Simulation and Computation*, **27**, 843-856.

s.arrow	<i>Plot of the factorial maps for the projection of a vector basis</i>
---------	--

Description

performs the scatter diagrams of the projection of a vector basis.

Usage

```
s.arrow(dfxy, xax = 1, yax = 2, label = row.names(dfxy),
        clabel = 1, pch = 20, cpoint = 0, boxes = TRUE, edge = TRUE, origin = c(0,0),
        xlim = NULL, ylim = NULL, grid = TRUE, addaxes = TRUE, cgrid = 1,
        sub = "", csub = 1.25, possub = "bottomleft", pixmap = NULL,
        contour = NULL, area = NULL, add.plot = FALSE)
```

Arguments

dfxy	a data frame containing the two columns for the axes
xax	the column number of x in dfxy
yax	the column number of y in dfxy
label	a vector of strings of characters for the point labels
clabel	if not NULL, a character size for the labels used with <code>par("cex")*clabel</code>
pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn.
boxes	if TRUE, labels are framed
edge	a logical value indicating whether the arrows should be plotted
origin	the fixed point in the graph space, by default <code>c(0,0)</code> the origin of axes. The arrows begin at cent.
xlim	the ranges to be encompassed by the x-axis, if NULL they are computed
ylim	the ranges to be encompassed by the y-axis, if NULL they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> , to indicate the mesh of the grid
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the legend position ("topleft", "topright", "bottomleft", "bottomright")
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
s.arrow(cbind.data.frame(runif(55,-2,3), runif(55,-3,2)))
```

s.chull

Plot of the factorial maps with polygons of contour by level of a factor

Description

performs the scatter diagrams with polygons of contour by level of a factor.

Usage

```
s.chull(dfxy, fac, xax = 1, yax = 2,
        optchull = c(0.25, 0.5, 0.75, 1), label = levels(fac), clabel = 1,
        cpoint = 0, col = rep(1, length(levels(fac))), xlim = NULL, ylim = NULL,
        grid = TRUE, addaxes = TRUE, origin = c(0,0), include.origin = TRUE,
        sub = "", csub = 1, possub = "bottomleft", cgrid = 1, pixmap = NULL,
        contour = NULL, area = NULL, add.plot = FALSE)
```

Arguments

dfxy	a data frame containing the two columns for the axes
fac	a factor partitioning the rows of the data frame in classes
xax	the column number of x in dfxy
yax	the column number of y in dfxy
optchull	the number of convex hulls and their interval
label	a vector of strings of characters for the point labels
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
col	a vector of colors used to draw each class in a different color
xlim	the ranges to be encompassed by the x axis, if NULL, they are computed
ylim	the ranges to be encompassed by the y axis, if NULL they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid

pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
xy <- cbind.data.frame(x = runif(200,-1,1), y = runif(200,-1,1))
posi <- factor(xy$x > 0) : factor(xy$y > 0)
coul <- c("black", "red", "green", "blue")
s.chull(xy, posi, cpoi = 1.5, col = coul)
```

s.class

Plot of factorial maps with representation of point classes

Description

performs the scatter diagrams with representation of point classes.

Usage

```
s.class(dfxy, fac, wt = rep(1, length(fac)), xax = 1,
  yax = 2, cstar = 1, cellipse = 1.5, axesell = TRUE,
  label = levels(fac), clabel = 1, cpoint = 1, pch = 20,
  col = rep(1, length(levels(fac))), xlim = NULL, ylim = NULL,
  grid = TRUE, addaxes = TRUE, origin = c(0,0),
  include.origin = TRUE, sub = "", csub = 1, possub = "bottomleft",
  cgrid = 1, pixmap = NULL, contour = NULL, area = NULL, add.plot = FALSE)
```

Arguments

dfxy	a data frame containing the two columns for the axes
fac	a factor partitioning the rows of the data frame in classes
wt	a vector of the point weightings of the data frame used for computing the means (star centers) and the ellipses of dispersion
xax	the column number of x in dfxy
yax	the column number of y in dfxy

cstar	a number between 0 and 1 which defines the length of the star size
cellipse	a positive coefficient for the inertia ellipse size
axesell	a logical value indicating whether the ellipse axes should be drawn
label	a vector of strings of characters for the point labels
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
col	a vector of colors used to draw each class in a different color
xlim	the ranges to be encompassed by the x, if NULL they are computed
ylim	the ranges to be encompassed by the y, if NULL they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position (" <code>opleft</code> ", " <code>opright</code> ", " <code>ottomleft</code> ", " <code>ottomright</code> ")
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```

xy <- cbind.data.frame(x = runif(200,-1,1), y = runif(200,-1,1))
posi <- factor(xy$x > 0) : factor(xy$y > 0)
coul <- c("black", "red", "green", "blue")
par(mfrow = c(2,2))
s.class(xy, posi, cpoi = 2)
s.class(xy, posi, cell = 0, cstar = 0.5)
s.class(xy, posi, cell = 2, axesell = FALSE, csta = 0, col = coul)
s.chull(xy, posi, cpoi = 1)
par(mfrow = c(1,1))

## Not run:
data(banque)
dudi1 <- dudi.acm(banque, scannf = FALSE)
coul = rainbow(length(levels(banque[,20])))
par(mfrow = c(2,2))
s.label(dudi1$li, sub = "Factorial map from ACM", csub = 1.5,
        possub = "topleft")
s.class(dudi1$li, banque[,20], sub = names(banque)[20],
        possub = "bottomright", cell = 0, cstar = 0.5, cgrid = 0, csub = 1.5)
s.class(dudi1$li, banque[,20], csta = 0, cell = 2, cgrid = 0,
        clab = 1.5)
s.class(dudi1$li, banque[,20], sub = names(banque)[20],
        possub = "topright", cgrid = 0, col = coul)
par(mfrow = c(1,1))

par(mfrow = n2mfrow(ncol(banque)))
for (i in 1:(ncol(banque)))
  s.class(dudi1$li, banque[,i], clab = 1.5, sub = names(banque)[i],
          csub = 2, possub = "topleft", cgrid = 0, csta = 0, cpoi = 0)
s.label(dudi1$li, clab = 0, sub = "Common background")
par(mfrow = c(1,1))

## End(Not run)

```

s.corcircle

Plot of the factorial maps of a correlation circle

Description

performs the scatter diagram of a correlation circle.

Usage

```

s.corcircle(dfxy, xax = 1, yax = 2, label = row.names(df),
            clabel = 1, grid = TRUE, sub = "", csub = 1, possub = "bottomleft",
            cgrid = 0, fullcircle = TRUE, box = FALSE, add.plot = FALSE)

```

Arguments

dfxy	a data frame with two coordinates
xax	the column number for the x-axis
yax	the column number for the y-axis
label	a vector of strings of characters for the point labels
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
grid	a logical value indicating whether a grid in the background of the plot should be drawn
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
fullcircle	a logical value indicating whether the complete circle should be drawn
box	a logical value indicating whether a box should be drawn
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
data(olympic)
dudi1 <- dudi.pca(olympic$tab, scan = FALSE) # a normed PCA
par(mfrow = c(2,2))
s.corcircle(dudi1$co, lab = names(olympic$tab))
s.corcircle(dudi1$co, cgrid = 0, full = FALSE, clab = 0.8)
s.corcircle(dudi1$co, lab = as.character(1:11), cgrid = 2,
  full = FALSE, sub = "Correlation circle", csub = 2.5,
  possub = "bottomleft", box = TRUE)
s.arrow(dudi1$co, clab = 1)
```

s.distri *Plot of a frequency distribution*

Description

performs the scatter diagram of a frequency distribution.

Usage

```
s.distri(dfxy, dfdistri, xax = 1, yax = 2, cstar = 1,
         cellipse = 1.5, axesell = TRUE, label = names(dfdistri),
         clabel = 0, cpoint = 1, pch = 20, xlim = NULL, ylim = NULL,
         grid = TRUE, addaxes = TRUE, origin = c(0,0),
         include.origin = TRUE, sub = "", csub = 1, possub = "bottomleft",
         cgrid = 1, pixmap = NULL, contour = NULL, area = NULL, add.plot = FALSE)
```

Arguments

dfxy	a data frame containing two columns for the axes
dfdistri	a data frame containing the mass distributions in columns
xax	the column number for the x-axis
yax	the column number for the y-axis
cstar	a number between 0 and 1 which defines the length of the star size
cellipse	a positive coefficient for the inertia ellipse size
axesell	a logical value indicating whether the ellipse axes should be drawn
label	a vector of strings of characters for the distribution centers labels
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
xlim	the ranges to be encompassed by the x, if NULL they are computed
ylim	the ranges to be encompassed by the y, if NULL they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>

possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
cgrid	a character size, parameter used with <code>par("cex")* cgrid</code> to indicate the mesh of the grid
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
xy <- cbind.data.frame(x = runif(200,-1,1), y = runif(200,-1,1))
distri <- data.frame(w1 = rpois(200, xy$x * (xy$x > 0)))
s.value(xy, distri$w1, cpoi = 1)
s.distri(xy, distri, add.p = TRUE)

w1 <- as.numeric((xy$x > 0) & (xy$y > 0))
w2 <- ((xy$x > 0) & (xy$y < 0)) * (1 - xy$y) * xy$x
w3 <- ((xy$x < 0) & (xy$y > 0)) * (1 - xy$x) * xy$y
w4 <- ((xy$x < 0) & (xy$y < 0)) * xy$y * xy$x

distri <- data.frame(a = w1 / sum(w1), b = w2 / sum(w2),
  c = w3 / sum(w3), d = w4 / sum(w4))
s.value(xy, unlist(apply(distri, 1, sum)), cleg = 0, csi = 0.75)
s.distri(xy, distri, clab = 2, add.p = TRUE)

data(rpjdl)
xy <- dudi.coa(rpjdl$fau, scan = FALSE)$li
par(mfrow = c(3,4))
for (i in c(1,5,8,20,21,23,26,33,36,44,47,49)){
  s.distri(xy, rpjdl$fau[,i], cell = 1.5, sub = rpjdl$frlab[i],
    csub = 2, cgrid = 1.5)}
par(mfrow = c(1,1))
```

`s.hist`*Display of a scatterplot and its two marginal histograms*

Description

performs a scatterplot and the two marginal histograms of each axis.

Usage

```
s.hist(dfxy, xax = 1, yax = 2, cgrid = 1, cbreaks = 2, adjust = 1, ...)
```

Arguments

<code>dfxy</code>	a data frame with two coordinates
<code>xax</code>	column for the x axis
<code>yax</code>	column for the y axis
<code>cgrid</code>	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
<code>cbreaks</code>	a parameter used to define the numbers of cells for the histograms. By default, two cells are defined for each interval of the grid displayed in <code>s.label</code> . With an increase of the integer <code>cbreaks</code> , the number of cells increases as well.
<code>adjust</code>	a parameter passed to <code>density</code> to display a kernel density estimation
<code>...</code>	further arguments passed from the <code>s.label</code> for the scatter plot

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
data(rpjd1)
coa1 <- dudi.coa(rpjd1$fau, scannf = FALSE, nf = 4)
s.hist(coa1$li)
s.hist(coa1$li, cgrid = 2, cbr = 3, adj = 0.5, clab = 0)
s.hist(coa1$co, cgrid = 2, cbr = 3, adj = 0.5, clab = 0)
```

s.image

*Graph of a variable using image and contour***Description**

performs a scatterplot

Usage

```
s.image(dfxy, z, xax = 1, yax = 2, span = 0.5, xlim = NULL, ylim = NULL,
        kgrid = 2, scale = TRUE, grid = FALSE, addaxes = FALSE, cgrid = 0,
        include.origin = FALSE, origin = c(0, 0), sub = "", csub = 1,
        possub = "topleft", neig = NULL, cneig = 1, image.plot = TRUE,
        contour.plot = TRUE, pixmap = NULL, contour = NULL, area = NULL,
        add.plot = FALSE)
```

Arguments

dfxy	a data frame containing the two columns for the axes
z	a vector of values on the dfxy rows
xax	the column number of x in dfxy
yax	the column number of y in dfxy
span	the parameter alpha which controls the degree of smoothing
xlim	the ranges to be encompassed by the x-axis, if NULL they are computed
ylim	the ranges to be encompassed by the y-axis, if NULL they are computed
kgrid	a number of points used to locally estimate the level line through the nodes of the grid, used by <code>kgrid*sqrt(length(z))</code>
scale	if TRUE, data are centered and reduced
grid	if TRUE, the background grid is traced
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
neig	an object of class neig
cneig	a size for the neighbouring graph lines used with <code>par("lwd")*cneig</code>

image.plot	if TRUE, the image is traced
contour.plot	if TRUE, the contour lines are plotted
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```

if (require(splancs, quiet = TRUE)){
  wxy=data.frame(expand.grid(-3:3,-3:3))
  names(wxy)=c("x","y")
  z=(1/sqrt(2))*exp(-(wxy$x^2+wxy$y^2)/2)
  par(mfrow=c(2,2))
  s.value(wxy,z)
  s.image(wxy,z)
  s.image(wxy,z,kgrid=5)
  s.image(wxy,z,kgrid=15)
}

## Not run:
data(t3012)
if (require(splancs, quiet = TRUE)){
  par(mfrow = c(4,4))
  for(k in 1:12) s.image(t3012$xy,scalewt(t3012$temp[,k]), kgrid = 3)
  par(mfrow = c(1,1))
}

data(elec88)
if (require(splancs, quiet = TRUE)){
  par(mfrow = c(4,4))
  for(k in 1:12)
  s.image(t3012$xy, scalewt(t3012$temp[,k]), kgrid = 3, sub = names(t3012$temp)[k],
    csub = 3, area = elec88$area)
  par(mfrow = c(1,1))
}

## End(Not run)

```

s.kde2d

*Scatter Plot with Kernel Density Estimate***Description**

performs a scatter of points without labels by a kernel Density Estimation in One or Two Dimensions

Usage

```
s.kde2d(dfxy, xax = 1, yax = 2, pch = 20, cpoint = 1, neig = NULL, cneig = 2,
        xlim = NULL, ylim = NULL, grid = TRUE, addaxes = TRUE, cgrid = 1,
        include.origin = TRUE, origin = c(0, 0), sub = "", csub = 1.25,
        possub = "bottomleft", pixmap = NULL, contour = NULL,
        area = NULL, add.plot = FALSE)
```

Arguments

dfxy	a data frame with at least two coordinates
xax	the column number for the x-axis
yax	the column number for the y-axis
pch	if cpoint > 0, an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with par("cex")*cpoint. If zero, no points are drawn
neig	a neighbouring graph
cneig	a size for the neighbouring graph lines used with par("lwd")*cneig
xlim	the ranges to be encompassed by the x axis, if NULL, they are computed
ylim	the ranges to be encompassed by the y axis, if NULL, they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with par("cex")* 'cgrid' to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example c(0,0) the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with par("cex")*csub
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
pixmap	an object pixmap displayed in the map background

contour a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)

area a data frame of class 'area' to plot a set of surface units in contour

add.plot if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
# To recognize groups of points
data(casitas)
casitas.fuz = fuzzygenet(casitas)
casitas.pop <- as.factor(rep(c("dome", "cast", "musc", "casi"), c(24,11,9,30)))
casitas.pca = dudi.pca(casitas.fuz, scannf = FALSE, scale = FALSE)
if (require(MASS, quiet=TRUE)) {s.kde2d(casitas.pca$li)
s.class(casitas.pca$li,casitas.pop, cell = 0, add.p = TRUE)
}
```

s.label

Scatter Plot

Description

performs the scatter diagrams with labels.

Usage

```
s.label(dfxy, xax = 1, yax = 2, label = row.names(dfxy),
        clabel = 1, pch = 20, cpoint = if (clabel == 0) 1 else 0, boxes = TRUE,
        neig = NULL, cneig = 2, xlim = NULL, ylim = NULL, grid = TRUE,
        addaxes = TRUE, cgrid = 1, include.origin = TRUE, origin = c(0,0),
        sub = "", csub = 1.25, possub = "bottomleft", pixmap = NULL,
        contour = NULL, area = NULL, add.plot = FALSE)
```

Arguments

dfxy a data frame with at least two coordinates

xax the column number for the x-axis

yax the column number for the y-axis

label a vector of strings of characters for the point labels

clabel if not NULL, a character size for the labels, used with par("cex")*clabel

pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
boxes	if TRUE, labels are framed
neig	a neighbouring graph
cneig	a size for the neighbouring graph lines used with <code>par("lwd")*cneig</code>
xlim	the ranges to be encompassed by the x axis, if NULL, they are computed
ylim	the ranges to be encompassed by the y axis, if NULL, they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position (" <code>opleft</code> ", " <code>opright</code> ", " <code>ottomleft</code> ", " <code>ottomright</code> ")
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (<code>x1,y1,x2,y2</code>)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
layout(matrix(c(1,2,3,2), 2, 2))
data(atlas)
s.label(atlas$xy, lab = atlas$names.district,
        area = atlas$area, inc = FALSE, addax = FALSE)
data(mafragh)
s.label(mafragh$xy, inc = FALSE, neig = mafragh$neig, addax = FALSE)
data(irishdata)
```

```

s.label(irisdata$xy, inc = FALSE, contour = irisdata$contour,
        addax = FALSE)

par(mfrow = c(2,2))
cha <- ls()
s.label(cbind.data.frame(runif(length(cha)),
                        runif(length(cha))), lab = cha)
x <- runif(50,-2,2) ; y <- runif(50,-2,2) ; z <- x^2 + y^2
s.label(data.frame(x,y), lab = as.character(z < 1))
s.label(data.frame(x,y), clab = 0, cpoi = 1, add.plot = TRUE)
symbols(0, 0, circles = 1, add = TRUE, inch = FALSE)
s.label(cbind.data.frame(runif(100,0,10), runif(100,5,12)),
        incl = FALSE, clab = 0)
s.label(cbind.data.frame(runif(100,-3,12),
                        runif(100,2,10)), cl = 0, cp = 2, include = FALSE)

```

s.logo

Representation of an object in a graph by a picture

Description

performs the scatter diagrams using pictures to represent the points

Usage

```

s.logo(dfxy, listlogo, klogo=NULL, clogo=1, rectlogo=TRUE,
       xax = 1, yax = 2, neig = NULL, cneig = 1, xlim = NULL, ylim = NULL,
       grid = TRUE, addaxes = TRUE, cgrid = 1, include.origin = TRUE,
       origin = c(0, 0), sub = "", csub = 1.25, possub = "bottomleft",
       pixmap = NULL, contour = NULL, area = NULL, add.plot = FALSE)

```

Arguments

dfxy	a data frame with at least two coordinates
listlogo	a list of pixmap pictures
klogo	a numeric vector giving the order in which pictures of listlogo are used; if NULL, the order is the same than the rows of dfxy
clogo	a numeric vector giving the size factor applied to each picture
rectlogo	a logical to decide whether a rectangle should be drawn around the picture (TRUE) or not (FALSE)
xax	the column number for the x-axis
yax	the column number for the y-axis
neig	a neighbouring graph
cneig	a size for the neighbouring graph lines used with par("lwd")*cneig
xlim	the ranges to be encompassed by the x axis, if NULL, they are computed

ylim	the ranges to be encompassed by the y axis, if NULL, they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with par("cex")* cgrid to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example c(0,0) the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with par("cex")*csub
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel and Thibaut Jombart <jombart@biomserv.univ-lyon1.fr>

Examples

```

if(require(pixmap, quiet=TRUE)){
  data(ggtortoises)
  a1 <- ggtortoises$area
  area.plot(a1)
  rect(min(a1$x), min(a1$y), max(a1$x), max(a1$y), col = "lightblue")
  invisible(lapply(split(a1, a1$id), function(x) polygon(x[, -1], col = "white")))
  s.label(ggtortoises$misc, grid=FALSE, include.ori=FALSE, addaxes=FALSE, add.p=TRUE)
  listico <- ggtortoises$ico[as.character(ggtortoises$pop$carap)]
  s.logo(ggtortoises$pop, listico, add.p=TRUE)

  data(capitales)
  index <- pmatch(tolower(attr(capitales$dist, "Labels")), names(capitales$logo))
  w1 <- capitales$area
  area.plot(w1)
  rect(min(w1$x), min(w1$y), max(w1$x), max(w1$y), col = "lightblue")
  invisible(lapply(split(w1, w1$id), function(x) polygon(x[, -1], col = "white")))
  s.logo(capitales$xy, capitales$logo, klogo=index, add.plot=TRUE, clogo=1) # depends on pixmap
}

```

s.match

*Plot of Paired Coordinates***Description**

performs the scatter diagram for a paired coordinates.

Usage

```
s.match(df1xy, df2xy, xax = 1, yax = 2, pch = 20, cpoint = 1,
        label = row.names(df1xy), clabel=1, edge = TRUE, xlim = NULL,
        ylim = NULL, grid = TRUE, addaxes = TRUE, cgrid = 1,
        include.origin = TRUE, origin = c(0,0), sub = "", csub = 1.25,
        possub = "bottomleft", pixmap = NULL, contour = NULL, area = NULL,
        add.plot = FALSE)
```

Arguments

df1xy	a data frame containing two columns from the first system
df2xy	a data frame containing two columns from the second system
xax	the column number for the x-axis of both the two systems
yax	the column number for the y-axis of both the two systems
pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
label	a vector of strings of characters for the couple labels
clabel	if not <code>NULL</code> , a character size for the labels, used with <code>par("cex")*clabel</code>
edge	If <code>TRUE</code> the arrows are plotted, otherwise only the segments are drawn
xlim	the ranges to be encompassed by the x axis, if <code>NULL</code> they are computed
ylim	the ranges to be encompassed by the y axis, if <code>NULL</code> they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>

possusb	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
pixmap	aan object pixmap displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
X <- data.frame(x = runif(50,-1,2), y = runif(50,-1,2))
Y <- X + rnorm(100, sd = 0.3)
par(mfrow = c(2,2))
s.match(X, Y)
s.match(X, Y, edge = FALSE, clab = 0)
s.match(X, Y, edge = FALSE, clab = 0)
s.label(X, clab = 1, add.plot = TRUE)
s.label(Y, clab = 0.75, add.plot = TRUE)
s.match(Y, X, clab = 0)
```

s.match.class

Scatterplot of two sets of coordinates and a partitioning into classes

Description

Performs a graphical representation of two sets of coordinates (different colors and symbols) and a partitionning into classes

Usage

```
s.match.class(df1xy, df2xy, fac, wt = rep(1/nrow(df1xy), nrow(df1xy)),
xax = 1, yax = 2, pch1 = 16, pch2 = 15, col1 = rep("lightgrey",
nlevels(fac)), col2 = rep("darkgrey", nlevels(fac)), cpoint = 1, label =
levels(fac), clabel = 1, cstar = 1, cellipse = 0, axesell = TRUE, xlim =
NULL, ylim = NULL, grid = TRUE, addaxes = TRUE, cgrid = 1,
include.origin = TRUE, origin = c(0, 0), sub = "", csub = 1.25, possusb =
"bottomleft", pixmap = NULL, contour = NULL, area = NULL, add.plot = FALSE)
```

Arguments

df1xy	a dataframe with the first system of coordinates
df2xy	a dataframe with the second system of coordinates
fac	a factor partitioning the rows of the data frame in classes
wt	a vector of weights
xax	a number indicating which column should be plotted on the x-axis
yax	a number indicating which column should be plotted on the y-axis
pch1	if cpoint > 0, an integer specifying the symbol or the single character to be used for plotting points
pch2	if cpoint > 0, an integer specifying the symbol or the single character to be used for plotting points
col1	a color for symbols
col2	a color for symbols
cpoint	a character size for plotting the points, used with par("cex")*cpoint. If zero, no points are drawn
label	a vector of strings of characters for the couple labels
clabel	if not NULL, a character size for the labels, used with par("cex")*clabel
cstar	a number between 0 and 1 which defines the length of the star size
cellipse	a positive coefficient for the inertia ellipse size
axesell	a logical value indicating whether the ellipse axes should be drawn
xlim	the ranges to be encompassed by the x axis, if NULL they are computed
ylim	the ranges to be encompassed by the y axis, if NULL they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
cgrid	a character size, parameter used with par("cex")*cgrid to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should belong to the graph space
origin	a fixed point in the graph space, for example c(0,0) for the origin of axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with par("cex")*csub
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
pixmap	a pixmap object
contour	a dataframe with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a dataframe of class 'area' to plot an areal map
add.plot	if TRUE, add the plot to the current graphic device

Value

The matched call.

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>

See Also

[s.class](#), [s.match](#)

Examples

```
xy <- data.frame(matrix(rnorm(100),50,2))
xy[,1] <- xy[,1] + rep(seq(0,12,by=3),rep(10,5))
xy[,2] <- xy[,2] + rep(seq(0,12,by=3),rep(10,5))
fac <- gl(5,10)
xy2<-xy + matrix(rnorm(100),50,2) + 1
s.match.class(xy,xy2,fac)
```

s.multinom

Graph of frequency profiles (useful for instance in genetic)

Description

The main purpose of this function is to draw categories using scores and profiles by their gravity center. Confidence intervals of the average position (issued from a multinomial distribution) can be superimposed.

Usage

```
s.multinom(dfxy, dfrowprof, translate = FALSE, xax = 1, yax = 2,
  labelcat = row.names(dfxy), clabelcat = 1, cpointcat = if (clabelcat == 0) 2 else 0,
  labelrowprof = row.names(dfrowprof), clabelrowprof = 0.75,
  cpointrowprof = if (clabelrowprof == 0) 2 else 0, pchrowprof = 20,
  coulrowprof = grey(0.8), proba = 0.95, n.sample = apply(dfrowprof, 1, sum),
  axesell = TRUE, ...)
```

Arguments

dfxy dfxy is a data frame containing at least two numerical variables. The rows of dfxy are categories such as 1,2 and 3 in the triangular plot.

dfrowprof dfrowprof is a data frame whose the columns are the rows of dfxy. The rows of dfxy are profiles or frequency distributions on the categories. The column number of dfrowprof must be equal to the row number of dfxy. row.names(dfxy) and names(dfrowprof) must be identical.

translate	a logical value indicating whether the plot should be translated(TRUE) or not. The origin becomes the gravity center weighted by profiles.
xax	the column number of dfxy for the x-axis
yax	the column number of dfxy for the y-axis
labelcat	a vector of strings of characters for the labels of categories
clabelcat	an integer specifying the character size for the labels of categories, used with par("cex")*clabelcat
cpointcat	an integer specifying the character size for the points showing the categories, used with par("cex")*cpointcat
labelrowprof	a vector of strings of characters for the labels of profiles (rows of dfrowprof)
clabelrowprof	an integer specifying the character size for the labels of profiles used with par("cex")*clabelrowprof
cpointrowprof	an integer specifying the character size for the points representative of the profiles used with par("cex")*cpointrowprof
pchrowprof	either an integer specifying a symbol or a single character to be used for the profile labels
coulrowprof	a vector of colors used for ellipses, possibly recycled
proba	a value lying between 0.500 and 0.999 to draw a confidence interval
n.sample	a vector containing the sample size, possibly recycled. Used n.sample = 0 if the profiles are not issued from a multinomial distribution and that confidence intervals have no sense.
axesell	a logical value indicating whether the ellipse axes should be drawn
...	further arguments passed from the s.label for the initial scatter plot.

Value

Returns in a hidden way a list of three components :

tra	a vector with two values giving the done original translation.
ell	a matrix, with 5 columns and for rows the number of profiles, giving the means, the variances and the covariance of the profile for the used numerical codes (column of dfxy)
call	the matched call

Author(s)

Daniel Chessel

Examples

```
par(mfrow = c(2,2))
par(mar = c(0.1,0.1,0.1,0.1))
proba <- matrix(c(0.49,0.47,0.04,0.4,0.3,0.3,0.05,0.05,0.9,0.05,0.7,0.25), ncol = 3, byrow = TRUE)
proba.df <- as.data.frame (proba)
names(proba.df) <- c("A","B","C") ; row.names(proba.df) <- c("P1","P2","P3","P4")
w.proba <- triangle.plot(proba.df, clab = 2, show = FALSE)
```

```

box()

w.tri = data.frame(x = c(-sqrt(1/2),sqrt(1/2),0), y = c(-1/sqrt(6),-1/sqrt(6),2/sqrt(6)))
L3 <- c("A","B","C")
row.names(w.tri) <- L3
s.multinom(w.tri, proba.df, n.sample = 0, coulrowprof = "black", clabelrowprof = 1.5)
s.multinom(w.tri, proba.df, n.sample = 30, coul = palette()[5])
s.multinom(w.tri, proba.df, n.sample = 60, coul = palette()[6], add.p = TRUE)
s.multinom(w.tri, proba.df, n.sample = 120, coul = grey(0.8), add.p = TRUE)

print(s.multinom(w.tri, proba.df[-3,], n.sample = 0, translate = TRUE)$tra)

```

s.traject

*Trajectory Plot***Description**

performs the scatter diagram with trajectories.

Usage

```

s.traject(dfxy, fac = factor(rep(1, nrow(dfxy))),
  ord = (1:length(fac)), xax = 1, yax = 2, label = levels(fac),
  clabel = 1, cpoint = 1, pch = 20, xlim = NULL, ylim = NULL,
  grid = TRUE, addaxes = TRUE, edge = TRUE, origin = c(0,0),
  include.origin = TRUE, sub = "", csub = 1, possub = "bottomleft",
  cgrid = 1, pixmap = NULL, contour = NULL, area = NULL, add.plot = FALSE)

```

Arguments

dfxy	a data frame containing two columns for the axes
fac	a factor partitioning the rows of the data frame in classes
ord	a vector of length equal to fac. The trajectory is drawn in an ascending order of the ord values
xax	the column number for the x-axis
yax	the column number for the y-axis
label	a vector of strings of characters for the point labels
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
xlim	the ranges to be encompassed by the x, if NULL they are computed
ylim	the ranges to be encompassed by the y, if NULL they are computed

grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted
edge	if TRUE the arrows are plotted, otherwise only the segments
origin	the fixed point in the graph space, for example c(0,0) the origin axes
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
pixmap	aan object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
rw <- function(a){
  x <- 0
  for(i in 1:49) x <- c(x,x[length(x)] + runif(1,-1,1))
  x
}
y <- unlist(lapply(1:5, rw))
x <- unlist(lapply(1:5, rw))
z <- gl(5,50)
s.traject(data.frame(x,y), z, edge = FALSE)
```

s.value

*Representation of a value in a graph***Description**

performs the scatter diagram with the representation of a value for a variable

Usage

```
s.value(dfxy, z, xax = 1, yax = 2, method = c("squaresize", "greylevel"),
        zmax=NULL, csize = 1, cpoint = 0, pch = 20, clegend = 0.75, neig = NULL,
        cneig = 1, xlim = NULL, ylim = NULL, grid = TRUE, addaxes = TRUE,
        cgrid = 0.75, include.origin = TRUE, origin = c(0,0), sub = "",
        csub = 1, possub = "topleft", pixmap = NULL, contour = NULL,
        area = NULL, add.plot = FALSE)
```

Arguments

dfxy	a data frame with two coordinates
z	a vector of the values corresponding to the rows of dfxy
xax	column for the x axis
yax	column for the y axis
method	a string of characters "squaresize" gives black squares for positive values and white for negative values with a proportional area equal to the absolute value. "greylevel" gives squares of equal size with a grey level proportional to the value. By default the first choice
zmax	a numeric value, equal by default to $\max(\text{abs}(z))$, can be used to impose a common scale of the size of the squares to several drawings in the same device
csize	a size coefficient for symbols
cpoint	a character size for plotting the points, used with $\text{par}(\text{"cex"}) * \text{cpoint}$. If zero, no points are drawn
pch	if $\text{cpoint} > 0$, an integer specifying the symbol or the single character to be used in plotting points
clegend	a character size for the legend used by $\text{par}(\text{"cex"}) * \text{clegend}$
neig	a neighbouring graph
cneig	a size for the neighbouring graph lines used with $\text{par}(\text{"lwd"}) * \text{cneig}$
xlim	the ranges to be encompassed by the x, if NULL they are computed
ylim	the ranges to be encompassed by the y, if NULL they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
addaxes	a logical value indicating whether the axes should be plotted

cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
pixmap	an object 'pixmap' displayed in the map background
contour	a data frame with 4 columns to plot the contour of the map : each row gives a segment (x1,y1,x2,y2)
area	a data frame of class 'area' to plot a set of surface units in contour
add.plot	if TRUE uses the current graphics window

Value

The matched call.

Author(s)

Daniel Chessel

Examples

```
xy <- cbind.data.frame(x = runif(500), y = runif(500))
z <- rnorm(500)
s.value(xy, z)

s.value(xy, z, method = "greylevel")

data(rpjdl)
fau.coa <- dudi.coa(rpjdl$fau, scan = FALSE, nf = 3)
s.value(fau.coa$li, fau.coa$li[,3], csi = 0.75, cleg = 0.75)

data(irishdata)
par(mfrow = c(3,4))
irq0 <- data.frame(scale(irishdata$stab, scale = TRUE))
for (i in 1:12) {
  z <- irq0[,i] ; nam <- names(irq0)[i]
  s.value(irishdata$xy, z, area = irishdata$area, csi = 3,
  csub = 2, sub = nam, cleg = 1.5, cgrid = 0, inc = FALSE,
  xlim = c(16,205), ylim = c(-50,268), adda = FALSE, grid = FALSE)
}
```

 santacatalina

Indirect Ordination

Description

This data set gives the densities per hectare of 11 species of trees for 10 transects of topographic moisture values (mean of several stations per class).

Usage

```
data(santacatalina)
```

Format

a data frame with 11 rows and 10 columns

Source

Gauch, H. G. J., Chase, G. B. and Whittaker R. H. (1974) Ordination of vegetation samples by Gaussian species distributions. *Ecology*, **55**, 1382–1390.

Examples

```
data(santacatalina)
par(mfrow = c(2,2))
table.value(log(santacatalina + 1), grid = TRUE)
table.value(log(santacatalina + 1)[,sample(10)], grid = TRUE)
coa1 <- dudi.coa(log(santacatalina + 1), scan = FALSE) # 2 factors
table.value(log(santacatalina + 1)[order(coa1$li[,1]),
  order(coa1$co[,1])], grid = TRUE)
scatter(coa1, posi = "bottom")
```

 sarcelles

Array of Recapture of Rings

Description

The data frame `sarcelles$tab` contains the number of the winter teals (*Anas C. Crecca*) for which the ring was retrieved in the area i during the month j ($n=3049$).

Usage

```
data(sarcelles)
```

Format

`sarcelles` is a list of 4 components.

tab is a data frame with 14 rows-areas and 12 columns-months.

xy is a data frame with the 2 spatial coordinates of the 14 region centers.

neig is the neighbouring graph between areas, object of the class `neig`.

col.names is a vector containing the month items

Source

Lebreton, J.D. (1973) Etude des déplacements saisonniers des Sarcelles d'hiver, Anas c. crecca L., hivernant en Camargue à l'aide de l'analyse factorielle des correspondances. *Compte rendu hebdomadaire des séances de l'Académie des sciences*, Paris, D, III, **277**, 2417–2420.

Examples

```
## Not run:
# depends of pixmap
if (require(pixmap, quietly=TRUE)) {
  bkgnd.pnm <- read.pnm(system.file("pictures/sarcelles.pnm", package = "ade4"))
  data(sarcelles)
  par(mfrow = c(4,3))
  for(i in 1:12) {
    s.distri(sarcelles$xy, sarcelles$tab[,i], pixmap = bkgnd.pnm,
             sub = sarcelles$col.names[i], clab = 0, csub = 2)
    s.value(sarcelles$xy, sarcelles$tab[,i], add.plot = TRUE, cleg = 0)
  }
}

## End(Not run)
```

scalewt

Compute or scale data using (weighted) means, variances and covariances (possibly for the levels of a factor)

Description

These utility functions compute (weighted) means, variances and covariances for dataframe partitioned by a factor. The scale transforms a numeric matrix in a centred and scaled matrix for any weighting.

Usage

```
covwt(x, wt, na.rm = FALSE)
varwt(x, wt, na.rm = FALSE)
scalewt(df, wt = rep(1/nrow(df), nrow(df)), center = TRUE, scale = TRUE)
meanfacwt(df, fac = NULL, wt = rep(1/nrow(df), nrow(df)), drop = FALSE)
```

```
varfacwt(df, fac = NULL, wt = rep(1/nrow(df), nrow(df)), drop = FALSE)
covfacwt(df, fac = NULL, wt = rep(1/nrow(df), nrow(df)), drop = FALSE)
scalefacwt(df, fac = NULL, wt = rep(1/nrow(df), nrow(df)), scale = TRUE, drop = FALSE)
```

Arguments

x	a numeric vector (varwt) or a matrix (covwt) containing the data.
na.rm	a logical value indicating whether NA values should be stripped before the computation proceeds.
df	a dataframe containing the data.
fac	a factor partitioning the data.
wt	a numeric vector of weights.
drop	a logical value indicating whether unused levels should be kept.
scale	a logical value indicating whether data should be scaled or not.
center	a logical value indicating whether data should be centered or not.

Details

Functions returns biased estimates of variances and covariances (i.e. divided by n and not n-1)

Value

For varwt, the weighted variance. For covwt, the matrix of weighted co-variances. For scalewt, the scaled dataframe. For other function a list (if fac is not null) of dataframes with appropriate values

Author(s)

Stephane Dray <stephane.dray@univ-lyon1.fr>

Examples

```
data(meau)
w <- rowSums(meau$spe)
varwt(meau$env, w)
varfacwt(meau$env, wt = w)
varfacwt(meau$env, wt = w, fac = meau$design$season)
covfacwt(meau$env, wt = w, fac = meau$design$season)
scalewt(meau$env, wt = w)
```

`scatter`*Graphical representation of the outputs of a multivariate analysis*

Description

`scatter` is a generic function that has methods for the classes `coa`, `dudi`, `fca`, `acm` and `pco`. It plots the outputs of a multivariate analysis by representing simultaneously the rows and the columns of the original table (biplot). The function `biplot` returns exactly the same representation.

The function `screepplot` represents the amount of inertia (usually variance) associated to each dimension.

Usage

```
scatter(x, ...)
## S3 method for class 'dudi'
biplot(x, ...)
## S3 method for class 'dudi'
screepplot(x, npcs = length(x$eig), type = c("barplot", "lines"),
  main = deparse(substitute(x)), col = c(rep("black", x$nf),
  rep("grey", npcs - x$nf)), ...)
```

Arguments

<code>x</code>	an object of the class <code>dudi</code> containing the outputs of a multivariate analysis
<code>npcs</code>	the number of components to be plotted
<code>type</code>	the type of plot
<code>main</code>	the title of the plot
<code>col</code>	a vector of colors
<code>...</code>	further arguments passed to or from other methods

Author(s)

Daniel Chessel
Stephane Dray <stephane.drays@univ-lyon1.fr>

See Also

[s.arrow](#), [s.chull](#), [s.class](#), [s.corcircle](#), [s.distri](#), [s.label](#), [s.match](#), [s.traject](#), [s.value](#),
[add.scatter](#)

Examples

```
data(rpjdl)
rpjdl.coa <- dudi.coa(rpjdl$fau, scannf = FALSE, nf = 4)
screepplot(rpjdl.coa)
biplot(rpjdl.coa)
```

 scatter.acm

Plot of the factorial maps in a Multiple Correspondence Analysis

Description

performs the scatter diagrams of a Multiple Correspondence Analysis.

Usage

```
## S3 method for class 'acm'
scatter(x, xax = 1, yax = 2, mfrow=NULL, csub = 2, possub = "topleft", ...)
```

Arguments

x	an object of class acm
xax	the column number for the x-axis
yax	the column number for the y-axis
mfrow	a vector of the form "c(nr,nc)", if NULL (the default) is computed by n2mfrow
csub	a character size for the legend, used with par("cex")*csub
possub	a string of characters indicating the legend position ("topleft", "topright", "bottomleft", "bottomright") in a array of figures
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(lascaux)
scatter(dudi.acm(lascaux$ornem, sca = FALSE), csub = 3)
```

 scatter.coa

Plot of the factorial maps for a correspondence analysis

Description

performs the scatter diagrams of a correspondence analysis.

Usage

```
## S3 method for class 'coa'
scatter(x, xax = 1, yax = 2, method = 1:3, clab.row = 0.75,
       clab.col = 1.25, posieig = "top", sub = NULL, csub = 2, ...)
```

Arguments

x	an object of class coa
xax	the column number for the x-axis
yax	the column number for the y-axis
method	an integer between 1 and 3 1 Rows and columns with the coordinates of lambda variance 2 Rows variance 1 and columns by averaging 3 Columns variance 1 and rows by averaging
clab.row	a character size for the rows
clab.col	a character size for the columns
posieig	if "top" the eigenvalues bar plot is upside,vif "bottom" it is downside, if "none" no plot
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with par("cex")*csub
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

References

Oksanen, J. (1987) Problems of joint display of species and site scores in correspondence analysis. *Vegetatio*, **72**, 51–57.

Examples

```
data(housetasks)
par(mfrow = c(2,2))
w <- dudi.coa(housetasks, scan = FALSE)
scatter.dudi(w, sub = "0 / To be avoided")
scatter.coa(w, method = 1, sub = "1 / Standard", posieig = "none")
scatter.coa(w, method = 2,
  sub = "2 / Columns -> averaging -> Rows", posieig = "none")
scatter.coa(w, method = 3,
  sub = "3 / Rows -> averaging -> Columns ", posieig = "none")
```

scatter.dudi

Plot of the Factorial Maps

Description

performs the scatter diagrams of objects of class dudi.

Usage

```
## S3 method for class 'dudi'
scatter(x, xax = 1, yax = 2, clab.row = 0.75, clab.col = 1,
        permute = FALSE, posieig = "top", sub = NULL, ...)
```

Arguments

x	an object of class dudi
xax	the column number for the x-axis
yax	the column number for the y-axis
clab.row	a character size for the rows
clab.col	a character size for the columns
permute	if FALSE, the rows are plotted by points and the columns by arrows. If TRUE it is the opposite.
posieig	if "top" the eigenvalues bar plot is upside, if "bottom" it is downside, if "none" no plot
sub	a string of characters to be inserted as legend
...	further arguments passed to or from other methods

Details

scatter.dudi is a factorial map of individuals and the projection of the vectors of the canonical basis multiplied by a constante of rescaling. In the eigenvalues bar plot, the used axes for the plot are in black, the other kept axes in grey and the other in white.

The permute argument can be used to choose between the distance biplot (default) and the correlation biplot (permute = TRUE).

Author(s)

Daniel Chessel

Examples

```
data(deug)
scatter(dd1 <- dudi.pca(deug$stab, scannf = FALSE, nf = 4),
        posieig = "bottomright")

data(rhone)
dd1 <- dudi.pca(rhone$stab, nf = 4, scann = FALSE)
scatter(dd1, sub = "Principal component analysis")
```

scatter.fca

Plot of the factorial maps for a fuzzy correspondence analysis

Description

performs the scatter diagrams of a fuzzy correspondence analysis.

Usage

```
## S3 method for class 'fca'
scatter(x, xax = 1, yax = 2, clab.moda = 1, labels = names(x$tab),
        sub = NULL, csub = 2, ...)
```

Arguments

x	an object of class fca
xax	the column number for the x-axis
yax	the column number for the y-axis
clab.moda	the character size to write the modalities
labels	a vector of strings of characters for the labels of the modalities
sub	a vector of strings of characters to be inserted as legend in each figure
csub	a character size for the legend, used with <code>par("cex")*csub</code>
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

References

Chevenet, F., Dolédec, S. and Chessel, D. (1994) A fuzzy coding approach for the analysis of long-term ecological data. *Freshwater Biology*, **31**, 295–309.

Examples

```
data(coleo)
coleo.fuzzy <- prep.fuzzy.var(coleo$tab, coleo$col.blocks)
fca1 <- dudi.fca(coleo.fuzzy, sca = FALSE, nf = 3)
scatter(fca1, labels = coleo$moda.names, clab.moda = 1.5,
        sub = names(coleo$col.blocks), csub = 3)
```

`scatterutil`*Graphical utility functions*

Description

These are utilities used in graphical functions.

Details

The functions scatter use some utilities functions :

scatterutil.base defines the layer of the plot for all scatters

scatterutil.sco defines the layer of the plot for sco functions

scatterutil.chull plots the polygons of the external contour

scatterutil.eigen plots the eigenvalues bar plot

scatterutil.ellipse plots an inertia ellipse for a weighting distribution

scatterutil.eti.circ puts labels on a correlation circle

scatterutil.eti puts labels centred on the points

scatterutil.grid plots a grid and adds a legend

scatterutil.legend.bw.square puts a legend of values by square size

scatterutil.legend.square.grey puts a legend by squares and grey levels

scatterutil.legendgris adds a legend of grey levels for the areas

scatterutil.scaling to fit a plot on a background bipmap

scatterutil.star plots a star for a weighting distribution

scatterutil.sub adds a string of characters in sub-title of a graph

scatterutil.convrot90 is used to rotate labels

Author(s)

Daniel Chessel, Stephane Dray <dray@biomserv.univ-lyon1.fr>

See Also

[s.arrow](#), [s.chull](#), [s.class](#), [s.corcircle](#), [s.distri](#), [s.label](#), [s.match](#), [s.traject](#), [s.value](#),
[add.scatter](#)

Examples

```

par(mfrow = c(3,3))
plot.new()
ade4:::scatterutil.legendgris(1:20, 4, 1.6)

plot.new()
ade4:::scatterutil.sub("lkn5555555551kn", csub = 2, possub = "bottomleft")
ade4:::scatterutil.sub("lkn5555555551kn", csub = 1, possub = "topleft")
ade4:::scatterutil.sub("jdj1", csub = 3, possub = "topright")
ade4:::scatterutil.sub("**", csub = 2, possub = "bottomright")

x <- c(0.5,0.2,-0.5,-0.2) ; y <- c(0.2,0.5,-0.2,-0.5)
eti <- c("toto", "kjbk", "gdgiglgl", "sdfg")
plot(x, y, xlim = c(-1,1), ylim = c(-1,1))
ade4:::scatterutil.eti.circ(x, y, eti, 2.5)
abline(0, 1, lty = 2) ; abline(0, -1, lty = 2)

x <- c(0.5,0.2,-0.5,-0.2) ; y <- c(0.2,0.5,-0.2,-0.5)
eti <- c("toto", "kjbk", "gdgiglgl", "sdfg")
plot(x, y, xlim = c(-1,1), ylim = c(-1,1))
ade4:::scatterutil.eti(x, y, eti, 1.5)

plot(runif(10,-3,5), runif(10,-1,1), asp = 1)
ade4:::scatterutil.grid(2)
abline(h = 0, v = 0, lwd = 3)

x <- runif(10,0,1) ; y <- rnorm(10) ; z <- rep(1,10)
plot(x,y) ; ade4:::scatterutil.star(x, y, z, 0.5)
plot(x,y) ; ade4:::scatterutil.star(x, y, z, 1)

x <- c(runif(10,0,0.5), runif(10,0.5,1))
y <- runif(20)
plot(x, y, asp = 1) # asp=1 is essential to have perpendicular axes
ade4:::scatterutil.ellipse(x, y, rep(c(1,0), c(10,10)), cell = 1.5, ax = TRUE)
ade4:::scatterutil.ellipse(x, y, rep(c(0,1), c(10,10)), cell = 1.5, ax = TRUE)

x <- c(runif(100,0,0.75), runif(100,0.25,1))
y <- c(runif(100,0,0.75), runif(100,0.25,1))
z <- factor(rep(c(1,2), c(100,100)))
plot(x, y, pch = rep(c(1,20), c(100,100)))
ade4:::scatterutil.chull(x, y, z, opt = c(0.25,0.50,0.75,1))
par(mfrow = c(1,1))

```

sco.boxplot

*Representation of the link between a variable and a set of qualitative variables***Description**

represents the link between a variable and a set of qualitative variables.

Usage

```
sco.boxplot(score, df, labels = names(df), clabel = 1, xlim = NULL,
            grid = TRUE, cgrid = 0.75, include.origin = TRUE, origin = 0,
            sub = NULL, csub = 1)
```

Arguments

score	a numeric vector
df	a data frame with only factors
labels	a vector of strings of characters for the labels of variables
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
xlim	the ranges to be encompassed by the x axis, if NULL they are computed
grid	a logical value indicating whether the scale vertical lines should be drawn
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the scale
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example 0 the origin axis
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>

Author(s)

Daniel Chessel

Examples

```
w1 <- rnorm(100,-1)
w2 <- rnorm(100)
w3 <- rnorm(100,1)
f1 <- gl(3,100)
f2 <- gl(30,10)
sco.boxplot(c(w1,w2,w3), data.frame(f1,f2))

data(banque)
banque.acm <- dudi.acm(banque, scan = FALSE, nf = 4)
par(mfrow = c(1,3))
sco.boxplot(banque.acm$11[,1], banque[,1:7], clab = 1.8)
sco.boxplot(banque.acm$11[,1], banque[,8:14], clab = 1.8)
sco.boxplot(banque.acm$11[,1], banque[,15:21], clab = 1.8)
par(mfrow = c(1,1))
```


sco.class

*1D plot of a numeric score and a factor with labels***Description**

Draws evenly spaced labels, each label linked to the corresponding values of the levels of a factor.

Usage

```
sco.class(score, fac, label = levels(fac), clabel = 1, horizontal = TRUE,
reverse = FALSE, pos.lab = 0.5, pch = 20, cpoint = 1, boxes = TRUE,
col = rep(1, length(levels(fac))), lim = NULL, grid = TRUE,
cgrid = 1, include.origin = TRUE, origin = c(0, 0), sub = "",
csub = 1.25, possub = "bottomleft")
```

Arguments

score	a numeric vector
fac	a factor
label	labels for the levels of the factor
clabel	a character size for the labels, used with <code>par("cex")*clabel</code>
horizontal	logical. If TRUE, the plot is horizontal
reverse	logical. If horizontal = TRUE and reverse=TRUE, the plot is at the bottom, if reverse = FALSE, the plot is at the top. If horizontal = FALSE, the plot is at the right (TRUE) or at the left (FALSE).
pos.lab	a values between 0 and 1 to manage the position of the labels.
pch	an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
boxes	if TRUE, labels are framed
col	a vector of colors used to draw each class in a different color
lim	the range for the x axis or y axis (if horizontal = FALSE), if NULL, they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should belong to the plot
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")

Value

The matched call.

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
data(meau)
envpca <- dudi.pca(meau$env, scannf=FALSE)
par(mfrow=c(2,1))
sco.class(envpca$li[,1],meau$design$season, col = 1:6)
sco.class(envpca$li[,1],meau$design$season, col = 1:4, reverse = TRUE)
```

sco.distri	<i>Representation by mean- standard deviation of a set of weight distributions on a numeric score</i>
------------	---

Description

represents the mean- standard deviation of a set of weight distributions on a numeric score.

Usage

```
sco.distri(score, df, y.rank = TRUE, csize = 1, labels = names(df),
  clabel = 1, xlim = NULL, grid = TRUE, cgrid = 0.75,
  include.origin = TRUE, origin = 0, sub = NULL, csub = 1)
```

Arguments

score	a numeric vector
df	a data frame with only positive or null values
y.rank	a logical value indicating whether the means should be classified in ascending order
csize	an integer indicating the size segment
labels	a vector of strings of characters for the labels of the variables
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
xlim	the ranges to be encompassed by the x axis, if NULL they are computed
grid	a logical value indicating whether the scale vertical lines should be drawn
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the scale
include.origin	a logical value indicating whether the point "origin" should be belonged to the graph space
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>

Value

returns an invisible data.frame with means and variances

Author(s)

Daniel Chessel

Examples

```
w <- seq(-1, 1, le = 200)
distri <- data.frame(lapply(1:50,
  function(x) sample((200:1)) * ((w >= (-x/50)) & (w <= x/50)) ))
names(distri) <- paste("w", 1:50, sep = "")
par(mfrow = c(1,2))
sco.distri(w, distri, csi = 1.5)
sco.distri(w, distri, y.rank = FALSE, csi = 1.5)
par(mfrow = c(1,1))

data(rpjdl)
coa2 <- dudi.coa(rpjdl$fau, FALSE)
sco.distri(coa2$li[,1], rpjdl$fau, lab = rpjdl$frlab, clab = 0.8)

data(doubs)
par(mfrow = c(2,2))
poi.coa <- dudi.coa(doubs$fish, scann = FALSE)
sco.distri(poi.coa$li[,1], doubs$fish)
poi.nsc <- dudi.nsc(doubs$fish, scann = FALSE)
sco.distri(poi.nsc$li[,1], doubs$fish)
s.label(poi.coa$li)
s.label(poi.nsc$li)

data(rpjdl)
fau.coa <- dudi.coa(rpjdl$fau, scann = FALSE)
sco.distri(fau.coa$li[,1], rpjdl$fau)
fau.nsc <- dudi.nsc(rpjdl$fau, scann = FALSE)
sco.distri(fau.nsc$li[,1], rpjdl$fau)
s.label(fau.coa$li)
s.label(fau.nsc$li)

par(mfrow = c(1,1))
```

sco.gauss

Relationships between one score and qualitative variables

Description

Draws Gauss curves with the same mean and variance as the scores of individuals belonging to categories of several qualitative variables.

Usage

```
sco.gauss(score, df, xlim = NULL, steps = 200, ymax = NULL, sub =
names(df), csub = 1.25, possub = "topleft", legen =TRUE, label = row.names(df),
clabel = 1, grid = TRUE, cgrid = 1, include.origin = TRUE, origin = c(0, 0))
```

Arguments

score	a numeric vector
df	a dataframe containing only factors, number of rows equal to the length of the score vector
xlim	starting point and end point for drawing the Gauss curves
steps	number of segments for drawing the Gauss curves
ymax	max ordinate for all Gauss curves. If NULL, ymax is computed and different for each factor
sub	vector of strings of characters for the lables of qualitative variables
csub	character size for the legend
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
legen	if TRUE, the first graphic of the series displays the score with evenly spaced labels (see sco.label)
label	labels for the score
clabel	a character size for the labels, used with par("cex")*clabel
grid	a logical value indicating whether a grid in the background of the plot should be drawn
cgrid	a character size, parameter used with par("cex")*cgrid to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should belong to the plot
origin	the fixed point in the graph space, for example c(0,0) the origin axes

Details

Takes one vector containing quantitative values (score) and one dataframe containing only factors that give categories to wich the quantitative values belong. Computes the mean and variance of the values in each category of each factor, and draws a Gauss curve with the same mean and variance for each category of each factor. Can optionally set the start and end point of the curves and the number of segments. The max ordinate (ymax) can also be set arbitrarily to set a common max for all factors (else the max is different for each factor).

Value

The matched call.

Author(s)

Jean Thioulouse, Stephane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
data(meau)
envpca <- dudi.pca(meau$env, scannf=FALSE)
dffac <- cbind.data.frame(meau$design$season, meau$design$site)
sco.gauss(envpca$li[,1], dffac, clabel = 2, csub = 2)
```

sco.label	<i>1D plot of a numeric score with labels</i>
-----------	---

Description

Draws evenly spaced labels, each label linked to the corresponding value of a numeric score.

Usage

```
sco.label(score, label = names(score), clabel = 1, horizontal = TRUE,
reverse = FALSE, pos.lab = 0.5, pch = 20, cpoint = 1, boxes = TRUE, lim
= NULL, grid = TRUE, cgrid = 1, include.origin = TRUE, origin = c(0, 0),
sub = "", csub = 1.25, possub = "bottomleft")
```

Arguments

score	a numeric vector
label	labels for the score
clabel	a character size for the labels, used with <code>par("cex")*clabel</code>
horizontal	logical. If TRUE, the plot is horizontal
reverse	logical. If horizontal = TRUE and reverse=TRUE, the plot is at the bottom, if reverse = FALSE, the plot is at the top. If horizontal = FALSE, the plot is at the right (TRUE) or at the left (FALSE).
pos.lab	a values between 0 and 1 to manage the position of the labels.
pch	an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
boxes	if TRUE, labels are framed
lim	the range for the x axis or y axis (if horizontal = FALSE), if NULL, they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
cgrid	a character size, parameter used with <code>par("cex")* cgrid</code> to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should belong to the plot
origin	the fixed point in the graph space, for example <code>c(0,0)</code> the origin axes

sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")

Value

The matched call.

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>, Jean Thioulouse

Examples

```
data(meau)
envpca <- dudi.pca(meau$env, scannf=FALSE)
par(mfrow=c(2,1))
sco.label(envpca$11[,1], row.names(envpca$11), lim=c(-1,3.5))
sco.label(envpca$co[,1], row.names(envpca$co), reverse = TRUE, lim=c(-1,3.5))
```

sco.match

1D plot of a pair of numeric scores with labels

Description

Draws evenly spaced labels, each label linked to the corresponding values of two numeric score.

Usage

```
sco.match(score1, score2, label = names(score1), clabel = 1,
  horizontal = TRUE, reverse = FALSE, pos.lab = 0.5, wmatch = 3,
  pch = 20, cpoint = 1, boxes = TRUE, lim = NULL, grid = TRUE,
  cgrid = 1, include.origin = TRUE, origin = c(0, 0), sub = "",
  csub = 1.25, possub = "bottomleft")
```

Arguments

score1	a numeric vector
score2	a numeric vector
label	labels for the score
clabel	a character size for the labels, used with <code>par("cex")*clabel</code>
horizontal	logical. If TRUE, the plot is horizontal
reverse	logical. If horizontal = TRUE and reverse=TRUE, the plot is at the bottom, if reverse = FALSE, the plot is at the top. If horizontal = FALSE, the plot is at the right (TRUE) or at the left (FALSE).

pos.lab	a values between 0 and 1 to manage the position of the labels.
wmatch	a numeric values to specify the width of the matching region in the plot. The width is equal to wmatch * the height of character
pch	an integer specifying the symbol or the single character to be used in plotting points
cpoint	a character size for plotting the points, used with par("cex")*cpoint. If zero, no points are drawn
boxes	if TRUE, labels are framed
lim	the range for the x axis or y axis (if horizontal = FALSE), if NULL, they are computed
grid	a logical value indicating whether a grid in the background of the plot should be drawn
cgrid	a character size, parameter used with par("cex")* cgrid to indicate the mesh of the grid
include.origin	a logical value indicating whether the point "origin" should belong to the plot
origin	the fixed point in the graph space, for example c(0,0) the origin axes
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with par("cex")*csub
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")

Value

The matched call.

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>

Examples

```
sco.match(-5:5, 2*(-5:5))
```

sco.quant	<i>Graph to Analyse the Relation between a Score and Quantitative Variables</i>
-----------	---

Description

represents the graphs to analyse the relation between a score and quantitative variables.

Usage

```
sco.quant (score, df, fac = NULL, clabel = 1, abline = FALSE,
           sub = names(df), csub = 2, possub = "topleft")
```

Arguments

score	a numeric vector
df	a data frame which rows equal to the score length
fac	a factor with the same length than the score
clabel	character size for the class labels (if any) used with <code>par("cex")*clabel</code>
abline	a logical value indicating whether a regression line should be added
sub	a vector of strings of characters for the labels of variables
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")

Author(s)

Daniel Chessel

Examples

```
w <- runif(100, -5, 10)
fw <- cut(w, 5)
levels(fw) <- LETTERS[1:5]
wX <- data.frame(matrix(w + rnorm(900, sd = (1:900) / 100), 100, 9))
sco.quant(w, wX, fac = fw, abline = TRUE, clab = 2, csub = 3)
```

score

Graphs for One Dimension

Description

score is a generic function. It proposes methods for the objects 'coa', 'acm', 'mix', 'pca'.

Usage

```
score(x, ...)
scoreutil.base(y, xlim, grid, cgrid, include.origin, origin, sub, csub)
```

Arguments

x	an object used to select a method
...	further arguments passed to or from other methods
y	a numeric vector
xlim	the ranges to be encompassed by the x axis, if NULL they are computed
grid	a logical value indicating whether the scale vertical lines should be drawn
cgrid	a character size, parameter used with <code>par("cex")*cgrid</code> to indicate the mesh of the scale

include.origin a logical value indicating whether the point "origin" should be belonged to the graph space

origin the fixed point in the graph space, for example 0 the origin axis

sub a string of characters to be inserted as legend

csub a character size for the legend, used with `par("cex")*csub`

Details

scoreutil.base is a utility function - not for the user - to define the bottom of the layout of all score.

Author(s)

Daniel Chessel

See Also

[sco.boxplot](#), [sco.distri](#), [sco.quant](#)

Examples

```
## Not run:
par(mar = c(1,1,1,1))
scoreutil.base (runif(20,3,7), xlim = NULL, grid = TRUE, cgrid = 0.8,
  include.origin = TRUE, origin = 0, sub = "Uniform", csub = 1)
## End(Not run)
# returns the value of the user coordinate of the low line.
# The user window id defined with c(0,1) in ordinate.
# box()
```

score.acm

Graphs to study one factor in a Multiple Correspondence Analysis

Description

performs the canonical graph of a Multiple Correspondence Analysis.

Usage

```
## S3 method for class 'acm'
score(x, xax = 1, which.var = NULL, mfrow = NULL,
  sub = names(oritab), csub = 2, possub = "topleft", ...)
```

Arguments

x	an object of class acm
xax	the column number for the used axis
which.var	the numbers of the kept columns for the analysis, otherwise all columns
mfrow	a vector of the form "c(nr,nc)", otherwise computed by a special own function n2mfrow
sub	a vector of strings of characters to be inserted as sub-titles, otherwise the variable names of the initial array
csub	a character size for the sub-titles
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(banque)
banque.acm <- dudi.acm(banque, scann = FALSE, nf = 3)
score(banque.acm, which = which(banque.acm$cr[,1] > 0.2), csub = 3)
```

score.coa

Reciprocal scaling after a correspondence analysis

Description

performs the canonical graph of a correspondence analysis.

Usage

```
## S3 method for class 'coa'
score(x, xax = 1, dotchart = FALSE, clab.r = 1, clab.c = 1,
      csub = 1, cpoi = 1.5, cet = 1.5, ...)
reciprocal.coa(x)
```

Arguments

x	an object of class coa
xax	the column number for the used axis
dotchart	if TRUE the graph gives a "dual scaling", if FALSE a "reciprocal scaling"
clab.r	a character size for row labels
clab.c	a character size for column labels

csub	a character size for the sub-titles, used with <code>par("cex")*csub</code>
cpoi	a character size for the points
cet	a coefficient for the size of segments in standard deviation
...	further arguments passed to or from other methods

Details

In a "reciprocal scaling", the reference score is a numeric code centred and normalized of the non zero cells of the array which both maximizes the variance of means by row and by column. The bars are drawn with half the length of this standard deviation.

Value

return a data.frame with the scores, weights and factors of correspondences (non zero cells)

Author(s)

Daniel Chessel

References

Thioulouse, J. and Chessel D. (1992) A method for reciprocal scaling of species tolerance and sample diversity. *Ecology*, **73**, 670–680.

Examples

```
layout(matrix(c(1,1,2,3), 2, 2), resp = FALSE)
data(aviurba)
dd1 <- dudi.coa(aviurba$fau, scan = FALSE)
score(dd1, clab.r = 0, clab.c = 0.75)
recscal <- reciprocal.coa(dd1)
head(recscal)
abline(v = 1, lty = 2, lwd = 3)
sco.distri(dd1$l1[,1], aviurba$fau)
sco.distri(dd1$c1[,1], data.frame(t(aviurba$fau)))

# 1 reciprocal scaling correspondence score -> species amplitude + sample diversity
# 2 sample score -> averaging -> species amplitude
# 3 species score -> averaging -> sample diversity

layout(matrix(c(1,1,2,3), 2, 2), resp = FALSE)
data(rpjdl)
rpjdl1 <- dudi.coa(rpjdl$fau, scan = FALSE)
score(rpjdl1, clab.r = 0, clab.c = 0.75)
if (require(MASS, quietly = TRUE)) {
  data(caith)
  score(dudi.coa(caith, scan = FALSE), clab.r = 1.5, clab.c = 1.5, cpoi = 3)
  data(housetasks)
  score(dudi.coa(housetasks, scan = FALSE), clab.r = 1.25, clab.c = 1.25,
        csub = 0, cpoi = 3)
}
```

```
par(mfrow = c(1,1))
score(rpjd11, dotchart = TRUE, clab.r = 0)
```

score.mix

Graphs to Analyse a factor in a Mixed Analysis

Description

performs the canonical graph of a mixed analysis.

Usage

```
## S3 method for class 'mix'
score(x, xax = 1, csub = 2, mfrow = NULL, which.var = NULL, ...)
```

Arguments

x	an object of class mix
xax	the column number for the used axis
csub	a character size for the sub-titles, used with par("cex")*csub
mfrow	a vector of the form "c(nr,nc)", otherwise computed by a special own function n2mfrow
which.var	the numbers of the kept columns for the analysis, otherwise all columns
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(lascaux)
w <- cbind.data.frame(lascaux$colo, lascaux$ornem)
dd <- dudi.mix(w, scan = FALSE, nf = 4, add = TRUE)
score(dd, which = which(dd$scr[,1] > 0.3))
```

score.pca

Graphs to Analyse a factor in PCA

Description

performs the canonical graph of a Principal Component Analysis.

Usage

```
## S3 method for class 'pca'
score(x, xax = 1, which.var = NULL, mfrow = NULL, csub = 2,
      sub = names(x$tab), abline = TRUE, ...)
```

Arguments

x	an object of class pca
xax	the column number for the used axis
which.var	the numbers of the kept columns for the analysis, otherwise all columns
mfrow	a vector of the form "c(nr,nc)", otherwise computed by a special own function n2mfrow
csub	a character size for sub-titles, used with par("cex")*csub
sub	a vector of string of characters to be inserted as sub-titles, otherwise the names of the variables
abline	a logical value indicating whether a regression line should be added
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel

Examples

```
data(deug)
dd1 <- dudi.pca(deug$tab, scan = FALSE)
score(dd1, csub = 3)

# The correlations are :
dd1$co[,1]
# [1] 0.7925 0.6532 0.7410 0.5287 0.5539 0.7416 0.3336 0.2755 0.4172
```

seconde

Students and Subjects

Description

The seconde data frame gives the marks of 22 students for 8 subjects.

Usage

```
data(seconde)
```

Format

This data frame (22,8) contains the following columns: - HGEO: History and Geography - FRAN: French literature - PHYS: Physics - MATH: Mathematics - BIOL: Biology - ECON: Economy - ANGL: English language - ESPA: Spanish language

Source

Personal communication

Examples

```
data(seconde)
scatter(dudi.pca(seconde, scan = FALSE), clab.r = 1, clab.c = 1.5)
```

sepan

Separated Analyses in a K-tables

Description

performs K separated multivariate analyses of an object of class ktab containing K tables.

Usage

```
sepan(X, nf = 2)
## S3 method for class 'sepan'
plot(x, mfrow = NULL, csub = 2, ...)
## S3 method for class 'sepan'
summary(object, ...)
## S3 method for class 'sepan'
print(x, ...)
```

Arguments

<code>X</code>	an object of class <code>ktab</code>
<code>nf</code>	an integer indicating the number of kept axes for each separated analysis
<code>x, object</code>	an object of class <code>'sepan'</code>
<code>mfrow</code>	a vector of the form <code>"c(nr,nc)"</code> , otherwise computed by a special own function <code>n2mfrow</code>
<code>csub</code>	a character size for the sub-titles, used with <code>par("cex")*csub</code>
<code>...</code>	further arguments passed to or from other methods

Details

The function plot on a `sepan` object allows to compare inertias and structures between arrays. In black, the eigenvalues of kept axes in the object `'sepan'`.

Value

returns a list of class `'sepan'` containing :

<code>call</code>	a call order
<code>tab.names</code>	a vector of characters with the names of tables
<code>blo</code>	a numeric vector with the numbers of columns for each table
<code>rank</code>	a numeric vector with the rank of the studied matrix for each table
<code>Eig</code>	a numeric vector with all the eigenvalues
<code>Li</code>	a data frame with the row coordinates
<code>L1</code>	a data frame with the row normed scores
<code>Co</code>	a data frame with the column coordinates
<code>C1</code>	a data frame with the column normed coordinates
<code>TL</code>	a data frame with the factors for <code>Li L1</code>
<code>TC</code>	a data frame with the factors for <code>Co C1</code>

Author(s)

Daniel Chessel

Examples

```
data(escopage)
w <- data.frame(scale(escopage$tab))
w <- ktab.data.frame(w, escopage$blo, tabnames = escopage$tab.names)
sep1 <- sepan(w)
sep1
summary(sep1)
plot(sep1)
```

 skulls

Morphometric Evolution

Description

This data set gives four anthropometric measures of 150 Egyptian skulls belonging to five different historical periods.

Usage

```
data(skulls)
```

Format

The skulls data frame has 150 rows (egyptean skulls) and 4 columns (anthropometric measures). The four variables are the maximum breadth (V1), the basibregmatic height (V2), the basialveolar length (V3) and the nasal height (V4). All measurements were taken in millimeters.

Details

The measurements are made on 5 groups and 30 Egyptian skulls. The groups are defined as follows :

- 1 - the early predynastic period (circa 4000 BC)
- 2 - the late predynastic period (circa 3300 BC)
- 3 - the 12th and 13th dynasties (circa 1850 BC)
- 4 - the Ptolemaic period (circa 200 BC)
- 5 - the Roman period (circa 150 BC)

Source

Thompson, A. and Randall-Maciver, R. (1905) *Ancient races of the Thebaid*, Oxford University Press.

References

Manly, B.F. (1994) *Multivariate Statistical Methods. A primer*, Second edition. Chapman & Hall, London. 1–215.

The example is treated pp. 6, 13, 51, 64, 72, 107, 112 and 117.

Examples

```
data(skulls)
pca1 <- dudi.pca(skulls, scan = FALSE)
fac <- gl(5, 30)
levels(fac) <- c("-4000", "-3300", "-1850", "-200", "+150")
dis.skulls <- discrimin(pca1, fac, scan = FALSE)
plot(dis.skulls, 1, 1)
```

statico	<i>STATIS and Co-Inertia : Analysis of a series of paired ecological tables</i>
---------	---

Description

Does the analysis of a series of pairs of ecological tables. This function uses Partial Triadic Analysis ([pta](#)) and [ktab.match2ktabs](#) to do the computations.

Usage

```
statico(KTX, KTY, scannf = TRUE)
```

Arguments

KTX	an objet of class ktab
KTY	an objet of class ktab
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed

Details

This function takes 2 ktabs and crosses each pair of tables of these ktabs with the function [ktab.match2ktabs](#). It then does a partial triadic analysis on this new ktab with [pta](#).

Value

a list of class ktab, subclass kcoinertia. See [ktab](#)

WARNING

IMPORTANT : KTX and KTY must have the same k-tables structure, the same number of columns, and the same column weights.

Author(s)

Jean Thioulouse <jean.thioulouse@univ-lyon1.fr>

References

Thioulouse J., Simier M. and Chessel D. (2004). Simultaneous analysis of a sequence of paired ecological tables. *Ecology* **85**, 272-283..

Simier, M., Blanc L., Pellegrin F., and Nandris D. (1999). Approche simultanée de K couples de tableaux : Application a l'étude des relations pathologie végétale - environnement. *Revue de Statistique Appliquée*, **47**, 31-46.

Examples

```

data(meau)
wit1 <- withinpca(meau$env, meau$design$season, scan = FALSE, scal = "total")
spepca <- dudi.pca(meau$spe, scale = FALSE, scan = FALSE, nf = 2)
wit2 <- wca(spepca, meau$design$season, scan = FALSE, nf = 2)
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5", "S6"), 4))
kta2 <- ktab.within(wit2, colnames = rep(c("S1", "S2", "S3", "S4", "S5", "S6"), 4))
statico1 <- statico(kta1, kta2, scan = FALSE)
plot(statico1)
kplot(statico1)

```

statis

STATIS, a method for analysing K-tables

Description

performs a STATIS analysis of a ktab object.

Usage

```

statis(X, scannf = TRUE, nf = 3, tol = 1e-07)
## S3 method for class 'statis'
plot(x, xax = 1, yax = 2, option = 1:4, ...)
## S3 method for class 'statis'
print(x, ...)

```

Arguments

X	an object of class 'ktab'
scannf	a logical value indicating whether the number of kept axes for the compromise should be asked
nf	if scannf FALSE, an integer indicating the number of kept axes for the compromise
tol	a tolerance threshold to test whether the distance matrix is Euclidean : an eigenvalue is considered positive if it is larger than $-tol * \lambda_1$ where λ_1 is the largest eigenvalue
x	an object of class 'statis'
xax, yax	the numbers of the x-axis and the y-axis
option	an integer between 1 and 4, otherwise the 4 components of the plot are displayed
...	further arguments passed to or from other methods

Value

statis returns a list of class 'statis' containing :

RV	a matrix with the all RV coefficients
RV.eig	a numeric vector with all the eigenvalues
RV.coo	a data frame with the array scores
tab.names	a vector of characters with the names of the arrays
RV.tabw	a numeric vector with the array weights
C.nf	an integer indicating the number of kept axes
C.rank	an integer indicating the rank of the analysis
C.li	a data frame with the row coordinates
C.Co	a data frame with the column coordinates
C.T4	a data frame with the principal vectors (for each table)
TL	a data frame with the factors (not used)
TC	a data frame with the factors for Co
T4	a data frame with the factors for T4

Author(s)

Daniel Chessel

References

Lavit, C. (1988) *Analyse conjointe de tableaux quantitatifs*, Masson, Paris.

Lavit, C., Escoufier, Y., Sabatier, R. and Traissac, P. (1994) The ACT (Statis method). *Computational Statistics and Data Analysis*, **18**, 97–119.

Examples

```
data(jv73)
kta1 <- ktab.within(withinpca(jv73$morpho, jv73$fac.riv, scann = FALSE))
statis1 <- statis(kta1, scann = FALSE)
plot(statis1)

dudi1 <- dudi.pca(jv73$poi, scann = FALSE, scal = FALSE)
wit1 <- wca(dudi1, jv73$fac.riv, scann = FALSE)
kta3 <- ktab.within(wit1)
data(jv73)
statis3 <- statis(kta3, scann = FALSE)
plot(statis3)

s.arrow(statis3$C.li, cgrid = 0)

kplot(statis3, traj = TRUE, arrow = FALSE, unique = TRUE,
      clab = 0, csub = 3, cpoi = 3)
statis3
```

 steppe

Transect in the Vegetation

Description

This data set gives the presence-absence of 37 species on 515 sites.

Usage

```
data(steppe)
```

Format

steppe is a list of 2 components.

tab is a data frame with 512 rows (sites) and 37 variables (species) in presence-absence.

esp.names is a vector of the species names.

Source

Estève, J. (1978) Les méthodes d'ordination : éléments pour une discussion. in J. M. Legay and R. Tomassone, editors. *Biométrie et Ecologie*, Société Française de Biométrie, Paris, 223–250.

Examples

```
par(mfrow = c(3,1))
data(steppe)
w1 <- col(as.matrix(steppe$tab[,1:15]))
w1 <- as.numeric(w1[steppe$tab[,1:15] > 0])
w2 <- row(as.matrix(steppe$tab[,1:15]))
w2 <- as.numeric(w2[steppe$tab[,1:15] > 0])
plot(w2, w1, pch = 20)
plot(dudi.pca(steppe$tab, scan = FALSE, scale = FALSE)$li[,1],
      pch = 20, ylab = "PCA", xlab = "", type = "b")
plot(dudi.coa(steppe$tab, scan = FALSE)$li[,1], pch = 20,
      ylab = "COA", xlab = "", type = "b")
par(mfrow = c(1,1))
```

 supcol

Projections of Supplementary Columns

Description

performs projections of supplementary columns.

Usage

```

supcol(x, ...)
## S3 method for class 'dudi'
supcol(x, Xsup, ...)
## S3 method for class 'coa'
supcol(x, Xsup, ...)

```

Arguments

x	an object used to select a method
Xsup	an array with the supplementary columns (Xsup and x\$tab have the same row number)
...	further arguments passed to or from other methods

Details

If `supcol.dudi` is used, the column vectors of `Xsup` are projected without prior modification onto the principal components of `dudi` with the scalar product associated to the row weightings of `dudi`.

Value

A list of two components:

tabsup	data frame containing the array with the supplementary columns transformed or not
cosup	data frame containing the coordinates of the supplementary projections

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

Examples

```

data(rpjdl)
rpjdl.coa <- dudi.coa(rpjdl$fau, scan = FALSE, nf = 4)
rpjdl.coa$co[1:3,]
supcol (rpjdl.coa, rpjdl$fau[,1:3])$cosup # the same

data(doubs)
dudi1 <- dudi.pca(doubs$fish, scal = FALSE, scan = FALSE)
s.arrow(dudi1$co)
s.arrow(supcol(dudi1, data.frame(scalewt(doubs$env))))$cosup,
  add.p = TRUE, clab = 2)
symbols(0, 0, circles = 1, inches = FALSE, add = TRUE)

```

suprow

Projections of Supplementary Rows

Description

performs projections of supplementary rows.

Usage

```
## S3 method for class 'coa'  
suprow(x, Xsup, ...)  
## S3 method for class 'dudi'  
suprow(x, Xsup, ...)  
## S3 method for class 'pca'  
suprow(x, Xsup, ...)
```

Arguments

x	an object of class dudi
Xsup	an array with the supplementary rows (Xsup and x\$tab have the same column number)
...	further arguments passed to or from other methods

Details

If `suprow.dudi` is used, the column vectors of `Xsup` are projected without prior modifications onto the principal components of `dudi` with the scalar product associated to the row weightings of `dudi`.

Value

returns a data frame containing the coordinates of the supplementary projections

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Gower, J. C. (1967) Multivariate analysis and multivariate geometry. *The statistician*, **17**, 13–28.

Examples

```

data(euro123)
par(mfrow = c(2,2))
w <- euro123[[2]]
dudi1 <- dudi.pca(w, scal = FALSE, scan = FALSE)
s.arrow(dudi1$c1, sub = "Classical", possub = "bottomright", csub = 2.5)
s.label(suprow(dudi1,w), add.plot = TRUE, clab = 0.75)

s.arrow(dudi1$c1,
        sub = "Without centring", possub = "bottomright", csub = 2.5)
s.label(suprow(dudi1,w), clab = 0.75, add.plot = TRUE)

triangle.plot(w, clab = 0.75, label = row.names(w), scal = FALSE)
triangle.plot(w, clab = 0.75, label = row.names(w), scal = TRUE)

data(rpjdl)
rpjdl.coa <- dudi.coa(rpjdl$fau, scann = FALSE, nf = 4)
rpjdl.coa$li[1:3,]
suprow(rpjdl.coa,rpjdl$fau[1:3,])$lisup # the same

data(deug)
deug.dudi <- dudi.pca(df = deug$tab, center = deug$cent,
                    scale = FALSE, scannf = FALSE)
suprow(deug.dudi, deug$tab[1:3,])$lisup # the supplementary individuals are centered
deug.dudi$li[1:3,] # the same

```

symbols.phylog	<i>Representation of a quantitative variable in front of a phylogenetic tree</i>
----------------	--

Description

symbols.phylog draws the phylogenetic tree and represents the values of the variable by symbols (squares or circles) which size is proportional to value. White symbols correspond to values which are below the mean, and black symbols correspond to values which are over.

Usage

```

symbols.phylog(phylog, circles, squares, csize = 1, clegend = 1,
              sub = "", csub = 1, possub = "topleft")

```

Arguments

phylog	an object of class phylog
circles	a vector giving the radii of the circles
squares	a vector giving the length of the sides of the squares
csize	a size coefficient for symbols
clegend	a character size for the legend used by par("cex")*clegend

sub a string of characters to be inserted as legend
 csub a character size for the legend, used with `par("cex")*csub`
 possub a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")

Author(s)

Daniel Chessel
 Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

See Also

[table.phylog](#) and [dotchart.phylog](#) for many variables

Examples

```
data(mjrochet)
mjrochet.phy <- newick2phylog(mjrochet$tre)
tab0 <- data.frame(scalewt(log(mjrochet$tab)))
par(mfrow=c(3,2))
for (j in 1:6) {
  w <- tab0[,j]
  symbols.phylog(phylog = mjrochet.phy, w, csi = 1.5, cleg = 1.5,
    sub = names(tab0)[j], csub = 3)
}
par(mfrow=c(1,1))
```

 syndicats

Two Questions asked on a Sample of 1000 Respondents

Description

This data set is extracted from an opinion poll (period 1970-1980) on 1000 respondents.

Usage

```
data(syndicats)
```

Format

The syndicats data frame has 5 rows and 4 columns.
 "Which politic family are you agreeing about ?" has 5 response items : extgauche (extreme left) left center right and extdroite (extreme right)
 "What do you think of the trade importance ?" has 4 response items : trop (too important) adequate insufficient nesaispas (no opinion)

Source

unknown

Examples

```
data(syndicats)
par(mfrow = c(1,2))
dudi1 <- dudi.coa(syndicats, scan = FALSE)
score (dudi1, 1, TRUE)
score (dudi1, 1, FALSE)
```

t3012

Average temperatures of 30 French cities

Description

This data set gives the average temperatures of 30 French cities during 12 months.

Usage

```
data(t3012)
```

Format

t3012 is a list of 3 objects:

xy is a data frame with 30 rows (cities) and 2 coordinates (x,y).

temp is a data frame with 30 rows (cities) and 12 columns (months). Each column contains the average temperature in tenth of degree Celsius.

contour is a data frame with 4 columns (x1,y1,x2,y2) for the contour display of France.

Source

Besse, P. (1979) *Etude descriptive d'un processus; approximation, interpolation*. Thèse de troisième cycle, Université Paul Sabatier, Toulouse.

Examples

```
data(t3012)
data(elec88)
area.plot(elec88$area)
s.arrow(t3012$xy, ori = as.numeric(t3012$xy["Paris",]),
  add.p = TRUE)
```

table.cont

*Plot of Contingency Tables***Description**

presents a graph for viewing contingency tables.

Usage

```
table.cont(df, x = 1:ncol(df), y = 1:nrow(df),
  row.labels = row.names(df), col.labels = names(df),
  clabel.row = 1, clabel.col = 1, abmean.x = FALSE, abline.x = FALSE,
  abmean.y = FALSE, abline.y = FALSE, csize = 1, clegend = 0, grid = TRUE)
```

Arguments

df	a data frame with only positive or null values
x	a vector of values to position the columns
y	a vector of values to position the rows
row.labels	a character vector for the row labels
col.labels	a character vector for the column labels
clabel.row	a character size for the row labels
clabel.col	a character size for the column labels
abmean.x	a logical value indicating whether the column conditional means should be drawn
abline.x	a logical value indicating whether the regression line of y onto x should be plotted
abmean.y	a logical value indicating whether the row conditional means should be drawn
abline.y	a logical value indicating whether the regression line of x onto y should be plotted
csize	a coefficient for the square size of the values
clegend	if not NULL, a character size for the legend used with <code>par("cex")*clegend</code>
grid	a logical value indicating whether a grid in the background of the plot should be drawn

Author(s)

Daniel Chessel

Examples

```

data(chats)
chatsw <- data.frame(t(chats))
chatscoa <- dudi.coa(chatsw, scann = FALSE)
par(mfrow = c(2,2))
table.cont(chatsw, abmean.x = TRUE, csi = 2, abline.x = TRUE,
           clabel.r = 1.5, clabel.c = 1.5)
table.cont(chatsw, abmean.y = TRUE, csi = 2, abline.y = TRUE,
           clabel.r = 1.5, clabel.c = 1.5)
table.cont(chatsw, x = chatscoa$c1[,1], y = chatscoa$l1[,1],
           abmean.x = TRUE, csi = 2, abline.x = TRUE, clabel.r = 1.5, clabel.c = 1.5)
table.cont(chatsw, x = chatscoa$c1[,1], y = chatscoa$l1[,1],
           abmean.y = TRUE, csi = 2, abline.y = TRUE, clabel.r = 1.5, clabel.c = 1.5)
par(mfrow = c(1,1))

## Not run:
data(rpjdl)
w <- data.frame(t(rpjdl$fau))
wcoa <- dudi.coa(w, scann = FALSE)
table.cont(w, abmean.y = TRUE, x = wcoa$c1[,1], y = rank(wcoa$l1[,1]),
           csi = 0.2, clabel.c = 0, row.labels = rpjdl$lalab, clabel.r = 0.75)

## End(Not run)

```

table.dist

Graph Display for Distance Matrices

Description

presents a graph for viewing distance matrices.

Usage

```
table.dist(d, x = 1:(attr(d, "Size")), labels = as.character(x),
          clabel = 1, csize = 1, grid = TRUE)
```

Arguments

d	an object of class dist
x	a vector of the row and column positions
labels	a vector of strings of characters for the labels
clabel	a character size for the labels
csize	a coefficient for the circle size
grid	a logical value indicating whether a grid in the background of the plot should be drawn

Author(s)

Daniel Chessel

Examples

```
data(eurodist)
table.dist(eurodist, labels = attr(eurodist, "Labels"))
```

table.paint

Plot of the arrays by grey levels

Description

presents a graph for viewing the numbers of a table by grey levels.

Usage

```
table.paint(df, x = 1:ncol(df), y = nrow(df):1,
            row.labels = row.names(df), col.labels = names(df),
            clabel.row = 1, clabel.col = 1, csize = 1, clegend = 1)
```

Arguments

df	a data frame
x	a vector of values to position the columns, used only for the ordered values
y	a vector of values to position the rows, used only for the ordered values
row.labels	a character vector for the row labels
col.labels	a character vector for the column labels
clabel.row	a character size for the row labels
clabel.col	a character size for the column labels
csize	if 'clegend' not NULL, a coefficient for the legend size
clegend	a character size for the legend, otherwise no legend

Author(s)

Daniel Chessel

Examples

```

data(rpjd1)
X <- data.frame(t(rpjd1$fau))
Y <- data.frame(t(rpjd1$mil))
layout(matrix(c(1,2,2,2,1,2,2,2,1,2,2,2,1,2,2,2), 4, 4))
coa1 <- dudi.coa(X, scan = FALSE)
x <- rank(coa1$co[,1])
y <- rank(coa1$li[,1])
table.paint(Y, x = x, y = 1:8, clabel.c = 0, cleg = 0)
abline(v = 114.9, lwd = 3, col = "red")
abline(v = 66.4, lwd = 3, col = "red")
table.paint(X, x = x, y = y, clabel.c = 0, cleg = 0,
  row.lab = paste(" ", row.names(X), sep = ""))
abline(v = 114.9, lwd = 3, col = "red")
abline(v = 66.4, lwd = 3, col = "red")

```

table.phylog

Plot arrays in front of a phylogenetic tree

Description

This function gives a graphical display for viewing the numbers of a table by square sizes in front of the corresponding phylogenetic tree.

Usage

```

table.phylog(df, phylog, x = 1:ncol(df), f.phylog = 0.5,
  labels.row = gsub("[_]", " ", row.names(df)), clabel.row = 1,
  labels.col = names(df), clabel.col = 1,
  labels.nod = names(phylog$nodes), clabel.nod = 0, cleaves = 1,
  cnodes = 1, csize = 1, grid = TRUE, clegend = 0.75)

```

Arguments

df : a data frame or a matrix

phylog : an object of class 'phylog'

x : a vector of values to position the columns

f.phylog : a size coefficient for tree size (a parameter to draw the tree in proportion to leaves labels)

labels.row : a vector of strings of characters for row labels

clabel.row : a character size for the leaves labels, used with `par("cex")*clabel.row`. If zero, no row labels are drawn

labels.col : a vector of strings of characters for columns labels

clabel.col : a character size for the leaves labels, used with `par("cex")*clabel.col`. If zero, no column labels are drawn

labels.nod	: a vector of strings of characters for the nodes labels
clabel.nod	: a character size for the nodes labels, used with <code>par("cex")*clabel.nodes</code> . If zero, no nodes labels are drawn
cleaves	: a character size for plotting the points that represent the leaves, used with <code>par("cex")*cleaves</code> . If zero, no points are drawn
cnodes	: a character size for plotting the points that represent the nodes, used with <code>par("cex")*cnodes</code> . If zero, no points are drawn
csize	: a size coefficient for symbols
grid	: a logical value indicating whether the grid should be plotted
clegend	: a character size for the legend (if 0, no legend)

Details

The function verifies that `sort(row.names(df))==sort(names(phylog$leaves))`. If `df` is a matrix the function uses `as.data.frame(df)`.

Author(s)

Daniel Chessel
Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>

See Also

[symbols.phylog](#) for one variable

Examples

```
## Not run:
data(newick.eg)
w.phy <- newick2phylog(newick.eg[[9]])
w.tab <- data.frame(matrix(rnorm(620), 31, 20))
row.names(w.tab) <- sort(names(w.phy$leaves))
table.phylog(w.tab, w.phy, csi = 1.5, f = 0.5,
  clabel.n = 0.75, clabel.c = 0.5)

## End(Not run)
```

table.value

Plot of the Arrays

Description

presents a graph for viewing the numbers of a table by square sizes.

Usage

```
table.value(df, x = 1:ncol(df), y = nrow(df):1,
            row.labels = row.names(df), col.labels = names(df), clabel.row = 1,
            clabel.col = 1, csize = 1, clegend = 1, grid = TRUE)
```

Arguments

df	a data frame
x	a vector of values to position the columns
y	a vector of values to position the rows
row.labels	a character vector for the row labels
col.labels	a character vector for the column labels
clabel.row	a character size for the row labels
clabel.col	a character size for the column labels
csize	a coefficient for the square size of the values
clegend	a character size for the legend (if 0, no legend)
grid	a logical value indicating whether the grid should be plotted

Author(s)

Daniel Chessel

Examples

```
data(olympic)
w <- olympic$tab
w <- data.frame(scale(w))
wpca <- dudi.pca(w, scann = FALSE)
par(mfrow = c(1,3))
table.value(w, csi = 2, clabel.r = 2, clabel.c = 2)
table.value(w, y = rank(wpca$li[,1]), x = rank(wpca$co[,1]), csi = 2,
            clabel.r = 2, clabel.c = 2)
table.value(w, y = wpca$li[,1], x = wpca$co[,1], csi = 2,
            clabel.r = 2, clabel.c = 2)
par(mfrow = c(1,1))
```

tarentaise

Mountain Avifauna

Description

This data set gives informations between sites, species, environmental and biological variables.

Usage

```
data(tarentaise)
```

Format

`tarentaise` is a list of 5 components.

`ecol` is a data frame with 376 sites and 98 bird species.

`frnames` is a vector of the 98 French names of the species.

`alti` is a vector giving the altitude of the 376 sites in m.

`envir` is a data frame with 14 environmental variables.

`traits` is a data frame with 29 biological variables of the 98 species.

Details

The attribute `col.blocks` of the data frame `tarentaise$traits` indicates it is composed of 6 units of variables.

Source

Original data from Hubert Tournier, University of Savoie and Philippe Lebreton, University of Lyon 1.

References

Lebreton, P., Tournier H. and Lebreton J. D. (1976) Etude de l'avifaune du Parc National de la Vanoise VI Recherches d'ordre quantitatif sur les Oiseaux forestiers de Vanoise. *Travaux Scientifiques du parc National de la vanoise*, **7**, 163–243.

Lebreton, Ph. and Martinot, J.P. (1998) Oiseaux de Vanoise. Guide de l'ornithologue en montagne. *Libris*, Grenoble. 1–240.

Lebreton, Ph., Lebrun, Ph., Martinot, J.P., Miquet, A. and Tournier, H. (1999) Approche écologique de l'avifaune de la Vanoise. *Travaux scientifiques du Parc national de la Vanoise*, **21**, 7–304.

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps038.pdf> (in French).

Examples

```
data(tarentaise)
coa1 <- dudi.coa(tarentaise$ecol, sca = FALSE, nf = 2)
s.class(coa1$li, tarentaise$envir$alti, wt = coa1$lw)
## Not run:
acm1 <- dudi.acm(tarentaise$envir, sca = FALSE, nf = 2)
s.class(acm1$li, tarentaise$envir$alti)

## End(Not run)
```

`taxo.eg`*Examples of taxonomy*

Description

This data sets contains two taxonomies.

Usage

```
data(taxo.eg)
```

Format

`taxo.eg` is a list containing the 2 following objects:

`taxo.eg[[1]]` is a data frame with 15 species and 3 columns.

`taxo.eg[[2]]` is a data frame with 40 species and 2 columns.

Details

Variables of the first data frame are : `genre` (a factor `genre` with 8 levels), `famille` (a factor `familiy` with 5 levels) and `ordre` (a factor `order` with 2 levels).

Variables of the second data frame are : `gen`(a factor `genre` with 29 levels), `fam` (a factor `family` with 19 levels).

Examples

```
data(taxo.eg)
taxo.eg[[1]]
as.taxo(taxo.eg[[1]])
class(taxo.eg[[1]])
class(as.taxo(taxo.eg[[1]]))

tax.phy <- taxo2phylog(as.taxo(taxo.eg[[1]]), add.tools = TRUE)
plot(tax.phy, clabel.l=1)

par(mfrow = c(1,2))
table.phylog(tax.phy$Bindica, tax.phy)
table.phylog(tax.phy$Bscores, tax.phy)
par(mfrow = c(1,1))

radial.phylog(taxo2phylog(as.taxo(taxo.eg[[2]])))
```

`testdim`*Function to perform a test of dimensionality*

Description

This functions allow to test for the number of axes in multivariate analysis. The procedure is only implemented for principal component analysis on correlation matrix. The procedure is based on the computation of the RV coefficient.

Usage

```
testdim(dudi, ...)  
## S3 method for class 'pca'  
testdim(dudi, nrepet = 99, nbax = dudi$rank, alpha = 0.05, ...)
```

Arguments

<code>dudi</code>	a duality diagram (an object of class <code>dudi</code>)
<code>nrepet</code>	the number of repetitions for the permutation procedure
<code>nbax</code>	the number of axes to be tested, by default all axes
<code>alpha</code>	the significance level
<code>...</code>	other arguments

Value

An object of the class `krandtest`. It contains also:

<code>nb</code>	The estimated number of axes to keep
<code>nb.cor</code>	The number of axes to keep estimated using a sequential Bonferroni procedure

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr>

References

Dray, S. (2007) On the number of principal components: A test of dimensionality based on measurements of similarity between matrices. *Computational Statistics and Data Analysis*, in press.

See Also

[dudi.pca](#), [RV.rtest](#)

Examples

```

tab <- data.frame(matrix(rnorm(200),20,10))
pca1 <- dudi.pca(tab,scannf=FALSE)
test1 <- testdim(pca1)
test1
test1$nb
test1$nb.cor
data(doubs)
pca2 <- dudi.pca(doubs$env,scannf=FALSE)
test2 <- testdim(pca2)
test2
test2$nb
test2$nb.cor

```

tintoodiel

Tinto and Odiel estuary geochemistry

Description

This data set contains informations about geochemical characteristics of heavy metal pollution in surface sediments of the Tinto and Odiel river estuary (south-western Spain).

Usage

```
data(tintoodiel)
```

Format

tintoodiel is a list containing the following objects :

xy : a data frame that contains spatial coordinates of the 52 sites

tab : a data frame with 12 columns (concentration of heavy metals) and 52 rows (sites)

neig : an object of class neig

Source

Borrego, J., Morales, J.A., de la Torre, M.L. and Grande, J.A. (2002) Geochemical characteristics of heavy metal pollution in surface sediments of the Tinto and Odiel river estuary (south-western Spain). *Environmental Geology*, **41**, 785–796.

Examples

```

data(tintoodiel)

## Not run:
if (require(pixmap, quiet = TRUE)){
  estuary.pnm <- read.pnm(system.file("pictures/tintoodiel.pnm",
    package = "ade4"))
}

```

```

s.label(tintoodiel$xy, pixmap = estuary.pnm, neig = tintoodiel$neig,
        clab = 0, cpoi = 2, cneig = 3, addax = FALSE, cgrid = 0, grid = FALSE)
}

## End(Not run)

estuary.pca <- dudi.pca(tintoodiel$tab, scan = FALSE, nf = 4)
if (require(maptools, quiet = TRUE) & require(spdep, quiet = TRUE)) {
  estuary.listw <- nb2listw(neig2nb(tintoodiel$neig))
  estuary.pca.ms <- multispati(estuary.pca, estuary.listw, scan = FALSE,
                              nfposi = 3, nfnega = 2)
  summary(estuary.pca.ms)
  par(mfrow = c(1,2))
  barplot(estuary.pca$eig)
  barplot(estuary.pca.ms$eig)
  par(mfrow = c(1,1))
}

```

tithonia

Phylogeny and quantitative traits of flowers

Description

This data set describes the phylogeny of 11 flowers as reported by Morales (2000). It also gives morphologic and demographic traits corresponding to these 11 species.

Usage

```
data(tithonia)
```

Format

tithonia is a list containing the 2 following objects :

tre is a character string giving the phylogenetic tree in Newick format.

tab is a data frame with 11 species and 14 traits (6 morphologic traits and 8 demographic).

Details

Variables of tithonia\$tab are the following ones :

- morho1: is a numeric vector that describes the seed size (mm)
- morho2: is a numeric vector that describes the flower size (mm)
- morho3: is a numeric vector that describes the female leaf size (cm)
- morho4: is a numeric vector that describes the head size (mm)
- morho5: is a integer vector that describes the number of flowers per head
- morho6: is a integer vector that describes the number of seeds per head
- demo7: is a numeric vector that describes the seedling height (cm)
- demo8: is a numeric vector that describes the growth rate (cm/day)
- demo9: is a numeric vector that describes the germination time

demo10: is a numeric vector that describes the establishment (per cent)
demo11: is a numeric vector that describes the viability (per cent)
demo12: is a numeric vector that describes the germination (per cent)
demo13: is a integer vector that describes the resource allocation
demo14: is a numeric vector that describes the adult height (m)

Source

Data were obtained from Morales, E. (2000) Estimating phylogenetic inertia in *Tithonia* (Asteraceae) : a comparative approach. *Evolution*, **54**, 2, 475–484.

Examples

```
data(tithonia)
phy <- newick2phylog(tithonia$tre)
tab <- log(tithonia$tab + 1)
table.phylog(scalewt(tab), phy)
gearymoran(phy$Wmat, tab)
gearymoran(phy$Amat, tab)
```

tortues

Morphological Study of the Painted Turtle

Description

This data set gives a morphological description (4 characters) of 48 turtles.

Usage

```
data(tortues)
```

Format

a data frame with 48 rows and 4 columns (length (mm), maximum width(mm), height (mm), gender).

Source

Jolicoeur, P. and Mosimann, J. E. (1960) Size and shape variation in the painted turtle. A principal component analysis. *Growth*, **24**, 339–354.

Examples

```

data(tortues)
xyz <- as.matrix(tortues[,1:3])
ref <- -svd(xyz)$u[,1]
pch0 <- c(1,20)[as.numeric(tortues$sex)]
plot(ref, xyz[,1], ylim = c(40,180), pch = pch0)
abline(lm(xyz[,1]~ -1 + ref))
points(ref,xyz[,2], pch = pch0)
abline(lm(xyz[,2]~ -1 + ref))
points(ref,xyz[,3], pch = pch0)
abline(lm(xyz[,3]~ -1 + ref))

```

toxicity

Homogeneous Table

Description

This data set gives the toxicity of 7 molecules on 17 targets expressed in $-\log(\text{mol/liter})$

Usage

```
data(toxicity)
```

Format

toxicity is a list of 3 components.

tab is a data frame with 7 columns and 17 rows

species is a vector of the names of the species in the 17 targets

chemicals is a vector of the names of the 7 molecules

Source

Devillers, J., Thioulouse, J. and Karcher W. (1993) Chemometrical Evaluation of Multispecies-Multichemical Data by Means of Graphical Techniques Combined with Multivariate Analyses. *Ecotoxicology and Environmental Safety*, **26**, 333–345.

Examples

```

data(toxicity)
table.paint(toxicity$tab, row.lab = toxicity$species,
            col.lab = toxicity$chemicals)

table.value(toxicity$tab, row.lab = toxicity$species,
            col.lab = toxicity$chemicals)

```

triangle.class	<i>Triangular Representation and Groups of points</i>
----------------	---

Description

Function to plot triangular data (i.e. dataframe with 3 columns of positive or null values) and a partition

Usage

```
triangle.class(ta, fac, col = rep(1, length(levels(fac))),
  wt = rep(1, length(fac)), cstar = 1, cellipse = 0, axesell = TRUE,
  label = levels(fac), clabel = 1, cpoint = 1, pch = 20, draw.line = TRUE,
  addaxes = FALSE, addmean = FALSE, labeltriangle = TRUE, sub = "", csub = 1,
  possub = "bottomright", show.position = TRUE, scale = TRUE, min3 = NULL,
  max3 = NULL)
```

Arguments

ta	a data frame with 3 columns of null or positive numbers
fac	a factor of length the row number of ta
col	a vector of color for showing the groups
wt	a vector of row weighting for the computation of the gravity centers by class
cstar	a character size for plotting the stars between 0 (no stars) and 1 (complete star) for a line linking a point to the gravity center of its belonging class.
cellipse	a positive coefficient for the inertia ellipse size
axesell	a logical value indicating whether the ellipse axes should be drawn
label	a vector of strings of characters for the labels of gravity centers
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
pch	if <code>cpoint > 0</code> , an integer specifying the symbol or the single character to be used in plotting points
draw.line	a logical value indicating whether the triangular lines should be drawn
addaxes	a logical value indicating whether the axes should be plotted
addmean	a logical value indicating whether the mean point should be plotted
labeltriangle	a logical value indicating whether the variable labels of ta should be drawn on the triangular sides
sub	a string of characters for the graph title
csub	a character size for plotting the graph title

possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
show.position	a logical value indicating whether the sub-triangle containing the data should be put back in the total triangle
scale	a logical value for the graph representation : the total triangle (FALSE) or the sub-triangle (TRUE)
min3	if not NULL, a vector with 3 numbers between 0 and 1
max3	if not NULL, a vector with 3 numbers between 0 and 1. Let notice that min3+max3 must equal c(1,1,1)

Author(s)

Daniel Chessel

Examples

```

data(euro123)
par(mfrow = c(2,2))
x = rbind.data.frame(euro123$in78, euro123$in86, euro123$in97)
triangle.plot(x)
triangle.class(x, as.factor(rep("G",36)), csta = 0.5, cell = 1)
triangle.class(x, euro123$plan$an)
triangle.class(x, euro123$plan$pays)
triangle.class(x, euro123$plan$an, cell = 1, axesell = TRUE)
triangle.class(x, euro123$plan$an, cell = 0, csta = 0,
  col = c("red", "green", "blue"), axesell = TRUE, clab = 2, cpoi = 2)
triangle.class(x, euro123$plan$an, cell = 2, csta = 0.5,
  axesell = TRUE, clab = 1.5)
triangle.class(x, euro123$plan$an, cell = 0, csta = 1, scale = FALSE,
  draw.line = FALSE, show.posi = FALSE)

```

triangle.plot

*Triangular Plotting***Description**

Graphs for a dataframe with 3 columns of positive or null values

triangle.plot is a scatterplot

triangle.biplot is a paired scatterplots

triangle.posipoint, triangle.param, add.position.triangle are utilities functions.

Usage

```

triangle.plot(ta, label = as.character(1:nrow(ta)), clabel = 0,
  cpoint = 1, draw.line = TRUE, addaxes = FALSE, addmean = FALSE,
  labeltriangle = TRUE, sub = "", csub = 0, possub = "topright",

```



```

    show.position = TRUE, scale = TRUE, min3 = NULL, max3 = NULL,
    box = FALSE)
triangle.biplot (ta1, ta2, label = as.character(1:nrow(ta1)),
    draw.line = TRUE, show.position = TRUE, scale = TRUE)

```

Arguments

ta, ta1, ta2,	data frame with three columns, will be transformed in percentages by rows
label	a vector of strings of characters for the point labels
clabel	if not NULL, a character size for the labels, used with <code>par("cex")*clabel</code>
cpoint	a character size for plotting the points, used with <code>par("cex")*cpoint</code> . If zero, no points are drawn
draw.line	a logical value indicating whether the lines into the triangle should be drawn
addaxes	a logical value indicating whether the principal axes should be drawn
addmean	a logical value indicating whether the mean should be plotted
labeltriangle	a logical value indicating whether the variable names should be wrote
sub	a string of characters to be inserted as legend
csub	a character size for the legend, used with <code>par("cex")*csub</code>
possub	a string of characters indicating the sub-title position ("topleft", "topright", "bottomleft", "bottomright")
show.position	a logical value indicating whether the used triangle should be shown in the complete one
scale	a logical value indicating whether the smaller equilateral triangle containing the plot should be used
min3	If scale is FALSE, a vector of three values for the minima e.g. <code>c(0.1,0.1,0.1)</code> can be used
max3	If scale is FALSE a vector of three values for the maxima e.g. <code>c(0.9,0.9,0.9)</code> can be used
box	a logical value indicating whether a box around the current plot should be drawn

Value

`triangle.plot` returns an invisible matrix containing the coordinates used for the plot. The graph can be supplemented in various ways.

Author(s)

Daniel Chessel

Examples

```

data (euro123)
tot <- rbind.data.frame(euro123$in78, euro123$in86, euro123$in97)
row.names(tot) <- paste(row.names(euro123$in78), rep(c(1,2,3), rep(12,3)),
    sep = "")

```

```

triangle.plot(tot, label = row.names(tot), clab = 1)

par(mfrow = c(2,2))
triangle.plot(euro123$in78, clab = 0, cpoi = 2, addmean = TRUE,
             show = FALSE)
triangle.plot(euro123$in86, label = row.names(euro123$in78), clab = 0.8)
triangle.biplot(euro123$in78, euro123$in86)
triangle.plot(rbind.data.frame(euro123$in78, euro123$in86), clab = 1,
             addaxes = TRUE, sub = "Principal axis", csub = 2, possub = "topright")

triangle.plot(euro123[[1]], min3 = c(0,0.2,0.3), max3 = c(0.5,0.7,0.8),
             clab = 1, label = row.names(euro123[[1]]), addax = TRUE)
triangle.plot(euro123[[2]], min3 = c(0,0.2,0.3), max3 = c(0.5,0.7,0.8),
             clab = 1, label = row.names(euro123[[1]]), addax = TRUE)
triangle.plot(euro123[[3]], min3 = c(0,0.2,0.3), max3 = c(0.5,0.7,0.8),
             clab = 1, label = row.names(euro123[[1]]), addax = TRUE)
triangle.plot(rbind.data.frame(euro123[[1]], euro123[[2]], euro123[[3]]))

wtriangleplot <- cbind.data.frame(a = runif(100), b = runif(100),
                                c = runif(100,4,5))
wtriangleplot <- triangle.plot(wtriangleplot)
points(wtriangleplot, col = "blue", cex = 2)
wtriangleplot <- colMeans(wtriangleplot)
points(wtriangleplot[1], wtriangleplot[2], pch = 20, cex = 3, col = "red")
rm(wtriangleplot)

```

trichometeo

Pair of Ecological Data

Description

This data set gives for trapping nights informations about species and meteorological variables.

Usage

```
data(trichometeo)
```

Format

trichometeo is a list of 3 components.

fau is a data frame with 49 rows (trapping nights) and 17 species.

meteo is a data frame with 49 rows and 11 meteorological variables.

cla is a factor of 12 levels for the definition of the consecutive night groups

Source

Data from P. Usseglio-Polatera

References

Usseglio-Polatera, P. and Auda, Y. (1987) Influence des facteurs météorologiques sur les résultats de piégeage lumineux. *Annales de Limnologie*, **23**, 65–79. (code des espèces p. 76)

See a data description at <http://pbil.univ-lyon1.fr/R/pps/pps034.pdf> (in French).

Examples

```
data(trichometeo)
faulog <- log(trichometeo$fau + 1)
pca1 <- dudi.pca(trichometeo$meteo, scan = FALSE)
niche1 <- niche(pca1, faulog, scan = FALSE)
s.label(niche1$ls, clab = 0)
s.distri(niche1$ls, faulog, clab = 0.6, add.p = TRUE,
         cell = 0, csta = 0.3)
s.arrow(7 * niche1$c1, clab = 1, add.p = TRUE)
```

ungulates

Phylogeny and quantitative traits of ungulates.

Description

This data set describes the phylogeny of 18 ungulates as reported by Pélabon et al. (1995). It also gives 4 traits corresponding to these 18 species.

Usage

```
data(ungulates)
```

Format

fission is a list containing the 2 following objects :

tre is a character string giving the phylogenetic tree in Newick format.

tab is a data frame with 18 species and 4 traits

Details

Variables of `ungulates$tab` are the following ones :

afbw: is a numeric vector that describes the adult female body weight (g)

mnw: is a numeric vector that describes the male neonatal weight (g)

fnw: is a numeric vector that describes the female neonatal weight (g)

ls: is a numeric vector that describes the litter size

Source

Data were obtained from Pélabon, C., Gaillard, J.M., Loison, A. and Portier, A. (1995) Is sex-biased maternal care limited by total maternal expenditure in polygynous ungulates? *Behavioral Ecology and Sociobiology*, **37**, 311–319.

Examples

```
data(ungulates)
ung.phy <- newick2phylog(ungulates$tre)
plot(ung.phy, clabel.l=1.25, clabel.n=0.75)
ung.x <- log(ungulates$tab[,1])
ung.y <- log((ungulates$tab[,2]+ungulates$tab[,3])/2)
names(ung.x) <- names(ung.phy$leaves)
names(ung.y) <- names(ung.x)
plot(ung.x, ung.y)
abline(lm(ung.y~ung.x))
symbols.phylog(ung.phy, ung.x-mean(ung.x))
dotchart.phylog(ung.phy, ung.x, cle=1.5, cno=1.5, cdot=1)
orthogram(ung.x, ung.phy$Bscores, nrep=9999)
ung.z <- residuals(lm(ung.y~ung.x))
names(ung.z) <- names(ung.phy$leaves)
dotchart.phylog(ung.phy, ung.z, cle=1.5, cno=1.5, cdot=1, ceti=0.75)
orthogram(ung.z, ung.phy$Bscores, nrep=9999)
```

uniquewt.df

Elimination of Duplicated Rows in a Array

Description

An utility function to eliminate the duplicated rows in a array.

Usage

```
uniquewt.df(x)
```

Arguments

x a data frame which contains duplicated rows

Value

The function returns a y which contains once each duplicated row of x.
y is an attribut 'factor' which gives the number of the row of y in which each row of x is found
y is an attribut 'length.class' which gives the number of duplicates in x with an attribut of each row of y with an attribut

Author(s)

Daniel Chessel

Examples

```

data(ecomor)
forsub.r <- uniquewt.df(ecomor$forsub)
attr(forsub.r, "factor")
forsub.r[1,]
ecomor$forsub[126,] #idem

dudi.pca(ecomor$forsub, scale = FALSE, scann = FALSE)$eig
# [1] 0.36845 0.24340 0.15855 0.09052 0.07970 0.04490
w1 <- attr(forsub.r, "len.class") / sum(attr(forsub.r, "len.class"))
dudi.pca(forsub.r, row.w = w1, scale = FALSE, scann = FALSE)$eig
# [1] 0.36845 0.24340 0.15855 0.09052 0.07970 0.04490

```

variance.phylog	<i>The phylogenetic ANOVA</i>
-----------------	-------------------------------

Description

This function performs the variance analysis of a trait on eigenvectors associated to a phylogenetic tree.

Usage

```

variance.phylog(phylog, z, bynames = TRUE,
na.action = c("fail", "mean"))

```

Arguments

phylog	: an object of class phylog
z	: a numeric vector of the values corresponding to the variable
bynames	: if TRUE checks if z labels are the same as phylog leaves label, possibly in a different order. If FALSE the check is not made and z labels must be in the same order than phylog leaves label
na.action	: if 'fail' stops the execution of the current expression when z contains any missing value. If 'mean' replaces any missing values by mean(z)

Details

phylog\$A_{mat} defines a set of orthonormal vectors associated the each nodes of the phylogenetic tree.

phylog\$A_{dim} defines the dimension of the subspace **A** defined by the first phylog\$A_{dim} vectors of phylog\$A_{mat} that corresponds to phylogenetic inertia.

variance.phylog performs the linear regression of z on **A**.

Value

Returns a list containing

- lm : an object of class `lm` that corresponds to the linear regression of `z` on `A`.
- anova : an object of class `anova` that corresponds to the anova of the precedent model.
- smry : an object of class `anova` that is a summary of the precedent object.

Author(s)

Sébastien Ollier <ollier@biomserv.univ-lyon1.fr>
Daniel Chessel

References

- Grafen, A. (1989) The phylogenetic regression. *Philosophical Transactions of the Royal Society London B*, **326**, 119–156.
- Diniz-Filho, J. A. F., Sant’Ana, C.E.R. and Bini, L.M. (1998) An eigenvector method for estimating phylogenetic inertia. *Evolution*, **52**, 1247–1262.

See Also

[phylog](#), [lm](#)

Examples

```
data(njplot)
njplot.phy <- newick2phylog(njplot$tre)
variance.phylog(njplot.phy, njplot$tauxcg)
par(mfrow = c(1,2))
table.phylog(njplot.phy$Ascores, njplot.phy, clabel.row = 0,
  clabel.col = 0.1, clabel.nod = 0.6, csize = 1)
dotchart.phylog(njplot.phy, njplot$tauxcg, clabel.nodes = 0.6)
orthogram(njplot$tauxcg, njplot.phy$Ascores)
```

vegtf

Vegetation in Trois-Fontaines

Description

This data set contains abundance values (Braun-Blanquet scale) of 80 plant species for 337 sites. Data have been collected by Sonia Said and Francois Debias.

Usage

```
data(vegtf)
```

Format

`vegtf` is a list containing the following objects :

veg is a data.frame with the abundance values of 80 species (columns) in 337 sites (rows).

xy is a data.frame with the spatial coordinates of the sites.

area is data.frame (area) which define the boundaries of the study site.

nb is a neighborhood object (class nb defined in package spdep)

Source

Dray, S., Said, S. and Debias, F. (2008) Spatial ordination of vegetation data using a generalization of Wartenberg's multivariate spatial correlation. *Journal of vegetation science*, **19**, 45–56.

Examples

```
if (require(spdep, quiet=TRUE)){
  data(vegtf)
  coa1 <- dudi.coa(vegtf$veg,scannf=FALSE)
  ms.coa1 <- multispazi(coa1,listw=nb2listw(vegtf$nb),nfposi=2,nfnega=0,scannf=FALSE)
  summary(ms.coa1)
  plot(ms.coa1)
  par(mfrow=c(2,2))
  s.value(vegtf$xy,coa1$li[,1],area=vegtf$area,include.origin=FALSE)
  s.value(vegtf$xy,ms.coa1$li[,1],area=vegtf$area,include.origin=FALSE)
  s.label(coa1$c1)
  s.label(ms.coa1$c1)
}
```

 veuvage

Example for Centring in PCA

Description

The data come from the INSEE (National Institute of Statistics and Economical Studies). It is an array of widower percentages in relation with the age and the socioprofessional category.

Usage

```
data(veuvage)
```

Format

`veuvage` is a list of 2 components.

tab is a data frame with 37 rows (widowers) 6 columns (socio-professional categories)

age is a vector of the ages of the 37 widowers.

Details

The columns contain the socioprofessional categories:

- 1- Farmers, 2- Craftsmen, 3- Executives and higher intellectual professions,
- 4- Intermediate Professions, 5- Others white-collar workers and 6- Manual workers.

Source

unknown

Examples

```
data(veuvage)
par(mfrow = c(3,2))
for (j in 1:6) plot(veuvage$age, veuvage$tab[,j],
  xlab = "age", ylab = "pourcentage de veufs",
  type = "b", main = names(veuvage$tab)[j])
```

wca

Within-Class Analysis

Description

Performs a particular case of an Orthogonal Principal Component Analysis with respect to Instrumental Variables (orthopcaiv), in which there is only a single factor as covariable.

Usage

```
within(dudi, fac, scannf = TRUE, nf = 2)
## S3 method for class 'dudi'
wca(x, fac, scannf = TRUE, nf = 2, ...)
```

Arguments

dudi	a duality diagram, object of class <code>dudi</code> obtained from the functions <code>dudi.coa</code> , <code>dudi.pca</code> ,...
x	a duality diagram, object of class <code>dudi</code> from one of the functions <code>dudi.coa</code> , <code>dudi.pca</code> ,...
fac	a factor partitioning the rows of <code>dudi\$tab</code> in classes
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if <code>scannf</code> <code>FALSE</code> , an integer indicating the number of kept axes
...	further arguments passed to or from other methods

Value

Returns a list of the sub-class within in the class dudi

tab	a data frame containing the transformed data (subtraction of the class mean)
call	the matching call
nf	number of kept axes
rank	the rank of the analysis
ratio	percentage of within-class inertia
eig	a numeric vector containing the eigenvalues
lw	a numeric vector of row weights
cw	a numeric vector of column weights
tabw	a numeric vector of class weights
fac	the factor defining the classes
li	data frame row coordinates
l1	data frame row normed scores
co	data frame column coordinates
c1	data frame column normed scores
ls	data frame supplementary row coordinates
as	data frame inertia axis onto within axis

Note

To avoid conflict names with the base::within function, the function within is now deprecated and will be removed. Use the generic wca function instead.

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

- Benzécri, J. P. (1983) Analyse de l'inertie intra-classe par l'analyse d'un tableau de correspondances. *Les Cahiers de l'Analyse des données*, **8**, 351–358.
- Dolédec, S. and Chessel, D. (1987) Rythmes saisonniers et composantes stationnelles en milieu aquatique I- Description d'un plan d'observations complet par projection de variables. *Acta Oecologica, Oecologia Generalis*, **8**, 3, 403–426.

Examples

```

data(meaudret)
par(mfrow = c(2,2))
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)
s.traject(pca1$li, meaudret$design$site,
  sub = "Principal Component Analysis", csub = 1.5)
wit1 <- wca(pca1, meaudret$design$site, scan = FALSE, nf = 2)
s.traject(wit1$li, meaudret$design$site,
  sub = "Within site Principal Component Analysis", csub = 1.5)
s.corcircle (wit1$as)
par(mfrow = c(1,1))
plot(wit1)

```

wca.rfq

Within-Class RLQ analysis

Description

Performs a particular RLQ analysis where a partition of sites (rows of R) is taken into account. The within-class RLQ analysis search for linear combinations of traits and environmental variables of maximal covariance.

Usage

```

## S3 method for class 'rlq'
wca(x, fac, scannf = TRUE, nf = 2, ...)
## S3 method for class 'witrlq'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'witrlq'
print(x, ...)

```

Arguments

x	an object of class <code>rlq</code> (created by the <code>rlq</code> function) for the <code>wca.rfq</code> function. An object of class <code>witrlq</code> for the <code>print</code> and <code>plot</code> functions
fac	a factor partitioning the rows of R
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if <code>scannf</code> FALSE, an integer indicating the number of kept axes
xax	the column number for the x-axis
yax	the column number for the y-axis
...	further arguments passed to or from other methods

Value

The `wca.rfq` function returns an object of class `'betrlq'` (sub-class of `'dudi'`). See the outputs of the `print` function for more details.

Author(s)

Stephane Dray <stephane.drays@univ-lyon1.fr>

References

Wesuls, D., Oldeland, J. and Dray, S. (2012) Disentangling plant trait responses to livestock grazing from spatio-temporal variation: the partial RLQ approach. *Journal of Vegetation Science*, **23**, 98–113.

See Also

[rlq](#), [wca](#), [wca.rlq](#)

Examples

```
data(piosphere)
afcl <- dudi.coa(log(piosphere$veg + 1), scannf = FALSE)
acpR <- dudi.pca(piosphere$env, scannf = FALSE, row.w = afcl$lw)
acpQ <- dudi.hillsmith(piosphere$traits, scannf = FALSE, row.w =
  afcl$cw)
rlq1 <- rlq(acpR, afcl, acpQ, scannf = FALSE)

wrlq1 <- wca(rlq1, fac = piosphere$habitat, scannf = FALSE)
wrlq1
plot(wrlq1)
```

westafrica

Freshwater fish zoogeography in west Africa

Description

This data set contains informations about faunal similarities between river basins in West africa.

Usage

```
data(westafrica)
```

Format

westafrica is a list containing the following objects :

tab : a data frame with absence/presence of 268 species (rows) at 33 embouchures (columns)

spe.names : a vector of string of characters with the name of species

spe.binames : a data frame with the genus and species (columns) of the 256 species (rows)

riv.names : a vector of string of characters with the name of rivers

atlantic : a data frame with the coordinates of a polygon that represents the limits of atlantic (see example)

riv.xy : a data frame with the coordinates of embouchures

lines : a data frame with the coordinates of lines to complete the representation (see example)

cadre : a data frame with the coordinates of points used to make the representation (see example)

Source

Data provided by B. Hugueny <hugueny@biomserv.univ-lyon1.fr>.

Paugy, D., Traoré, K. and Diouf, P.F. (1994) Faune ichtyologique des eaux douces d’Afrique de l’Ouest. In *Diversité biologique des poissons des eaux douces et saumâtres d’Afrique. Synthèses géographiques*, Teugels, G.G., Guégan, J.F. and Albaret, J.J. (Editors). Annales du Musée Royal de l’Afrique Centrale, Zoologie, **275**, Tervuren, Belgique, 35–66.

Hugueny, B. (1989) *Biogéographie et structure des peuplements de Poissons d’eau douce de l’Afrique de l’ouest : approches quantitatives*. Thèse de doctorat, Université Paris 7.

References

Hugueny, B., and Lévêque, C. (1994) Freshwater fish zoogeography in west Africa: faunal similarities between river basins. *Environmental Biology of Fishes*, **39**, 365–380.

Examples

```
data(westafrica)

s.label(westafrica$cadre, xlim = c(30,500), ylim = c(50,290),
        cpoi = 0, clab = 0, grid = FALSE, addax = 0)
old.par <- par(no.readonly = TRUE)
par(mar = c(0.1, 0.1, 0.1, 0.1))
rect(30,0,500,290)
polygon(westafrica$atlantic,col = "lightblue")
points(westafrica$riv.xy, pch = 20, cex = 1.5)
apply(westafrica$lines, 1, function(x) segments(x[1], x[2], x[3],
        x[4], lwd = 1))
apply(westafrica$riv.xy,1, function(x) segments(x[1], x[2], x[3],
        x[4], lwd = 1))
text(c(175,260,460,420), c(275,200,250,100), c("Senegal","Niger",
        "Niger","Volta"))
par(srt = 270)
text(westafrica$riv.xy$x2, westafrica$riv.xy$y2-10,
        westafrica$riv.names, adj = 0, cex = 0.75)
par(old.par)
rm(old.par)

# multivariate analysis
afri.w <- data.frame(t(westafrica$tab))
afri.dist <- dist.binary(afri.w,1)
afri.pco <- dudi.pco(afri.dist, scan = FALSE, nf = 3)
par(mfrow = c(3,1))
barplot(afri.pco$li[,1])
barplot(afri.pco$li[,2])
barplot(afri.pco$li[,3])
```

```

if (require(spdep, quiet = TRUE)){
  #multivariate spatial analysis
  afri.neig <- neig(n.line = 33)
  afri.nb <- neig2nb(afri.neig)
  afri.listw <- nb2listw(afri.nb)
  afri.ms <- multispati(afri.pco, afri.listw, scan = FALSE,
    nfposi = 6, nfneig = 0)
  par(mfrow = c(3,1))
  barplot(afri.ms$li[,1])
  barplot(afri.ms$li[,2])
  barplot(afri.ms$li[,3])

  par(mfrow = c(2,2))
  s.label(afri.ms$li, clab = 0.75, cpoi = 0, neig = afri.neig,
    cneig = 1.5)
  s.value(afri.ms$li, afri.ms$li[,3])
  s.value(afri.ms$li, afri.ms$li[,4])
  s.value(afri.ms$li, afri.ms$li[,5])
  summary(afri.ms)
}

par(mfrow = c(1,1))
plot(hclust(afri.dist,"ward"),h=-0.2)

```

within

Within-Class Analysis

Description

Outputs and graphical representations of the results of a within-class analysis.

Usage

```

## S3 method for class 'within'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'within'
print(x, ...)
## S3 method for class 'witcoi'
plot(x, xax = 1, yax = 2, ...)
## S3 method for class 'witcoi'
print(x, ...)

```

Arguments

x	an object of class <code>within</code> or <code>witcoi</code>
xax	the column index for the x-axis
yax	the column index for the y-axis
...	further arguments passed to or from other methods

Author(s)

Daniel Chessel
 Anne B Dufour <dufour@biomserv.univ-lyon1.fr>
 Stephane Dray <stephane.dray@univ-lyon1.fr>

References

Benzécri, J. P. (1983) Analyse de l'inertie intra-classe par l'analyse d'un tableau de correspondances. *Les Cahiers de l'Analyse des données*, **8**, 351–358.

Dolédec, S. and Chessel, D. (1987) Rythmes saisonniers et composantes stationnelles en milieu aquatique I- Description d'un plan d'observations complet par projection de variables. *Acta Oecologica, Oecologia Generalis*, **8**, 3, 403–426.

See Also

[wca.dudi](#), [wca.coinertia](#)

Examples

```
data(meaudret)
par(mfrow = c(2,2))
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)
s.traject(pca1$li, meaudret$design$site,
  sub = "Principal Component Analysis", csub = 1.5)
wit1 <- wca(pca1, meaudret$design$site, scan = FALSE, nf = 2)
s.traject(wit1$li, meaudret$design$site,
  sub = "Within site Principal Component Analysis", csub = 1.5)
s.corcircle (wit1$as)
par(mfrow = c(1,1))
plot(wit1)
```

withinco inertia	<i>Within-class coinertia analysis</i>
------------------	--

Description

Performs a within-class analysis after a coinertia analysis

Usage

```
withinco inertia(obj, fac, scannf = TRUE, nf = 2)
## S3 method for class 'coinertia'
wca(x, fac, scannf = TRUE, nf = 2, ...)
```

Arguments

obj	a coinertia analysis (object of class coinertia) obtained by the function coinertia
x	a coinertia analysis (object of class coinertia) obtained by the function coinertia
fac	a factor partitioning the rows in classes
scannf	a logical value indicating whether the eigenvalues barplot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
...	further arguments passed to or from other methods

Details

This analysis is equivalent to do a within-class analysis on each initial dudi, and a coinertia analysis on the two within analyses. This function returns additional outputs for the interpretation.

Value

An object of the class `witcoi`. Outputs are described by the `print` function

Note

To avoid conflict names with the `base::within` function, the function `within` is now deprecated and will be removed. To be consistent, the `withincoinertia` function is also deprecated and is replaced by the method `wca.coinertia` of the generic `wca` function.

Author(s)

Stephane Dray <dray@biomserv.univ-lyon1.fr> and Jean Thioulouse <jean.thioulouse@univ-lyon1.fr>

References

Franquet E., Doledec S., and Chessel D. (1995) Using multivariate analyses for separating spatial and temporal effects within species-environment relationships. *Hydrobiologia*, **300**, 425–431.

See Also

[coinertia](#), [within](#)

Examples

```
data(meaudret)
pca1 <- dudi.pca(meaudret$env, scan = FALSE, nf = 4)
pca2 <- dudi.pca(meaudret$spe, scal = FALSE, scan = FALSE, nf = 4)

wit1 <- wca(pca1, meaudret$design$site, scan = FALSE, nf = 2)
wit2 <- wca(pca2, meaudret$design$site, scan = FALSE, nf = 2)
coiw <- coinertia(wit1, wit2, scannf = FALSE)

coi <- coinertia(pca1,pca2,scannf = FALSE,nf=3)
coi.w <- wca(coi,meaudret$design$site, scannf = FALSE)
## coiw and coi.w are equivalent
```

```
plot(coi.w)
```

withinpca	<i>Normed within principal component analysis</i>
-----------	---

Description

Performs a normed within Principal Component Analysis.

Usage

```
withinpca(df, fac, scaling = c("partial", "total"),
          scannf = TRUE, nf = 2)
```

Arguments

df	a data frame with quantitative variables
fac	a factor partitioning the rows of df in classes
scaling	a string of characters as a scaling option : if "partial", the sub-table corresponding to each class is centred and normed. If "total", the sub-table corresponding to each class is centred and the total table is then normed.
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes

Details

This functions implements the 'Bouroche' standardization. In a first step, the original variables are standardized (centred and normed). Then, a second transformation is applied according to the value of the scaling argument. For "partial", variables are standardized in each sub-table (corresponding to each level of the factor). Hence, variables have null mean and unit variance in each sub-table. For "total", variables are centred in each sub-table and then normed globally. Hence, variables have a null mean in each sub-table and a global variance equal to one.

Value

returns a list of the sub-class within of class dudi. See [within](#)

Author(s)

Daniel Chessel
Anne B Dufour <dufour@biomserv.univ-lyon1.fr>

References

Bouroche, J. M. (1975) *Analyse des données ternaires: la double analyse en composantes principales*. Thèse de 3ème cycle, Université de Paris VI.

Examples

```

data(meaudret)
wit1 <- withinpca(meaudret$env, meaudret$design$season,
  scannf = FALSE, scaling = "partial")
kta1 <- ktab.within(wit1, colnames = rep(c("S1", "S2", "S3", "S4", "S5"), 4))
unclass(kta1)
# See pta
plot(wit1)

```

witwit.coa

Internal Correspondence Analysis

Description

witwit.coa performs an Internal Correspondence Analysis. witwitsepan gives the computation and the barplot of the eigenvalues for each separated analysis in an Internal Correspondence Analysis.

Usage

```

witwit.coa(dudi, row.blocks, col.blocks, scannf = TRUE, nf = 2)
## S3 method for class 'witwit'
summary(object, ...)
witwitsepan(ww, mfrow = NULL, csub = 2, plot = TRUE)

```

Arguments

dudi	an object of class coa
row.blocks	a numeric vector indicating the row numbers for each block of rows
col.blocks	a numeric vector indicating the column numbers for each block of columns
scannf	a logical value indicating whether the eigenvalues bar plot should be displayed
nf	if scannf FALSE, an integer indicating the number of kept axes
object	an object of class witwit
...	further arguments passed to or from other methods
ww	an object of class witwit
mfrow	a vector of the form "c(nr,nc)", otherwise computed by a special own function 'n2mfrow'
csub	a character size for the sub-titles, used with par("cex")*csub
plot	if FALSE, numeric results are returned

Value

returns a list of class `witwit`, `coa` and `dudi` (see [as.dudi](#)) containing

<code>rbvar</code>	a data frame with the within variances of the rows of the factorial coordinates
<code>lbw</code>	a data frame with the marginal weighting of the row classes
<code>cvar</code>	a data frame with the within variances of the columns of the factorial coordinates
<code>cbw</code>	a data frame with the marginal weighting of the column classes

Author(s)

Daniel Chessel Anne B Dufour <dufour@biomserv.univ-lyon1.fr> Correction by Campo Elías PARDO <cepardot@cable.net.co>

References

Cazes, P., Chessel, D. and Dolédec, S. (1988) L'analyse des correspondances internes d'un tableau partitionné : son usage en hydrobiologie. *Revue de Statistique Appliquée*, **36**, 39–54.

Examples

```
data(ardeche)
coa1 <- dudi.coa(ardeche$tab, scann = FALSE, nf = 4)
ww <- witwit.coa(coa1, ardeche$row.blocks, ardeche$col.blocks, scann = FALSE)
ww
s.class(ww$co, ardeche$sta.fac, clab = 1.5, cell = 0, axesell = FALSE)
s.label(ww$co, add.p = TRUE, clab = 0.75)
summary(ww)

witwitsepan(ww, c(4,6))
```

woangers

Plant assemblages in woodlands of the conurbation of Angers (France)

Description

This data set gives the presence of plant species in relevés of woodlands in the conurbation of Angers; and their biological traits.

Usage

```
data(woangers)
```

Format

woangers is a list of 2 components.

1. flo: is a data frame that contains the presence/absence of species in each sample site. In the codes for the sample sites (first column of the data frame), the first three letters provide the code of the woodland and the numbers represent the 5 quadrats sampled in each site. Codes for the woodlands are based on either their local name when they have one or on the name of the nearest locality.
2. traits: is a data frame that contains the values of the 13 functional traits considered in the paper. One trait can be encoded by several columns. The codes are as follows:
 - Column 1: Species names;
 - Column 2: li, nominal variable that indicates the presence (y) or absence (n) of ligneous structures;
 - Column 3: pr, nominal variable that indicates the presence (y) or absence (n) of prickly structures;
 - Column 4: fo, circular variable that indicates the month when the flowering period starts (from 1 January to 9 September);
 - Column 5: he, ordinal variable that indicates the maximum height of the leaf canopy;
 - Column 6: ae, ordinal variable that indicates the degree of aerial vegetative multiplication;
 - Column 7: un, ordinal variable that indicates the degree of underground vegetative multiplication;
 - Column 8: lp, nominal variable that represents the leaf position by 3 levels (ros = rosette, semiros = semi-rosette and leafy = leafy stem);
 - Column 9: le, nominal variable that represents the mode of leaf persistence by 5 levels (seasaes = seasonal aestival, seashib = seasonal hibernal, seasver = seasonal vernal, everalw = always evergreen, everparti = partially evergreen);
 - Columns 10, 11 and 12: fuzzy variable that describes the modes of pollination with 3 levels (auto = autopollination, insects = pollination by insects, wind = pollination by wind); this fuzzy variable is expressed as proportions, i.e. for each row, the sum of the three columns equals 1;
 - Columns 13, 14 and 15: fuzzy variable that describes the life cycle with 3 levels (annual, monocarpic and polycarpic); this fuzzy variable is expressed as proportions, i.e. for each row, the sum of the three column equals 1;
 - Columns 16 to 20: fuzzy variable that describes the modes of dispersion with 5 levels (elaio = dispersion by ants, endozoo = injection by animals, epizoo = external transport by animals, wind = transport by wind, unsp = unspecialized transport); this fuzzy variable is expressed as proportions, i.e. for each row, the sum of the three columns equals 1;
 - Column 21: lo, quantitative variable that provides the seed bank longevity index;
 - Column 22: lf, quantitative variable that provides the length of the flowering period.

Source

Pavoine, S., Vallet, J., Dufour, A.-B., Gachet, S. and Daniel, H. (2009) On the challenge of treating various types of variables: Application for improving the measurement of functional diversity. *Oikos*, **118**, 391–402.

Examples

```

# Loading the data
data(woangers)

# Preparing of the traits
traits <- woangers$traits
# Nominal variables 'li', 'pr', 'lp' and 'le'
# (see table 1 in the main text for the codes of the variables)
tabN <- traits[, c(1:2, 7, 8)]
# Circular variable 'fo'
tabC <- traits[3]
tabCp <- prep.circular(tabC, 1, 12)
# The levels of the variable lie between 1 (January) and 12 (December).
# Ordinal variables 'he', 'ae' and 'un'
tabO <- traits[, 4:6]
# Fuzzy variables 'mp', 'pe' and 'di'
tabF <- traits[, 9:19]
tabFp <- prep.fuzzy(tabF, c(3, 3, 5), labels = c("mp", "pe", "di"))
# 'mp' has 3 levels, 'pe' has 3 levels and 'di' has 5 levels.
# Quantitative variables 'lo' and 'lf'
tabQ <- traits[, 20:21]

# Combining the traits
ktab1 <- ktab.list.df(list(tabN, tabCp, tabO, tabFp, tabQ))
## Not run:
# Calculating the distances for all traits combined
distrat <- dist.ktab(ktab1, c("N", "C", "O", "F", "Q"))
is.euclid(distrat)

# Calculating the contribution of each trait in the combined distances
contrib <- kdist.cor(ktab1, type = c("N", "C", "O", "F", "Q"))
contrib
dotchart(sort(contrib$glocor), labels = rownames(contrib$glocor)[order(contrib$glocor[, 1])])

## End(Not run)

```

worksurv

French Worker Survey (1970)

Description

The worksurv data frame gives 319 response items and 4 questions providing from a French Worker Survey.

Usage

```
data(worksurv)
```

Format

This data frame contains the following columns:

1. pro: Professional elections. In professional elections in your firm, would you rather vote for a list supported by?
 - CGT
 - CFDT
 - FO
 - CFTC
 - Auton Autonomous
 - Abst
 - Nonaffi Not affiliated
 - NR No response
2. una: Union affiliation. At the present time, are you affiliated to a Union, and in the affirmative, which one?
 - CGT
 - CFDT
 - FO
 - CFTC
 - Auton Autonomous
 - CGC
 - Notaffi Not affiliated
 - NR No response
3. pre: Presidential election. On the last presidential election (1969), can you tell me the candidate for whom you havevoted?
 - Duclos
 - Deferre
 - Krivine
 - Rocard
 - Poher
 - Ducatel
 - Pompidou
 - NRabs No response, abstention
4. pol: political sympathy. Which political party do you feel closest to, as a rule ?
 - Communist (PCF)
 - Socialist (SFIO+PSU+FGDS)
 - Left (Party of workers,...)
 - Center MRP+RAD.
 - RI
 - Right INDEP.+CNI
 - Gaullist UNR
 - NR No response

Details

The data frame `worksurv` has the attribute `'counts'` giving the number of responses for each item.

Source

Rouanet, H. and Le Roux, B. (1993) *Analyse des données multidimensionnelles*. Dunod, Paris.

References

Le Roux, B. and Rouanet, H. (1997) Interpreting axes in multiple correspondence analysis: method of the contributions of points and deviation. Pages 197-220 in B. J. and M. Greenacre, editors. *Visualization of categorical data*, Academic Press, London.

Examples

```
data(worksurv)
acm1 <- dudi.acm(worksurv, row.w = attr(worksurv, "counts"),
  scan = FALSE)
par(mfrow = c(2,2))
apply(worksurv, 2, function(x) s.class(acm1$li, factor(x),
  attr(worksurv, 'counts')))
```

yanomama

Distance Matrices

Description

This data set gives 3 matrices about geographical, genetic and anthropometric distances.

Usage

```
data(yanomama)
```

Format

`yanomama` is a list of 3 components:

geo is a matrix of 19-19 geographical distances

gen is a matrix of 19-19 SFA (genetic) distances

ant is a matrix of 19-19 anthropometric distances

Source

Spielman, R.S. (1973) Differences among Yanomama Indian villages: do the patterns of allele frequencies, anthropometrics and map locations correspond? *American Journal of Physical Anthropology*, **39**, 461–480.

References

Table 7.2 Distance matrices for 19 villages of Yanomama Indians. All distances are as given by Spielman (1973), multiplied by 100 for convenience in: Manly, B.F.J. (1991) *Randomization and Monte Carlo methods in biology* Chapman and Hall, London, 1–281.

Examples

```
data(yanomama)
gen <- quasieucld(as.dist(yanomama$gen)) # depends of mva
ant <- quasieucld(as.dist(yanomama$ant)) # depends of mva
par(mfrow = c(2,2))
plot(gen, ant)
t1 <- mantel.randtest(gen, ant, 99);
plot(t1, main = "gen-ant-mantel") ; print(t1)
t1 <- procuste.rtest(pcoscaled(gen), pcoscaled(ant), 99)
plot(t1, main = "gen-ant-procuste") ; print(t1)
t1 <- RV.rtest(pcoscaled(gen), pcoscaled(ant), 99)
plot(t1, main = "gen-ant-RV") ; print(t1)
```

zealand

Road distances in New-Zealand

Description

This data set gives the road distances between 13 towns in New-Zealand.

Usage

```
data(zealand)
```

Format

zealand is a list of 3 components:

road is a data frame with 13 rows (New Zealand towns) and 13 columns (New Zealand towns) containing the road distances between these towns.

xy is a data frame containing the coordinates of the 13 towns.

neig is a object of class 'neig', a neighbour graph to visualize the map shape.

Source

Manly, B.F. (1994) *Multivariate Statistical Methods. A primer.*, Second edition, Chapman and Hall, London, 1–215, page 172.

Examples

```
data(zealand)

d0 = as.dist(as.matrix(zealand$road))
d1 = cailliez (d0)
d2 = lingoies(d0)
s.label(zealand$xy,lab=as.character(1:13),neig=zealand$neig)
par(mfrow = c(2,2))
  s.label(cmdscale(dist(zealand$xy)),lab = as.character(1:13),
    neig = zealand$neig, sub = "Distance canonique", csub = 2)
  s.label(cmdscale(d0), lab = as.character(1:13), neig = zealand$neig,
    sub = "Distance routiere", csub = 2)
  s.label(cmdscale(d1), lab = as.character(1:13), neig = zealand$neig,
    sub = "Distance routiere / Cailliez", csub = 2)
  s.label(cmdscale(d2), lab = as.character(1:13), neig = zealand$neig,
    sub = "Distance routiere / Lingoies", csub = 2)
```


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